

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

DIVISIONAL APPLICATION

In re Application of Petkovich, P. Martin, *et al.*  
Title: Retinoid Metabolizing Protein  
Atty's Docket No.: 57600/00035

**Box Patent Application**

The Commissioner of Patents and Trademarks  
Washington, D.C. 20331  
U.S.A.

Dear Sir:

This is a request for filing a divisional application.

We enclose the following:

1. Copy of prior application, Serial No. 08/882,164 filed June 25, 1997:

	No. of pages
Cover sheet:	1
Description:	69
Claims	4
Drawings	20
<u>Abstract:</u>	<u>1</u>
Total:	95

2. Copy of signed Declaration and Power of Attorney from prior application.
3. Copy of PTO letter dated August 29, 2000 indicating that Applicant's petition under 37 C.F.R. § 1.137(b), filed May 3, 2000, to revive U.S.S.N. 08/882,164 has been granted.
4. Return Receipt Post Card.

The filing fee is not enclosed. Please charge any fee which may be required in order that a filing date be granted on the enclosed application to Account No. 02-2553.

**Prior Application Information:**

U.S.S.N. 08/882,164 filed June 25, 1997  
International patent application No. PCT/CA 97/00440, filed June 23, 1997  
U.S.S.N. 08/724,466, filed October 1, 1996  
U.S.S.N. 08/667,546, filed June 21, 1996

Please amend the specification so that the first sentence reads:

- This application is a divisional application of United States Patent Application Serial No. 08/882,164, filed June 25, 1997, issued as United States Patent No. \_\_\_\_\_, on \_\_\_\_\_, which application is a continuation-in-part application of international patent application No. PCT/CA 97/00440, filed under the same title on June 23, 1997 through the Canadian Intellectual Property Office as Receiving Office and designating the United States; U.S. patent application Serial No. 08/724,466, filed October 1, 1996, issued as United States Patent No. 6,063,606 on May 16, 2000, which application is a continuation-in-part application of U.S. patent application Serial No. 08/667,546, filed June 21, 1996, the specifications of which applications are incorporated herein by reference, and which latter application is now abandoned. --

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Yours very truly,



John C. Hunt  
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September 22, 2000

Date

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UNITED STATES OF AMERICA

TO ALL WHOM IT MAY CONCERN:

BE IT KNOWN THAT WE,

P. Martin Petkovich, of 875 Everitt Avenue, Kingston, Ontario K7M 4R1 Canada, Citizen of Canada; Jay A. White, of 913 Purcell Crescent, Kingston, Ontario K7P 1C1 Canada, Citizen of Canada; Barbara R. Beckett, of 112 Westmorland Road, Kingston, Ontario K7M 1J7 Canada, Citizen of Canada; and Glenville Jones, of 66 Inverness Crescent, Kingston, Ontario K7M 6N7 Canada, Citizen of Canada

have invented:

**RETINOID METABOLIZING PROTEIN**

of which the following is a specification.

## Retinoid Metabolizing Protein

This application is a continuation-in-part application of international patent application No. \_\_\_\_\_, filed under the same title on June 23, 1997 through the Canadian Intellectual Property Office as Receiving Office and designating the United States; U.S. patent application Serial No. 08/724,466, filed October 1, 1996, which application is a continuation-in-part application of U.S. patent application Serial No. 08/667,546, filed June 21, 1996, the specifications of which applications are incorporated herein by reference, and which latter application is now abandoned.

### Background of the Invention

Vitamin A metabolism gives rise to several active forms of retinoic acid (RA) which are involved in regulating gene expression during development, regeneration, and in the growth and differentiation of epithelial tissues [Maden, 1992; Chambon, 1995; Mangelsdorf, 1995; Gudas, 1994; Lotan, 1995; Morriss-Kay, 1996]; has been linked apoptosis, or programmed cell death in a number of cell types; and to have anticarcinogenic and antitumoral properties [Lotan, 1996].

Early studies of retinol deficiency indicated a correlation between vitamin A depletion and a higher incidence of cancer and increased susceptibility to chemical carcinogenesis [Chytil, 1984]. Several animal models have been used to demonstrate the effectiveness of retinoids in suppressing carcinogenesis in a variety of tissues including skin, mammary epithelia, oral cavity, aerodigestive tract, liver, bladder and prostate [Moon, 1994]. These studies have led to the preventative use of retinoids to treat premalignant lesions including actinic keratosis and oral leukoplakia, as well as in the prevention of secondary tumours of the head and neck and the recurrence of non-small cell lung carcinomas, and basal cell carcinomas [Hong, 1994; Lippman, 1995]. RA itself has been found to be useful therapeutically, notably in the treatment of cancers, including acute promyelocytic leukemia (APL), tumors of the head and neck, and skin cancer, as well as in the treatment of skin disorders such as the premalignancy associated actinic keratoses, acne, psoriasis and ichthyosis. There is evidence that the effectiveness of RA as an anti-tumour agent is at least partially due to induction of cellular differentiation and/or inhibition of proliferation [Lotan, 1996]. Studies over the past several years indicate that a high proportion of patients with acute promyelocytic leukemia (APL) achieve complete remission after a short period of treatment with all-*trans* RA. Unfortunately, this high rate of remission is in most cases brief. Following relapse, patients are clinically resistant to further treatment with RA [Warrell, 1994; Warrell, *et al.*, 1994; Chomienne, 1996; Muindi, 1992]. The nature of this resistance is unknown. Interestingly, leukemic cells taken from patients exhibiting clinical resistance to RA have been shown to be sensitive to the differentiating action of RA when grown *in vitro* (Muindi, 1992; Muindi, 1994). This suggests that pharmacokinetic mechanisms may account for the acquired resistance to RA. This possibility is supported by studies showing that peak plasma concentrations of RA were much higher in patients after initial administration than in patients treated following relapse. This decrease in peak plasma RA concentration was accompanied by a 10-fold increase in urinary 4-oxo-retinoic acid concentration. In addition, ketoconazole, a broad spectrum inhibitor of cytochrome P450 function was shown to modulate RA

pharmacokinetics in vivo [Muindi, 1992; Muindi, 1994]. It is therefore likely that RA increases the rate of its own metabolism which in turn results in the inability to sustain effective therapeutic doses of RA. Therapeutic administration of RA can result in a variety of undesirable side effects and it is therefore important to establish and maintain the minimal requisite doses of RA in treatment. For example, RA treatments during pregnancy can lead to severe teratogenic effects on the fetus. Adverse reactions to RA treatment also include headache, nausea, chelitis, facial dermatitis, conjunctivitis, and dryness of nasal mucosa. Prolonged exposure to RA can cause major elevations in serum triglycerides and can lead to severe abnormalities of liver function, including hepatomegaly, cirrhosis and portal hypertension.

10 RA metabolism may also account for the lack of response of certain tumors to RA treatment. For example, recent studies have shown that cytochrome P450 inhibitors that block RA metabolism, resulting in increased tissue levels of RA, may be useful therapeutic agents in the treatment of prostate cancer [Wouters, 1992; De Coster, 1996]. Thus RA metabolizing cytochrome P450s may be useful targets for the treatment of a number of  
15 different types of cancer.

The classical view of vitamin A metabolism holds that all *trans*-RA, the most active metabolite is derived from conversion of retinol to retinaldehyde to RA through two oxidation steps and that RA is further metabolized to the polar derivatives 4-OH RA and 4-oxo RA [Blaner, 1994; Napoli, 1995; Formelli, 1996; Napoli, 1996]. It is unknown whether the 4-  
20 oxo- and 4-OH- metabolites are simply intermediates in the RA catabolic pathway or whether they can also have specific activities which differ from those of all-*trans* RA and 9-*cis* RA. Pijnappel et al. [Pijnappel, 1993] have shown that, in *Xenopus*, 4-oxo-RA can efficiently modulate positional specification in early embryos and exhibits a more potent ability to regulate Hoxb-9 and Hoxb-4 gene expression than all-*trans* RA. 4-oxo-RA has been found to bind to  
25 retinoic acid receptor- $\beta$  (RAR- $\beta$ ) with affinity comparable to all-*trans* RA [Pijnappel, 1993] but poorly to RAR- $\gamma$  [Reddy, 1992], suggesting that this metabolite exhibits some receptor selectivity. 4-oxo-RA also binds to cellular retinoic acid binding protein (CRABP) but with an affinity slightly lower than that of all-*trans* RA [Fiorella, 1993]. Takatsuka et al. [Takatsuka, 1996] have shown that growth inhibitory effects of RA correlate with RA metabolic activity but it  
30 is unknown whether there is a causal relationship between production of RA metabolites and growth inhibition. The asymmetric distribution of these metabolites in developing embryos suggests that they may be preferentially sequestered or generated by tissue specific isomerases [Creech Kraft, 1994]. The normal balance of these metabolites is dependent upon rate of formation from metabolic precursors, retinol and retinaldehyde [Leo, 1989], and rate of  
35 catabolism. Little is presently known about the enzymes involved in this metabolic scheme, in particular the catabolism of RA.

The catabolism of RA is thought to be initiated by hydroxylation either at the C4-, or C18-position of the  $\beta$ -ionone ring of RA [Napoli, 1996]. The C4- hydroxylation step is mediated by cytochrome P450 activity, as judged by the ability of broad spectrum P450 inhibitors such as  
40 ketocoazole and liarazole to block 4-hydroxylation [Williams, 1987, Van Wauwe, 1988; Van

Wauwe, 1990, Van Wauwe, 1992, Wouters, 1992]. In certain tissues, including testis, skin and lung and in numerous cell lines, such as NIH3T3 fibroblasts, HL 60 myelomonocytic leukemic cells, F9 and P19 murine embryonal carcinoma cell lines and MCF7, RA metabolism can be induced by RA pretreatment [Frolik, 1979, Roberts, 1979a and b; Duell, 1992; Wouters, 1992].

- 5 Studies involving targetted disruption of RAR genes in F9 cells suggest that RAR- $\alpha$  and RAR- $\gamma$  isoforms may play a role in regulating the enzymes responsible for this increased metabolism [Boylan, 1995].

The glucuronidation of RA is a significant metabolic step in the inactivation of RA [Blaner, 1994; Formelli, 1996]. The elimination of RA may require oxidation to 4-oxo, followed by conjugation to form the 4-oxo all-trans RA glucuronide. This is supported by studies in both primates and humans showing that the 4-oxo RA glucuronide is the only retinoid conjugate found in urine [Muindi, 1992; Muindi, 1994]. The fact that following RA therapy, 4-oxo RA is not present or barely detectable in serum, suggests that oxidation may be the rate limiting step in this process.

- 15 It has recently been shown that 4-oxoretinol (4-oxo-ROL) can have greater biological activity than retinol. The 4-oxo-ROL is inducible by RA in F9 and P19 mouse teratocarcinoma cells [Blumberg *et al.*, 1995; Achkar *et al.*, 1996].

It is known that zebrafish fins regenerate through an RA sensitive process which utilizes many gene regulatory pathways involved in early vertebrate development [White, 1994; Akimenko, 1995a & b].

- 20 As far as the inventors are aware, cytochrome P450s involved in the metabolism of RA in extrahepatic tissues remain uncharacterized at the molecular level.

## SUMMARY OF THE INVENTION

The present inventors are the first to identify, clone and sequence a gene (cDNA) encoding a retinoic acid-inducible, retinoic acid-metabolizing protein, including a cDNA which is RA-inducible in humans. The protein has been found to be expressed in epithelia.

A cDNA has been isolated from zebra fish and sequenced. A protein encoded by the cDNA has been expressed and shown to have the ability to hydroxylate retinoic acid at the 4 position of the  $\beta$ -ionone ring of retinoic acid. The protein has been found to be inducible in epithelial cells exposed to retinoic acid.

- 30 A human cDNA encoding a protein with similar functionality has also been isolated and sequenced.

A cDNA has also been isolated from mouse and sequenced.

Homology between sequences from the three species, be they nucleic acids encoding the protein, or the amino acid sequences of the proteins, has been found to be relatively high and all three proteins contain a heme-binding motif characteristic of the group of proteins known as cytochrome P450s. The overall homology between the amino acid sequences of these newly obtained proteins and known cytochrome P450s is less than 30%. Notwithstanding this relatively low overall homology, a higher degree of homology has been

observed in the heme binding region for certain other P450s. For example, homology between the approximately 20 amino acids defining respective heme binding regions of the new zebrafish protein and CYP4503A12 is about 50% and between the new zebrafish protein and hCYTFAOH is 65%. The homology between the heme binding region itself of a protein of the present invention and another P450 could well be 70%, 75%, 80%, 85%, 90%, 95% or even 100%.

A first aspect of the present invention is thus a purified protein having the ability to oxidize a retinoid, and having an amino acid sequence which is at least about 30% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or identified as SEQ ID NO:32, or a functionally equivalent homolog thereof. The amino acid sequence identified as SEQ ID NO:2 is of the protein, termed here "zP450RAI", obtained from zebrafish. The amino acid sequence of the human protein is identified as SEQ ID NO:4 and the protein is referred to herein as "hP450RAI". The sequence of the mouse protein is identified as SEQ ID NO:32 and the protein is referred to herein as mP450RAI.

Such a protein which is at least about 35% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 (zebrafish) or identified as SEQ ID NO:4 (human) or identified as SEQ ID NO:32 (mouse), or a functionally equivalent homolog thereof, also forms part of the invention disclosed herein. Likewise, the degree of sequence conservation of a protein could be 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95% or of course 100% of either SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:32 or a functionally equivalent homolog thereof, variants being possible so long as the ability of the native protein to oxidize a retinoid is retained. Also within the scope of the invention is any such protein which has the ability to hydroxylate retinoic acid at the 4 position of the  $\beta$ -ionone ring. Of course, conservatively substituted variants of proteins disclosed are within the scope of the present invention.

A retinoid oxidized by a protein of the present invention may be a retinoic acid or a retinol and the protein may have the ability to oxidize the carbon occupying the 4-position of the  $\beta$ -ionone ring of the retinoid. In particular, all-*trans* retinoids may be metabolized by proteins of the present invention.

In the context of this specification, the term "conserved" describes similarity between sequences. The degree of conservation between two sequences can be determined by optimally aligning the sequences for comparison, as is commonly known in the art, and comparing a position in the first sequence with a corresponding position in the second sequence. When the compared positions are occupied by the same nucleotide or amino acid, as the case may be, the two sequences are conserved at that position. The degree of conservation between two sequences is often expressed, as it is here, as a percentage representing the ratio of the number of matching positions in the two sequences to the total number of positions compared.

The generic term "retinoids" means a group of compounds which includes retinoic acid, vitamin A (retinol) and a series of natural and synthetic derivatives that can exert

profound effects on development and differentiation in a wide variety of systems. For purposes of this disclosure "retinoid" is also intended to encompass an equivalent thereof having the same functional characteristics which may be produced, for example, by computational chemistry.

5 In another aspect, the present invention is an isolated nucleic acid molecule encoding a protein of the present invention.

The present invention thus includes an isolated nucleic acid molecule encoding a protein having an amino acid sequence which is at least about 30% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or  
10 identified as SEQ ID NO:32, or a functionally equivalent homolog thereof, for example, or a nucleic acid strand capable of hybridizing with the nucleic acid molecule under stringent hybridization conditions. Of course, the degree of conservation of the protein which the nucleic acid encodes can be higher, that is, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95% or more.

15 Particularly, the invention is an isolated nucleic acid molecule encoding a protein having the ability to oxidize a retinoid at the carbon occupying the 4-position of the  $\beta$ -ionone ring of the retinoid ring, and more particularly, having all-*trans* retinoic acid 4-hydroxylase activity. For the purposes of this invention, the term "isolated" refers to a nucleic acid that is substantially free of other cellular material or culture medium when produced by  
20 recombinant DNA techniques, or chemical precursors or other chemicals when produced by chemical synthesis.

Cellular expression of preferred proteins of the present invention, preferred embodiments being described in more detail below, can for certain types of cells be induced by exposure of the cells to a retinoid, particularly, retinoic acid. A protein of the present invention,  
25 when described as being a "retinoic acid inducible protein", is a protein normally encoded by DNA of a cell and whose expression by that cell can be induced by exposure of the cell to retinoic acid. It will be appreciated that not every cell, even if it contains DNA encoding such a protein, possesses all the attributes necessary to express the protein on exposure to RA.

Nucleotide sequences possessing promoter activity associated with the  
30 proteins have also been isolated and identified by the inventors. The human nucleotide sequence having promoter activity is identified as SEQ ID NO:33. The murine nucleotide sequence having promoter activity is identified as SEQ ID NO:34. The zebrafish nucleotide sequence having promoter activity is identified as SEQ ID NO:35.

In fact, the invention includes genomic sequences for the human (SEQ ID  
35 Nos:36 and 37) and mouse (SEQ ID NO:38).

In another respect, the present invention is thus a microbial cell containing and expressing heterologous DNA encoding a retinoic acid inducible protein having all-*trans* retinoic acid 4-hydroxylase activity.

The sequence of a nucleic acid molecule of the present invention can  
40 correspond to a part of a human genome or of a fish genome or of a mouse genome, or vary



therefrom due to the degeneracy of the genetic code. More particularly, a nucleic acid molecule of the present invention can be a DNA molecule having the sequence identified as SEQ ID NO:3 (zP450RAI) or SEQ ID NO:5 (hP450RAI), or SEQ ID NO:31 (mP450RAI), or the sequence can be one which varies from one of these sequences due to the degeneracy of the genetic code, or it can be a nucleic acid strand capable of hybridizing with at least one of these nucleic acid molecules under high or low stringency hybridization conditions.

"Stringent hybridization conditions" takes on its common meaning to a person skilled in the art here. Appropriate stringency conditions which promote nucleic acid hybridization, for example, 6x sodium chloride/sodium citrate (SSC) at about 45°C are known to those skilled in the art. The following examples are found in Current Protocols in Molecular Biology, John Wiley & Sons, NY (1989), 6.3.1-6.3.6: For 50 ml of a first suitable hybridization solution, mix together 24 ml formamide, 12 ml 20x SSC, 0.5 ml 2 M Tris-HCl pH 7.6, 0.5 ml 100x Denhardt's solution, 2.5 ml deionized H<sub>2</sub>O, 10 ml 50% dextran sulfate, and 0.5 ml 10% SDS. A second suitable hybridization solution can be 1% crystalline BSA (fraction V), 1 mM EDTA, 0.5 M Na<sub>2</sub>HPO<sub>4</sub> pH 7.2, 7% SDS. The salt concentration in the wash step can be selected from a low stringency of about 2x SSC at 50°C to a high stringency of about 0.2x SSC at 50°C. Both of these wash solutions may contain 0.1% SDS. In addition, the temperature in the wash step can be increased from low stringency conditions at room temperature, about 22°C, to high stringency conditions, at about 65°C. The cited reference gives more detail, but appropriate wash stringency depends on degree of homology and length of probe. If homology is 100%, a high temperature (65°C to 75°C) may be used. If homology is low, lower wash temperatures must be used. However, if the probe is very short (<100bp), lower temperatures must be used even with 100% homology. In general, one starts washing at low temperatures (37°C to 40°C), and raises the temperature by 3-5°C intervals until background is low enough not to be a major factor in autoradiography.

Another aspect of this invention is isolated mRNA transcribed from DNA having a sequence encoding a protein of the present invention.

In another aspect, the present invention is isolated DNA having a sequence according to a nucleotide sequence described above operatively linked in a recombinant cloning vector. In the context of this invention, the two-part term "operatively linked" means both that the regulatory sequence contains sufficient element(s) to allow expression of the nucleic acid in question and that the nucleic acid is linked to the regulatory sequence appropriately. For example, the nucleic acid of the invention is in the appropriate orientation and in phase with an initiation codon. The present invention thus includes a stably transfected cell line which expresses a protein having the ability to hydroxylate retinoic acid at the 4 position of the  $\beta$ -ionone ring of retinoic acid. The invention includes a culture of cells transformed with a recombinant DNA molecule having a nucleic acid sequence which encodes a protein having the ability to hydroxylate retinoic acid at the 4 position of the  $\beta$ -ionone ring of retinoic acid.

Another aspect of the present invention is a host cell that has been engineered genetically to produce a protein of the invention described above, the cell having incorporated

expressibly therein heterologous DNA encoding said protein. The cell may be selected such that production of the protein is inducible by exposing the cell to a retinoid, preferably, retinoic acid. The cell can be eukaryotic.

- The present invention also includes a process for producing an above-
- 5 described protein of the invention. Such a process includes:
- preparing a DNA fragment containing a nucleotide sequence which encodes the protein;
  - incorporating the DNA fragment into an expression vector to obtain a recombinant DNA molecule which contains the DNA fragment and is capable of undergoing replication;
  - transforming a host cell with the recombinant DNA molecule to produce a transformant
  - 10 which can express the protein;
  - culturing the transformant to produce the protein; and
  - recovering the protein from resulting cultured mixture.

- The present invention includes an antibody to a protein of the invention. Here, the term "antibody" is intended to include a Fab fragment and it can be a monoclonal antibody.
- 15 The antibody can be specifically raised to the amino acid sequence identified as SEQ ID NO:4, i.e., hP450RAI.

- The present invention includes a purified protein for use in metabolizing retinoic acid in an organism or cell in need of such metabolizing. Likewise, the invention includes a method for metabolizing retinoic acid in an organism or cell in need of retinoic acid
- 20 metabolizing wherein the method includes administering a protein of the invention as described above.

- The invention includes a method for inhibiting retinoic acid hydroxylation in an organism in need of such inhibition, comprising introducing into cells of the organism an effective amount of an antisense RNA or oligonucleotide substantially complementary to at
- 25 least a portion of the sequence identified as SEQ ID NO:5. The organism can be human and/or the organism can be in need of treatment against a cancerous disease or a disease selected from the group consisting of cancer, actinic keratosis, oral leukoplakia, a secondary tumor of the head and/or neck, a non-small cell lung carcinoma, a basal cell carcinoma, acute promyelocytic leukemia, skin cancer, and a premalignancy associated actinic keratosis, acne,
- 30 psoriasis and/or ichthyosis. Such a method can include use of at least one delivery vehicle or technique selected from the set of viral vectors, microinjection, electroporation, coprecipitation, liposomes, aerosol delivery and lavage. The portion of the sequence may be 5 bases in length, between 5 and 50 bases in length, 5 and 30 bases in length between 10 and 20 bases in length, or another suitable length may be found. The organism may be a human patient and
- 35 the method can include treating the patient against a cancerous disease.

- The invention also includes a method of inhibiting retinoic acid hydroxylation in an organism in need of such inhibition by administering to the organism an effective amount of an antibody, such antibodies being described above. A particularly useful antibody for the treatment of a human would be an antibody to the protein having the amino acid sequence
- 40 identified as SEQ ID NO:4, or a portion thereof. It would be advantageous to adapt such an

antibody for administration to a human by "humanizing" the antibody, as is understood by those skilled in the art [Hozumi, 1993].

The invention includes a method for producing a desired protein, comprising providing a cell which can produce an endogenous protein in response to exposure to a  
5 retinoid; incorporating into DNA of the cell a DNA sequence encoding the desired protein at or near a site which is normally occupied by a DNA sequence encoding the endogenous protein; and exposing the cell to the retinoid so as to induce production of the desired protein.

In another embodiment, the present invention is a kit for determining the presence of a protein having the ability to oxidize a retinoid, and having an amino acid  
10 sequence which is at least about 30% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:32, or more likely for determining the presence of a protein (or peptide fragment thereof) having an amino acid sequence identified as SEQ ID NO:4, the human protein. The kit includes an antibody to the protein linked to a  
15 reporter system wherein the reporter system produces a detectable response when a predetermined amount of the protein and the antibody are bound together.

In another aspect, the present invention is a kit for determining the presence of a nucleic acid encoding a protein of invention, or having the sequence identified as SEQ ID  
20 NO:3 or SEQ ID NO:5 or SEQ ID NO:31, or which varies from the sequence due to the degeneracy of the genetic code, or a nucleic acid strand capable of hybridizing with at least one said nucleic acid under stringent hybridization conditions. The kit includes a nucleic acid molecule capable of hybridizing with at least a portion of a said nucleic acid or nucleic acid strand under stringent conditions in which the nucleic acid molecule is linked to a reporter  
25 system wherein the reporter system produces a detectable response when a predetermined amount of the nucleic acid or nucleic acid strand and nucleic acid molecule are hybridized with each other. The molecule can be 5 bases in length or longer; between 5 and 50 bases in length, between 5 and 30 or 40 bases in length, or between 10 and 20 bases in length. Of course it might be possible to find a more suitable base length.

The present invention includes an isolated DNA molecule having a nucleotide sequence selected from the group consisting of:

- 30 (a) SEQ ID NO:33;  
(b) SEQ ID NO:34;  
(c) SEQ ID NO:35; and  
(d) a fragment of (a), (b) or (c),

wherein the DNA molecule possesses promoter activity.

35 In a more particular aspect, the invention includes a DNA molecule of one of the indicated sequences in which the DNA molecule includes the sequence TGAAC(N)<sub>x</sub>TGAAC, wherein x has a value of up to 5. This sequence is conserved in all three of the nucleotide sequences identified as having promoter activity, particularly where x is 5. Even more particularly, the invention includes such a DNA molecule in which the sequence  
40 TCTGASSAAGKTAAC occurs downstream from the sequence TGAAC(N)<sub>x</sub>TGAAC. Even

more particularly, the sequence includes the sequence AATT between the sequence TGAAGT(N)<sub>x</sub>TGAAGT and the sequence TCTGASSAAGKTAAC, the AATT having been found immediately downstream of the sequence TGAAGT(N)<sub>x</sub>TGAAGT. It has been observed that there can be up to six nucleotides between the sequence TGAAGT(N)<sub>x</sub>TGAAGT and the sequence TCTGASSAAGKTAAC. There can also be the sequence CAATTAAAGA upstream of the sequence TGAAGT(N)<sub>x</sub>TGAAGT. In a particular aspect, the nucleotide sequence having promoter activity includes the sequence CAATTAAAGATGAAGTTTGGGTGAAGTAAATT and the sequence TATAA. Particularly, the sequence TATAA is downstream of the sequence TGAAGT(N)<sub>x</sub>TGAAGT, and more particularly, the sequence TATAA is downstream of the sequence TCTGASSAAGKTAAC and it can be spaced up to about 55 nucleotides downstream from the sequence TGAAGT(N)<sub>x</sub>TGAAGT.

The invention includes a recombinant DNA having such a DNA sequence having promoter activity. The recombinant DNA can also include one or more structural genes. The structural gene(s) can encode cytochrome P450 protein(s). The invention includes an expression plasmid including such recombinant DNA and an isolated cell containing the recombinant DNA. The cell is usually eukaryotic.

The invention includes a process for the production of recombinant protein(s), including steps of culturing a cell according to the invention containing recombinant DNA and recovering the protein(s) produced. The protein(s) can be a cytochrome P450, but is not necessarily so. The protein(s) can be a fusion protein.

The invention further includes a method of screening drugs for their effect on activity of a retinoic acid inducible protein, comprising exposing a purified said protein to a said drug and determining the effect on the activity. The activity can be hydroxylation of a retinoic acid, particularly all-*trans* retinoic acid. In particular, the activity can be hydroxylation of a retinoic acid, particularly all *trans*-retinoic acid, at the 4 position of the  $\beta$ -ionone ring thereof.

The invention provides a method of screening drugs for their effect on activity of any protein having P450RAI activity of the present invention, particularly ascertaining the effect a potential drug has on the oxidation of retinoic acid, specifically all-*trans* retinoic acid.

In yet another aspect, the invention includes a method of screening drugs for their effect on expression of a gene wherein the gene is inducible by a retinoid, including the step of exposing a recombinant DNA described above to a potential drug and determining the effect on gene expression. This can include exposing the recombinant DNA in the presence of a said retinoid. The gene can be a reporter gene which does not metabolize retinoic acid. The method can include ascertaining the effect on transcription of the gene.

The invention includes a method of screening a drug for its effect on the metabolism of a retinoid by a cytochrome P450 encoded by a nucleotide sequence of the invention which is incorporated into an expression system so as to be under control of a nucleotide sequence having native promoter activity for a said cytochrome P450, comprising: exposing the system to the drug in the presence of the retinoid so as to determine the effect of the drug on metabolism of the retinoid.

The retinoid can be retinoic acid and it can be all *trans*-retinoic acid.

The invention includes any drug identified according to such a method as having the effect of modulating, and preferably reducing, the activity of a said protein or as having the effect of modulating, and preferably reducing, gene expression.

- 5           The invention includes a method for inhibiting retinoic acid metabolism in an organism in need of such inhibition, comprising administering to the organism an effective amount of a drug identified according to a method of the invention.

#### BRIEF DESCRIPTION OF THE DRAWINGS

- 10           Figure 1 is a schematic representation of the steps used to isolate retinoid-regulated genes using differential display of mRNA.

          Figure 2(a) shows a polyacrylamide gel of PCR amplified mRNA in duplicate obtained using retinoic acid-treated fish (lanes 1 and 2) and dimethyl sulfoxide (DMSO-) treated control fish (lanes 3 and 4). The arrow indicates a PCR amplified band present in the RA-treated samples and not observed in the controls.

- 15           Figure 2(b) shows the nucleotide sequence (SEQ ID NO:1) of the 337 base pair PCR product isolated from the band (arrow) of Figure 2(a). The arrows indicate the nucleotide sequences where the upstream and downstream priming sites for differential display PCR amplification were located in the 3'- untranslated portion of zP450RAI.

- 20           Figure 2(c) shows an amino acid sequence (SEQ ID NO:2) corresponding to cDNA (492 amino acid open reading frame). The boxed residues indicate the heme-binding motif characteristic of cytochrome P450s.

- Figure 2(d) shows amino acid sequence comparisons between zP450RAI and several other cytochrome P450s (SEQ ID Nos: 6,7,8,9,10) in the area of the conserved heme-binding motif found in the superfamily. The cysteine, designated 0 in the figure, which has been  
25           shown to be directly involved in heme-binding [Gotoh, 1989] is surrounded by several highly conserved amino acids.

- Figure 3(a) shows Northern blot analysis of mRNAs obtained from regenerate tissue of RA-treated fish in lane 5, and controls (DMSO-treated fish) in lane 4, using a zP450RAI cDNA probe. Comparison to an RNA ladder (lane 1) shows the major zP450RAI  
30           transcript to be in the 1.4-2.4 kb range.

- Figure 3(b) shows localization of zP450RAI transcripts in regenerating caudal fin tissue 72 hours post-amputation by whole mount *in situ* hybridization. (i) zP450RAI transcripts were found to be undetectable in DMSO-treated regenerates. The original plane of amputation is indicated by the white line with arrowhead; m (soft mesenchyme) and r (bony rays) are  
35           labelled. (ii) In a sample obtained from an RA-treated fish, zP450RAI transcripts, indicated by the black arrowhead, were found to be localized to a band of cells extending across the distal tip of the regenerate. Lower levels of expression of zP450RAI were also evident in non-regenerate tissue at the proximal base of the isolated fin, as indicated by the black line with arrowhead. The plane of amputation is indicated by the white line with arrowhead as in Figure

3(b)(i). (iii) A histological section taken through the plane is indicated by the line. (iv) A histological section of RA-treated fins post-hybridization is shown. Localized expression of zP450RAI was detected in a subset of epithelial cells (black arrowhead) which lie at the distal tip of the regenerate. Basement membrane separating the dense blastemae and the wound epithelium is indicated by the grey arrowhead.

Figure 4 shows elution profiles of lipid soluble extracts obtained from treated media of pSG5-zP450RAI transfected COS-1 cells and pSG5 transfected control cells. Figures 4(a) and 4(b) are plots of cpm vs fraction number for cells incubated with 575 pM [11,12-<sup>3</sup>H]RA for 4 hours and 24 hours, respectively, pSG5-zP450RAI COS-1 cells (—) and control cells (- - -). Metabolism of [11,12-<sup>3</sup>H]RA to total aqueous soluble metabolites was measured using aliquots of the aqueous soluble extract subjected to  $\beta$ -scintillation counting. See insets of Figures 4(a) and (b). Figures 4(c) and 4(d) are plots of absorbance vs retention time for cells incubated with 1  $\mu$ M RA for 4 and 24 hours, respectively. Peaks observed in zP450RAI transfected cell are shaded black. The region of the chromatogram from 4 to 6 min has been expanded (see insets of Figures 4(c) and (d)). In cells transfected with zP450RAI cDNA, the generation of peaks corresponding to 4-oxo RA and 4-OH RA was observed.

Figure 5 shows results obtained with human cell lines probed with a  $\alpha$ -[<sup>32</sup>P]-dATP labeled probe having the sequence identified as SEQ ID NO:11: HEK293; EL-E; HL-60; MCF10A; LC-T; SK-LC6; and MCF7. (+) indicates pretreatment with 10<sup>-6</sup>M RA and (-) indicates no RA pretreatment. The blot was also probed with hGAPDH to control for RNA loading of the gel, shown in the bottom panel.

Figure 6 is similar to Figure 5 for the cell lines U937 and HepG2.

Figure 7 is similar to Figure 5 for the NT2 cell line.

Figure 8 is similar to Figure 5 for a normal NB4 cell line (first two lanes) and three individually derived retinoic acid resistant NB4 derivative cell lines.

Figure 9 shows the murine cDNA amino acid sequence (top row, SEQ ID NO:32) aligned with the human P450RAI amino acid sequence (middle row; SEQ ID NO:4) and the zebrafish cDNA amino acid sequence (bottom row: SEQ ID NO:2).

Figure 10(a) shows elution profiles of lipid soluble extracts obtained from treated media of pSG5-hP450RAI transfected COS-1 cells and pSG5 transfected control cells. Plots of cpm vs fraction number for cells incubated with [11,12-<sup>3</sup>H]RA for 24 hours of pSG5-hP450RAI COS-1 cells (—) and control cells (—) are shown. Figure 10(b) shows measurement of aliquots of the aqueous soluble extract subjected to  $\beta$ -scintillation counting taken to determine metabolism of [11,12-<sup>3</sup>H]RA to total aqueous soluble metabolites. Figure 10(c) shows plots of absorbance vs retention time for hP450RAI transfected cell (—) and control cells (—) cells incubated with 1  $\mu$ M RA for 24 hours. The inset is the region around 10 minutes, expanded for clarity.

Figure 11(a) shows 4-oxo-RA production of pSG5-hP450RAI transfected COS-1 cells and pSG5 transfected control cells.

Figure 11(b) shows 4-OH-RA production of pSG5-hP450RAI transfected COS-1 cells and pSG5 transfected control cells.

Figure 11(c) shows formation of aqueous soluble metabolites of pSG5-hP450RAI transfected COS-1 cells and pSG5 transfected control cells.

5                    Figure 11(d) shows unmetabolized RA of pSG5-hP450RAI transfected COS-1 cells and pSG5 transfected control cells.

Figure 12(a) shows elution profiles of lipid soluble extracts obtained from media of MCF10A cells exposed to RA and unexposed MCF10A control cells. Plots of cpm vs fraction number for cells incubated with [11,12-<sup>3</sup>H]RA for 24 hours of RA-induced MCF10A  
10 cells (—) and control (—) are shown.

Figure 12(b) shows elution profiles of lipid soluble extracts obtained from treated media of MCF7 cells exposed to RA and unexposed MCF7 control cells. Plots of cpm vs fraction number for cells incubated with [11,12-<sup>3</sup>H]RA for 24 hours of RA-induced MCF7 cells (—) and control (—) are shown.

15                    Figure 12(c) shows the total aqueous soluble metabolites measured using aliquots of the aqueous soluble extracts of the cell lines described in Figures 12(a) and (b) subjected to  $\beta$ -scintillation counting. The first two bars are for unexposed MCF7 cells and MCF7 cells exposed to RA, respectively. The third and fourth bars are for unexposed MCF10A cells and MCF10A cells exposed to RA, respectively.

20                    Figure 13(a) shows elution profiles of lipid soluble extracts obtained from microsomal preparations after incubation with radiolabelled RA for ninety minutes, as described in Example 7. Plots of dpm vs fraction number for HeLa P microsomes (■,▲) and HeLa RAI microsomes (▼,◆).

Figure 13(b) shows fractions 5 to 15 of Figure 13(a) on a larger scale.

25                    Figure 14(a) shows SDS-PAGE of the GST-hP450RAI fusion protein. Lane 1 shows the whole cell lysate of *E. coli* expressing the plasmid containing the fusion protein, without induction. Lane 2 shows the whole cell lysate of *E. coli* expressing the plasmid containing the fusion protein after induction with 0.1 mM IPTG.

Figure 14(b) shows SDS-PAGE of the purified GST-hP450RAI fusion protein  
30 after induction with 0.1 mM IPTG, with molecular weight markers at the right hand side indicating the fusion protein to have a molecular weight of about 75 kDa.

Figure 15 shows promoter sequences of human, mouse and zebrafish P450RAI. The boxed regions show highly conserved regions while the arrows indicate spaced apart consensus sequences of RAREs.

35                    Figure 16 shows relative luciferase activity induced in cells containing a luciferase vector into which was cloned a portion of the putative promoter for mP450RAI. Luciferase activity was measured in cell extract supernatants from cells transfected with 3 concentrations of expression vectors comprising cDNAs encoding RXR $\alpha$  and RAR $\gamma$  (100 ng, 500 ng, and 1 $\mu$ g) along with a luciferase-based reporter gene including either the sense or

antisense promoter sequence, or no promoter sequence, grown in presence and absence of RA.

Figure 17 shows inhibition of P450RAI mRNA in MCF7 cells by 4-hydroxy-phenylretinamide (4-HPR).

5                    Figure 18 shows expression of cytochrome P450RAI in MCF7 cells by northern blot analysis, over time, after administration of all-*trans* retinoic acid.

Figure 19 shows expression of cytochrome P450RAI in MCF7 cells by northern blot analysis, over time, after administration of all-*trans* retinoic acid alone, and in the presence of all-*trans* retinoic acid and ketoconazole.

10                   Figure 20 shows expression of cytochrome P450RAI in MCF7 cells by northern blot analysis, over time, after administration of all-*trans* retinoic acid and after administration of Am580.

#### DESCRIPTION OF PREFERRED EMBODIMENTS

Figure 1 outlines the steps used to isolate retinoid-regulated genes using  
15 differential display of mRNA. The cloned products isolated in step 6 of Figure 1 were used for sequencing and screening of *Danio rerio* (*D. rerio*) cDNA libraries. P1, P2 and P3 correspond to fragments from RA induced mRNAs. P4 is a PCR product from a down-regulated mRNA. Details of procedures followed in determination of gene sequences described herein follow.

#### *Danio rerio* stocks

20                   *D. rerio* were kept at 28.5°C in 40 L tanks with 25-30 fish per tank on a 14 hour light-10 hour dark cycle. Tap water was conditioned by the addition of 10 ml of Water Conditioner (Sera Aquatan) and 10 ml of 250 g/L Aquarium Salt (Nutra Fin) per 20 L. 2-3 L of water was changed daily. Amputation of fins was carried out following anaesthetization of the fish in a solution of 0.2% ethyl-m-aminobenzoate methanesulfonic acid (ICN) in conditioned  
25 water. Retinoic acid treatment was performed by adding all-*trans* RA, to a final concentration of 10<sup>-6</sup> M, directly into the tank water two days following amputation. Both control- and RA-treated fish were kept in the dark during the experiments.

#### Differential Display of mRNAs

Differential mRNA display was performed essentially as described by Liang  
30 and Pardee (1992) with appropriate modifications as described herein. Regenerating tissues were collected 3 days post-amputation (24 hours post-RA addition) and quick frozen in liquid nitrogen. Poly (A)<sup>+</sup> RNA was isolated using the Micro Fast-Track kit. Duplicate independent reverse transcription reactions were performed on the isolated poly(A)<sup>+</sup> RNA from both the treated and untreated samples for each specific 3' poly-T primer used (5'-T<sub>12</sub>VN-3'). The  
35 symbol "V" represents A or C or G and not T or U. Several combinations of the 3' poly-T primers given in the first column of Table 1 and the upstream primers given in the second column were utilized for PCR amplification. For each reaction 0.1 µg poly(A)<sup>+</sup> RNA was reverse



transcribed in a 20 µl reaction volume containing 300U Superscript Reverse Transcriptase (Gibco/BRL), 1X Buffer, 20 µM each dGTP, dATP, dCTP and dTTP, 10 µM dithiothreitol (DTT) and 5 pmol of 5'-T<sub>12</sub>VN-3' primer. The reactions were mixed and incubated at 35°C for 60 minutes, followed by 5 minutes at 95°C. PCR amplification was performed in a Perkin Elmer Cetus PCR machine as follows: 1 µl cDNA synthesis reaction, 5U *Taq* DNA polymerase (Gibco/BRL), 1X PCR Buffer, 2 µM each dGTP, dATP, dCTP and dTTP, 10 µCi α-[<sup>35</sup>S]dATP (*redivue*, Amersham) 1.2 mM MgCl<sub>2</sub>, 0.5 µM upstream primer and 0.5 µM of the corresponding 5'-T<sub>12</sub>VN-3' primer. PCR conditions were as follows: 1 cycle, 94°C for 5 minutes; 40 cycles, 94°C for 30 seconds, 42°C for 1 minute, 72°C for 30 seconds; followed by a final extension of 5 minutes at 72°C. 4 µl of the PCR reactions were loaded onto a 6% non-denaturing polyacrylamide gel and electrophoresed at 60 watts, 45°C. The gel was dried and exposed for 12 to 24 hours on Kodak XAR film at room temperature.

Table 1. Sequences of the downstream Poly (T) oligonucleotides for the differential display procedure.	
3'- Poly(T) primers:	5'-degenerate primers:
5'-TTT TTT TTT TTT GG-3'	5'-AAG CGA CCG A-3'
5'-TTT TTT TTT TTT GA-3'	5'-TGT TCG CCA G-3'
5'-TTT TTT TTT TTT GT-3'	5'-TGC CAG TGG A-3'
5'-TTT TTT TTT TTT GC-3'	5'-GGC TGC AAA C-3'
	5'-CCT AGC GTT G-3'
5'-TTT TTT TTT TTT AG-3'	
5'-TTT TTT TTT TTT AA-3'	
5'-TTT TTT TTT TTT AT-3'	
5'-TTT TTT TTT TTT AC-3'	
5'-TTT TTT TTT TTT CG-3'	
5'-TTT TTT TTT TTT CA-3'	
5'-TTT TTT TTT TTT CT-3'	
5'-TTT TTT TTT TTT CC-3'	

In Table 1, the sequences in the first column are identified as SEQ ID NOs: 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22 and 23, respectively. The sequences in the second column are identified as SEQ ID NOs: 24, 25, 26, 27 and 28, respectively.

#### Gel Purification and Reamplification

Bands demonstrating reproducible differential amplifications (see Figure 2a) were found for the upstream-downstream primer combination of 5'-TGCCAGTGGGA-3'- poly-T primer, 5'-TTT TTT TTT TTT AG-3' (SEQ ID NOs: 26 and 16, respectively). These bands were excised from the gel by overlaying the X-ray film and cutting out the corresponding piece of dried gel and filter paper. The PCR product corresponding to a fragment of the protein described herein was isolated from the band in Figure 2(a). Samples were placed in 100 µl of nuclease free water, incubated for 10 minutes at room temperature, then boiled for 15 minutes. The supernatant was recovered following a 15 minute centrifugation at 12,000 x g.

In order to facilitate cloning of the PCR products, several changes were made to the reactions. Primers which included EagI restriction endonuclease sites were used in the reamplification. Based on results obtained in the differential display analysis, the upstream 5'-TGCCAGTGGA-3' primer was replaced by 5'-GTAGCGGCCGCTGCCAGTGGA-3' (SEQ ID NO: 29) and the downstream poly-T primer, 5'-TTT TTT TTT TTT AG-3', was replaced by 5'-GTAGCGGCCGCT<sub>12</sub>-3' (SEQ ID NO:30). In addition, the reaction volume was increased to 40 µl, isotope was omitted and 20 as opposed to 40 cycles were performed. 5 µl aliquots of the PCR reactions were removed and the products were visualized by electrophoresis in a 1% agarose gel followed by ethidium bromide staining and UV illumination.

#### 10 Cloning PCR Products

The reamplified products were purified by phenol/chloroform extraction followed by ethanol precipitation. The resulting DNA pellet was resuspended in 17 µl of sterile water and digested at 37°C for 1 hour by the inclusion of 10U EagI (New England Biolabs), and 1X NEB 3 buffer. EagI restriction endonuclease was heat inactivated by incubation at 65°C for 20 minutes. pBluescript SK<sup>+</sup> vector was prepared by digestion with EagI, followed by dephosphorylation using calf intestinal alkaline phosphatase (CAP, Promega). Restriction digests were purified using the GeneClean II Kit (Bio 101) following electrophoresis in a 1% agarose gel. In a total ligation volume of 10 µl, 2 µl of digested PCR product, 1 µl digested SK<sup>+</sup>, 1U T4 DNA ligase (Gibco/BRL) and 1X buffer were incubated at 16°C overnight. *E. coli* bacterial strain JM109 was transformed with 1 µl of the ligation product using a BioRad Gene Pulser, then plated on LB+ampicillin plates and incubated overnight at 37°C.

#### Colony Selection

Individual colonies were transferred in duplicate to fresh LB plates and grown overnight at 37°C. Colonies were transferred to nitrocellulose membrane and denatured in a solution of 1.5M NaCl, 0.5M NaOH for 5 minutes, neutralized in 1.5M NaCl, 0.5M Tris-HCl, pH 8.0 for 5 minutes, followed by two 5 minute washes in 2X SSC. Membranes were then UV cross-linked (Stratalinker UV Crosslinker, Stratagene). Prehybridization and hybridization were performed using Quickhyb (Stratagene) following the manufacturer's directions. Each colony lift was probed with the corresponding PCR product isolated during the gel reamplification and purification step. α-[<sup>32</sup>P]-dATP labelled probes were generated using the Prime-It Kit II (Stratagene). Subsequent to hybridization, filters were washed twice for 20 minutes in 2X SSC, 0.1% SDS solution at room temperature and exposed to Kodak X-omat autoradiography film overnight at -70°C. Positive colonies were selected from the duplicate plates, grown overnight in LB+ampicillin (100 µg/ml) and plasmid DNA isolated using the Qiaprep Spin Plasmid Kit (Qiagen).

Clones were sequenced using the T7 Sequencing Kit (Pharmacia Biotech). Sequence comparisons were generated using the GeneWorks software package (Intelligenetics).

### Screening of a *D. rerio* cDNA Library

A random primed *D. rerio* 6-18 hour embryo cDNA library constructed in Uni-ZAP II (Stratagene) was produced.  $4.5 \times 10^5$  independent pfu were screened using the random primed,  $\alpha$ - $^{32}\text{P}$ -dATP labelled 337 bp PCR fragment isolated by mRNA differential display as a probe. Filters were prehybridized for 1-4 hours at 42°C in 50% formamide, 5X SSPE, 1X Denhardt's solution, 0.2 mg/ml denatured salmon sperm DNA. Hybridization was performed at 42°C by adding denatured probe to the prehybridization solution. Filters were washed two times for 20 minutes in 2X SSC, 0.05% SDS at room temperature and exposed to Kodak XAR film overnight at -70°C. Positive plaques were picked into 500  $\mu$ l SM buffer and subjected to additional rounds of rescreening until purified. Positive plaques were exposed to the *in vivo* excision protocol following the manufacturer's directions (Stratagene). pBluescript containing colonies were plated onto LB+amp plates and grown overnight at 37°C. Sequence data were generated using the T7 Sequencing Kit (Pharmacia) and analysed using the GeneWorks software package (Intelligenetics).

### 15 Whole Mount *in situ* hybridization

RA- and DMSO-treated regenerates were isolated 72 hours post-amputation (24 hours post RA/DMSO addition), washed in PBS and prepared for whole mount *in situ* hybridization. *In situ* hybridizations were undertaken as previously described [White, 1994].

### Northern Blot Analysis

20 Fish were allowed to regenerate their caudal fins for 72 hours. At 48 hours  $10^{-6}\text{M}$  all-*trans* RA in DMSO vehicle or DMSO alone was added directly to the tank water. mRNA was prepared using the Micro Fast-Track mRNA isolation kit (Invitrogen, CA) according to the manufacturer's directions. 3.0-5.0  $\mu$ g poly A<sup>+</sup> RNA was electrophoresed, blotted and probed using a previously described method [White, 1994] with the full length zP450RAI cDNA  
25 obtained as described below. Ethidium bromide stained agarose gel showed that equivalent amounts of mRNA were used in the blotting experiments. See lanes 2 and 3 of Figure 3(a).

### HPLC Analysis

Media from transfected cells incubated with 575 pM [11,12- $^3\text{H}$ ]RA (Figures 4(a) and 4(b)) or 1  $\mu\text{M}$  RA (Figures 4(c) and 4(d)) for either 4 hrs (Figures 4 (a) and 4(c)) or 24 hrs (Figures 4(b) and 4(d)) were acidified with 0.1% acetic acid. Lipid soluble metabolites were separated from aqueous soluble metabolites using a total lipid extraction of the medium [Bligh, 1957]. Metabolism of [11,12- $^3\text{H}$ ]RA to total aqueous soluble metabolites was measured using aliquots of the aqueous soluble extract subjected to  $\beta$ -scintillation counting (See the insets of  
35 Figures 4(a) and 4(b)). Lipid soluble extracts were evaporated to dryness under a stream of nitrogen and resuspended in 93.5/5/1/0.5 hexane/isopropanol/methanol/acetic acid (H/I/M/AA). Metabolites were separated by HPLC using a Zorbax-SIL (3 $\mu$ , 8 x 0.62 cm) column eluted with a solvent system of 93.5/5/1/0.5 H/I/M/AA at a flow rate of 1 ml/min.

## EXAMPLE 1

### Characterization of a novel cytochrome P450

Transcripts present in fin tissue regenerating in the presence or absence of RA were compared using the differential display PCR technique developed by Liang and Pardee [Liang, 1992] (Fig 2(a)). One of the differential display products which exhibited a dependence on the presence of RA for its expression, indicated by the arrow in Figure 2(a), was isolated and sequenced. The sequence is identified as SEQ ID NO:1 and is also shown in Figure 2(b). The amino acid sequence corresponding to the cDNA, termed here, "zP450RAI", is shown in Figure 2(c) and identified as SEQ ID NO:2. BLAST search analyses revealed sequence homology between zP450RAI and multiple members of the cytochrome P450 superfamily. Alignments between zP450RAI cDNA deduced amino acid sequence and those of other cytochrome P450s indicated that zP450RAI exhibited less than 30% overall amino acid identity with members of previously defined subfamilies [Nelson, 1993]. zP450RAI contains many of the structural motifs which are common to cytochrome P450 family members, including the heme-binding domain located in the C-terminal portion of the protein. See Figure 2(d). The P450RAI family has been designated "CYP26".

## EXAMPLE 2

### Cell specific induction of zP450RAI by all-trans RA

Northern blot analysis of mRNAs expressed in regenerate tissue isolated from control (dimethyl sulfoxide-treated) and RA-treated fish was performed with a full-length zP450RAI cDNA probe. zP450RAI transcripts were not detectable in regenerate tissue from control fish (Figure 3(a), lane 4) but were very noticeably present in tissues isolated from fish exposed to RA for 24 hours (Figure 3(a), lane 5).

Whole mount *in situ* hybridization was used to determine the cellular localization of zP450RAI expression in regenerating fin tissue. Figure 3(b) shows regenerating fins from control and RA-treated fish. zP450RAI transcripts are not detectable in control fin tissue (Figure 3(b)(i)). In regenerating tissue from RA-treated fish, zP450RAI transcripts were found to be abundant in a layer of epithelial cells extending across the distal edge of the wound epithelium as indicated by the black arrowhead in Figure 3(b)(ii). Some low level staining was also observed in inter-ray tissue as indicated by the black line with arrowhead in Figure 3(b)(ii). A histological section of an RA-treated fin, taken along the line shown in Figure 3(b)(iii), is shown in Figure 3(b)(iv). The section indicates that cells expressing zP450RAI are located deep within the epithelial layer at the distal tip of the blastemal mesenchyme. Whole mount *in situ* hybridization thus illustrates the usefulness of nucleic acid probes of the invention for the localization of cytochrome P450RAI mRNA in whole tissue.

### EXAMPLE 3

#### Metabolism of all-*trans* RA by zP450RAI transfected cells

Retinoic acid as a substrate of zP450RAI was studied. The full-length zebrafish zP450RAI cDNA was cloned into the eukaryotic expression vector pSG5 [Green, 1988]. COS-1 cells were transiently transfected with either pSG5 or pSG5-zP450RAI and then incubated with either picomolar concentrations of [11,12-<sup>3</sup>H]all-*trans*-RA or micromolar concentrations of non-radioactive all-*trans*-RA. COS-1 cells are an African green monkey kidney "fibroblast-like" cell line. zP450RAI expression in COS-1 cells promoted the rapid conversion of RA into both lipid- and aqueous-soluble metabolites. See Figures 4(a) and 4(b). Fractions of total lipid extracts of transfected cells were initially separated by normal-phase HPLC on Zorbax-SIL. Comparison between extracts from pSG5 and pSG5-zP450RAI-transfected cells indicated that zP450RAI significantly increased RA metabolism. Incubation of zP450RAI-transfected cells with 575 pM [11,12-<sup>3</sup>H]all-*trans*-RA for either 4 or 24 hours resulted in accumulation of RA metabolites, one of which co-migrated on a column with synthetic standards 4-OH-RA and 18-OH-RA, and a second slightly less polar metabolite which co-migrated with 4-oxo-RA standard (Figures 4(a) and 4(b)). Rechromatography of RA metabolites using other HPLC systems confirmed the identity of these two metabolites as 4-OH-RA and 4-oxo-RA (Table 2). It is possible that the aqueous-soluble radioactivity represents glucuronides of RA metabolites or glucuronides of RA itself. Rapid glucuronidation of 4- and 18-hydroxy-RA in mammalian cell extracts has been reported by others [Wouters, 1992; Takatsuka, 1996].

**Table 2 Chromatographic properties of RA metabolites.**

	Metabolite	Retention Time (min)		
		Z-Sil <sup>a</sup>	Z-CN <sup>b</sup>	Z-ODS <sup>c</sup>
25	RA (std)	2.57	4.47	19.92
	4-oxo-RA (std)	4.79	11.33	11.73
	4-OH-RA (std)	5.17	9.65	12.65
	18-OH-RA (std)	5.06	9.53	14.03
	Peak 1 (RA)	2.57	4.48	19.73
30	Peak 2 (4-oxo-RA)	4.87	11.38	11.57
	Peak 3 (4-OH-RA)	5.16	9.68	12.68

<sup>a</sup>HPLC conditions: Zorbax-SIL column eluted with 93.5/5/1/0.5 H<sub>2</sub>O/MeOH/A.A. (1 ml/min)

<sup>b</sup>HPLC conditions: Zorbax-CN column eluted with 93.5/5/1/0.5 H<sub>2</sub>O/MeOH/A.A. (1 ml/min)

<sup>c</sup>HPLC conditions: Zorbax-ODS column eluted with a 20 min linear gradient with solvent containing 10 mM ammonium acetate which ranged from 55.45 to 5.95 H<sub>2</sub>O /MeOH (2 ml/min).

A similar pattern of zP450RAI-dependent metabolism was also observed using a much higher RA concentration (1 μM). zP450RAI-transfected COS-1 cells incubated for 4 or 24 hours with 1 μM RA generated two closely-running peaks which were discernible in a 350nm HPLC trace shown in Figures 4(c) and 4(d), but which were essentially undetectable in control pSG5-transfected cells (See the insets of Figures 4(c) and 4(d)). These peaks co-migrated with those of 4-oxo-RA and 4-OH-RA standards, respectively. Diode array spectrophotometric

detection of the zP450RAI-generated peaks showed that the spectral properties of the two metabolite peaks matched the standard retinoids [In hexane-based solvents: 4-OH-RA,  $\lambda_{\max}$ =350nm; 4-oxo-RA,  $\lambda_{\max}$ =355nm; in methanol-based solvents: 4-OH-RA,  $\lambda_{\max}$ =340nm; 4-oxo-RA,  $\lambda_{\max}$ =360nm].

- 5           The invention thus includes a retinoic acid metabolizing protein belonging to the family of cytochrome P450s, designated CYP26, and generation of the protein in zebrafish caudal fin wound epithelium being induced in response to RA treatment. While RA metabolizing activity has previously been detected in epithelial tissues of several species [Frolik, 1979; Roberts, 1979; Wouters, 1992; Duell, 1992], an actual enzyme responsible for such
- 10   activity has heretofore been unknown.

zP450RAI is up-regulated by RA treatment and apparently this up-regulation occurs in a specific set of cells in the wound epithelium of regenerating zebrafish caudal fins.

- It might be of relevance to the regulation of the generation of this enzyme *in vivo* that experiments with F9 cells where RARs have been selectively ablated indicate that
- 15   RAR- $\alpha$  and RAR- $\gamma$  might have a role in the regulation of RA metabolism [Boylan, 1995]. The expression of both RAR- $\alpha$  and RAR- $\gamma$  in the regenerating caudal fin is consistent with the suggestion that they may be involved in the regulation of P450RAI expression by RA [White, 1994].

#### EXAMPLE 4

##### 20   Cloning of Human P450RAI

- The amino acid sequence corresponding to the DNA of zebrafish P450RAI (zP450RAI) (SEQ ID NO:2) was used to search an express sequence tag (EST) database. A commercially available EST clone (SEQ ID NO:11) having a high degree of homology with a C-terminal portion of the zP450RAI (from Glu 292 to Phe 410 of SEQ ID NO:2) was purchased
- 25   (Research Genetics, Huntsville, AL). The clone is reportedly from a human infant brain cDNA library (Bento Soares and M. Fatima Bonaldo) and is apparently otherwise unpublished. The purchased clone was sequenced using the T7 sequencing kit (Pharmacia) and sequence data was generated using the Geneworks Software Package (Intelligenetics).

- A cDNA library generated from an NT2 cell line treated with retinoic acid is
- 30   commercially available (Stratagene, cat#939231) and this product was used for further studies. The cDNA library was probed with a nucleic acid having a sequence identified as SEQ ID NO:11. Eleven positively hybridizing clones were isolated and purified according to the manufacturer's directions. Sequence data for these clones were generated using the T7 Sequencing Kit (Pharmacia) and analyzed using the Geneworks package (Intelligenetics). The
- 35   human DNA sequence is identified as SEQ ID NO:5 and the corresponding polypeptide as SEQ ID NO:4.

## EXAMPLE 5

### Isolating Mouse P450RAI

It has been found by the present inventors (results not shown) that RA can induce mRNA transcripts which cross hybridize with a hP450RAI cDNA probe in either of the F9 and P19 mouse cell lines having 4-hydroxylase activity, as described by Blumberg *et al.* [Blumberg *et al.*, 1995; Achkar *et al.*, 1996].

A retinoic acid-treated P19 teratocarcinoma cDNA library in lambda Unizap XR (Stratagene) was screened using <sup>32</sup>P-labelled full length human P450RAI as a probe. Nitrocellulose filters were hybridized overnight at 37°C in a buffer containing 50% formamide, 1x Denhardt's, 5x SSPE, 0.1% SDS, and 100 µg/ml boiled salmon sperm DNA, and washed in 2x SSC, 0.1% SDS for 2x 15 minutes at room temperature followed by 20 minutes at 37°C. From a total of 600,000 plaque forming units of phage, 72 positives were isolated. Of these, 18 were purified, as described above, and one was found to have a full-length cDNA insert 1.7 kb in length homologous to the full-length human clone previously isolated. The murine DNA sequence is identified as SEQ ID NO:31 and the corresponding polypeptide as SEQ ID NO:32. Figure 9 shows aligned portions of the amino acid sequence of the mouse protein (SEQ ID NO:32) with those of the human protein (SEQ ID NO:4) and the zebrafish protein (SEQ ID NO:2).

## EXAMPLE 6

### Transient Transfection Analysis

COS-1 cells were subcultured 20 hours prior to transfection which was carried out according to the standard DEAE-dextran method [Sambrook, 1989; Maniatis, 1982]. Cells were transfected with pE-AR (adrenodoxin expression vector, 1 µg/P100 plate) and pE-ADX (adrenodoxin reductase expression vector, 1 µg/P100 plate) together with 3 µg per plate of either pSG5 (control) or hP450RAI-pSG5 (experimental). [11,12-<sup>3</sup>H]all-*trans* retinoic acid (60,000 cpm per plate) was added 24 hours after transfection. Analyses were carried out as described in Example 3 and results obtained are shown in Figures 10 and 11(a) to 11(d). As indicated in the Figures, hP450RAI expression in COS-1 cells promoted conversion of RA into 4-OH-RA and 4-oxo-RA. Total amounts of 4-oxo-RA and 4-OH-RA produced in the transfected cells in comparison to amounts produced in the control cells are shown in Figures 11(a) and (b), respectively. Overall, greater amounts of aqueous soluble metabolites were produced in the transfected cells (Figure 11(c)) and greater amounts of unmetabolized RA were found in control cells (Figure 11(d)).

The clone sequence (SEQ ID NO:11) was prepared as a <sup>32</sup>[P]-dATP labeled probe to study the inducibility of hP450RAI by RA in several cell lines: HEK293; EL-E; HL-60; MCF10A; LC-T; SK-LC6; MCF7; U937; HepG2; NT2 (See Figures 5 to 7). As can be seen, a variety of expression patterns were observed. The SK-LC6 human lung (epithelial) line appeared to constitutively express corresponding mRNA. There was apparently some increase in expression in the HEK293 (human embryonic kidney), LC-T (human lung epithelial), HepG2

(human liver, epithelial in morphology), NT2 (pluripotent human embryonic carcinoma) and U937 (human monomyelocytes) cell lines in response to addition of RA. There was a large dependence on exposure to RA in the MCF7 (human breast carcinoma (epithelial)) cell line. Some cell lines showed no expression in the absence or presence of RA: EL-E; HL-60 and

5 MCF10A.

The  $^{32}$ [P]-dATP labeled probe was also used to study hP450RAI mRNA expression in a human acute promyelocytic leukemia cell line. Experiments were carried out using the NB4 cell line, isolated from a human acute promyelocytic leukemia patient, and three retinoic acid resistant cell lines were independently derived from NB4. Results are shown in

10 Figure 8. As can be seen, the normal cells expressed hP450RAI mRNA after treatment with  $10^{-6}$ M RA, while such expression appeared to be absent for the other cell lines both in the absence and presence of RA.

Analysis of metabolites of MCF10A and MCF7 cell lines exposed to RA was carried out, MCF10A cells having displayed no expression of mRNA and latter having displayed

15 a large dependence of mRNA expression on exposure to RA. The results are shown in Figures 12(a) to 12(c). Consistent with the results shown in Figure 5, the results shown in Figure 12(a) indicate there was little difference in the lipid soluble activity profiles of the MCF10A cell line exposed to RA and the control. The last two bars of Figure 12 (c) indicate that total aqueous soluble metabolites were about the same for both the induced and control MCF10A cells. As

20 indicated in Figure 12(b), the MCF7 cell line exposed to RA had an elution profile which indicated significantly greater concentrations of 4-OH-RA and 4-oxo-RA than the same cell line not exposed to RA. Figure 12(c) indicates that the amount of total aqueous soluble metabolites of the MCF7 cells exposed to RA was much greater than that for the control cells. Again, these results are consistent with those obtained in the Northern blot analysis shown in Figure 5 for the

25 MCF7 cell line.

#### EXAMPLE 7

##### Generation of a Stable Cell Line using Human P450RAI: HeLa Cells Expressing Sense Construct

For expression in HeLa cells, the human cytochrome P450RAI cDNA (SEQ ID

30 NO:5) was inserted into the XhoI-NotI sites of the multiple cloning site of the Epstein-Barr virus-based vector pCEBV7 [Wilson, 1995]. Stable transfection was performed via the calcium phosphate method [Sambrook, 1989]. Prior to the day of transfection, HeLa cells were seeded at  $3.0 \times 10^6$  cells per 100 mm plate. Approximately 12  $\mu$ g of DNA were transfected per plate and triplicate plates were employed for the transfection. Selection using hygromycin B began

35 three days after the transfection and continued for approximately three weeks until the development of foci on the plates. The concentration of hygromycin B (100  $\mu$ g/ml) was chosen for selection of cells with high expression of the construct. A killing curve was determined prior to selection which showed that 50  $\mu$ g/ml of hygromycin was sufficient to kill 50% of the cells in 4



days. Confirmation of the selected HeLa cells expressing the sense construct was determined by Northern blot analysis and probing with full length hP450RAI cDNA (data not shown).

- 5        Microsomes were prepared from HeLa cells transfected with the pCEBV7 alone (HeLa P) or from the HeLa cells expressing the P450RAI (HeLa RAI) and exposed to radiolabelled RA for ninety minutes. The results are shown in Figures 13(a) and 13(b), in which it can be seen that when microsomes prepared from these cells are incubated with RA, only those from HeLa RAI showed any conversion of retinoic acid to 4-hydroxy-retinoic acid or 4-oxo-retinoic acid .

#### EXAMPLE 8

##### 10    **Expressing a Fusion Protein Containing Human P450RAI**

- The cDNA of hP450RAI was inserted into the GST gene fusion vector, pGEX3X using the Sma1 site. The construct generated lacked 177 bp of the N-terminal portion of the hP450RAI cDNA, which encodes a hydrophobic region characteristic of a signal sequence. *E. coli* (JM109) cells were used for expression of vectors and construct. After  
15    screening for positive colonies expressing the construct in the proper orientation, expression of the fusion protein was carried out according to the manual provided by Pharmacia.

- GST-hP450RAI fusion protein was generated via electroporation of *E.coli* (JM109) cells with the construct, followed by induction of gene expression. Expression of the fusion protein is under the control of the *tac* promoter which is inducible by the lactose analog,  
20    isopropylbeta-thiogalactoside (IPTG).

- Cultures were grown overnight in LB-ampicillin broth. The following day, approximately 1:100 dilution factor of the overnight culture was transferred to a fresh tube containing LB-ampicillin broth. The culture was allowed to grow for three hours and induced with IPTG to a final concentration of 0.1 mM for an additional 2.5 hours. After induction, the  
25    culture was centrifuged at 7,700x g for 10 minutes at 4°C to sediment the cells. The pellet was resuspended in a ratio of 50µl of culture/ml of ice-cold 1X PBS. The cells were lysed with a sonicator at 30 second intervals while submerged in ice-water. Triton X-100 was added to the lysed cells to a final concentration of 1% and the mixture was incubated for 30 minutes at 4°C to allow solubilization. The suspension was subjected to centrifugation at 12,000x g for 10  
30    minutes at 4°C.

- Binding of the GST portion of the fusion protein to glutathione beads was used to purify the protein. 2 ml of a 50% slurry of reduced glutathione beads equilibrated with 1 X PBS were used per 100 ml of sonicate mixture from above. After the addition of the beads, the material was gently agitated for 30 minutes at room temperature to allow binding of the fusion  
35    protein to the beads. The suspension was centrifuged at 500 x g for 5 minutes to separate glutathione beads with bound fusion protein from other cellular components. The glutathione beads were washed three times with cold PBS to remove non-specifically bound material.

          The fusion protein was eluted from the glutathione beads using an elution buffer containing reduced glutathione. Approximately 1.0 ml of glutathione elution buffer per

100 ml of sonicate suspension was used. The bound glutathione beads were allowed to mix at room temperature for 10 minutes to elute the fusion protein. Centrifugation of the eluted beads was performed at 500 x g for 5 minutes. Both the elution and centrifugation were performed three times. Figures 14(a) and (b) show the results of SDS-PAGE of the GST-hP450RAI fusion protein.

It is generally possible to carry out some functional tests with the GST protein covalently attached to the protein of interest, but removal is preferred for the production of antibodies. The GST-hP450RAI fusion protein contains a site recognized by factor Xa. Treatment of the eluted fusion protein with factor Xa yields both the GST protein and hP450RAI protein. Alternatively, it is possible to cleave the fusion protein prior to its elution from the beads, but inadequate cleavage many occur. Approximately 10 µg of factor Xa/mg of fusion protein is used. The material is mixed gently and incubated for approximately 2-16 hours at room temperature. Completion of digestion is checked by gel eletrophoresis at various time points. To purify the protein of interest from the GST protein, binding of cleaved fusion protein suspension with glutathione beads is carried out, followed by centrifugation. The protein of interest remains in the supernatant.

The GST fusion protein can be assayed immunologically or biochemically. Biochemically, treatment of the GST fusion protein with the GST substrate, 1-chloro-2,4-dinitrobenzene (CDNB), indicates the amount of the fusion protein. The GST protein catalyzes the conjugation of CDNB with glutathione, resulting in a CDNB-glutathione product with a strong molar absorptivity at 340 nm. Immunologically, a Western blot of the fusion protein can be probed with an anti-GST antibody which can be readily detected using a secondary antibody such as an anti-goat IgG alkaline phosphatase conjugate, and the standard colorimetric reaction.

## EXAMPLE 9

### Mapping of Human P450RAI

A 1.3 kb cDNA of hP450RAI was mapped using a P-1 derived artificial chromosome (PAC) library. Mapping of the cDNA and genomic PAC clone was performed by fluorescence *in situ* hybridization [Lichter, 1990] to normal human lymphocyte chromosomes counterstained with propidium iodide and DAPI. Biotinylated probe was detected with avidin-fluorescein isothiocyanate (FITC). Images of metaphase preparations were captured by a thermoelectrically cooled charge coupled camera (Photometrics, Tucson, Arizona). Separate images of DAPI banded chromosomes [Heng, 1993] and FITC targeted chromosomes were obtained. Hybridization signals were acquired and merged using image analysis software and pseudo colored blue (DAPI) and yellow (FTIC) [Boyle, 1992] and overlaid electronically.

Positive hybridization signals were found to be localized to 10q23-24. The band assignment was determined by measuring the fractional chromosome length and by analyzing the banding pattern generated by the DAPI counterstained image.

#### EXAMPLE 10

##### Regulation of P450RAI transcription - Cloning of P450RAI Promoters

**Cloning of Zebrafish P450RAI promoter.** An adult zebrafish genomic library ( $1.0 \times 10^6$  pfu)

was screened with the full length cDNA corresponding to zP450RAI and positive plaques

- 5 purified through secondary and tertiary screening. Lambda DNA corresponding to positive plaques was prepared, and the inserts from these clones were excised by restriction enzyme digestion with NotI and subcloned into the plasmid SK+ (Stratagene). Genomic clones were analyzed by restriction enzyme digestion using enzymes from the polylinker of SK+, followed by
- 10 Southern blotting using an oligonucleotide (5'-GTAGCACGGATGGTG-3') which hybridizes to the nucleotide sequence encoding the N-terminus of the zP450RAI cDNA to identify fragments of the genomic clones which encode the N-terminal region. A 772 base pair PstI fragment which hybridized with the oligonucleotide probe was purified, ligated into the vector SK+ and sequenced using the Core Facility for Protein/DNA Chemistry at Queen's University, Kingston, Canada. Sequence analysis identified this clone as containing the putative initial methionine
- 15 followed by 129 base pairs of coding sequence, plus 651 nucleotides upstream (5'). Within this 772 base pair fragment, a 402 base pair HindIII fragment was found to contain the putative retinoic acid response element (RARE). This fragment was subcloned into the pGL3B luciferase vector, in both the forward and reverse orientations, (Promega) for transient transfection analyses.

- 20 **Cloning Human P450RAI promoter.** The full length hP450RAI cDNA was used as a probe to identify PAC (P1 artificial chromosome) clones which contain the hP450RAI gene. cDNA from hP450RAI was sent to Canadian Genome Analysis and Technology Program at the Hospital for Sick Children in Toronto, Ontario, Canada for screening of PAC libraries. 5 PAC clones were obtained from this screening, which were verified to contain the hP450RAI gene by restriction
- 25 enzyme digestion and Southern blotting using the full length hP450RAI cDNA as a probe. One of these clones, 245C7, was found to hybridize to an N-terminal probe from hP450RAI. The probe used was an approximately 130 bp NotI fragment generated from the hP450RAI cDNA. Digestion and Southern blotting of clone 245C7 identified an approximately 3.5 Kb BamHI fragment which hybridized with the NotI fragment. This fragment was subcloned into the
- 30 plasmid SK+ and sequence data generated at the Core Facility for Protein/DNA Chemistry at Queen's University, Kingston, Canada. Comparison of the sequence data generated with the hP450RAI cDNA identified this 3.5 Kb clone as containing the potential initial methionine and approximately 675 bp upstream (5').

**Cloning of mouse P450RAI promoter.** A clone of mouse P450RAI genomic DNA

- 35 approximately 17 kb long was isolated from an SV129  $\lambda$ DASH library and subcloned into SK. DNA prepared from this plasmid was digested with various restriction endonucleases, electrophoresed on an agarose gel, and Southern blotted onto nitrocellulose. The resulting blot was hybridized with a  $^{32}\text{P}$ -labelled 230 base pair probe from the N-terminal region of a mouse P450RAI cDNA clone. A SacI fragment approximately 520 base pairs in length was
- 40 found to hybridize strongly to the probe. This fragment was subcloned into SK cleaved with

SacI. Sequence analysis revealed the presence of a DR5-type RARE in the proximal promoter. Flanking this RARE were two BssHII sites 193 base pairs apart. This 193 base pair BssHII fragment was subcloned into the MluI site of pGL3B. Diagnostic restriction digests with XhoI were done to isolate clones with the BssHII fragments in both forward and reverse orientations.

- 5 Within the 193 base pair fragment is also found the TATA box, 45 base pairs downstream of the RARE. Also included in the 193 base pair BssHII fragment is 83 base pairs of DNA upstream of the DR5 RARE.

- The existence of retinoic acid response elements (RAREs), which bind retinoic acid receptors (RARs) and retinoid-x-receptors RXRs is known. The three promoters of
- 10 P450RAIs from different species described above were thus examined for the presence of RAREs, which might indicate that the regulation of expression of P450RAI is at the transcriptional level. RAREs bind RARs in the form of a heterodimer and typically include a direct repeat of the consensus motif 5'-(A/G)G(T/G)TCA-3'. The repeated sequences can be separated by 1, 2 or 5 nucleotides. Figure 15 shows the promoter sequences for the zebrafish,
- 15 human and mouse P450RAI genes. There is a highly conserved motif, indicated by the arrows, which corresponds to a consensus retinoic acid response element known as a DR5, consisting of a direct repeat of a motif AGTTCA separated by 5 nucleotides. This motif in the P450RAI promoters extends over the sequence TGAAC (the complement of AGTTCA). There is also a highly conserved region extending on either side of the DR5, the entire region being boxed.
- 20 The degree of conservation of this region of the promoter between all three species suggests that this region is important to the control of RA response and that RAR/RXRs are acting in conjunction or competition with other factors binding to this region whose identities are currently unknown. A portion of the conserved region corresponds to the sequence 5'-CAATTAAA-3' which corresponds very closely to a homeobox binding motif, suggesting that homeobox
- 25 proteins may facilitate or compete with RAR/RXR heterodimers in the regulation of P450RAI expression.

#### EXAMPLE 11

##### Transfection with Mouse P450RAI Promoter

- A 195 basepair fragment of genomic DNA containing a portion of the putative
- 30 promoter for mP450RAI was cloned into the luciferase vector, pGL3B (Promega). For analyses of promoter activity, HepG2 cells were transfected in 6-well dishes with 2 µg BssHII-pGL3B sense or antisense constructs, described in Example 10, using 5 µl lipofectAMINE reagent (Gibco, BRL).

- On the first day, 48 hours prior to transfection, cells were replated into 6-well
- 35 plates, with 2.5-3 mls Minimal Essential Medium (MEM) (Gibco, BRL)+ 10% Fetal Calf Serum (Gibco, BRL). On the third day, cells are generally about 50% confluent (HepG2). Before beginning transfection, the medium was replaced with fresh medium and the cells were allowed to grow while preparing DNA/lipofectamine mix. In individual wells of a 48 well plate were mixed 1-5 µg DNA in 100 µl optiMEM (Gibco, BRL) and 5 µl lipofectAMINE reagent in 100

µl optiMEM, by addition of the DNA/optiMEM to the lipofectAMINE/optiMEM with gentle mixing. This was left to sit 15 min to 1 hour at room temperature. To each well were added 800 µl optiMEM to obtain a final volume of 1 ml. The cells were washed 2 times in 1X PBS and once in optiMEM. The 1 ml DNA/lipofectamine/optiMEM mixture was added to the cells and  
5 incubated for 20 hours at 37°C.

The effects of retinoic acid on promoter activity were analyzed by cotransfecting with varying amounts (100 ng to 1 µg) of expression vectors including cDNA sequences encoding zebrafish retinoic acid receptor gamma (RAR-γ) and zebrafish retinoid x receptor alpha (RXR-α). Comparisons were made between cells transfected with the control  
10 pGL3B vector, those with the sense construct and those with the antisense construct by incubating one set of the transfected cells with DMSO and the other set with 10<sup>-6</sup> M RA in DMSO.

On the fourth day, the medium was removed and replaced with fresh medium (+ 10% FBS). The RA or vehicle was added as required to the wells and mixed gently by  
15 swirling the plate. For a 6 well plate 3 µl of 10<sup>-3</sup> M RA in DMSO were added. Control wells received 3 µl DMSO. The plates were covered in foil and incubated for 24 hours at 37°C.

On the fifth day, cells were harvested by removing the medium and washing twice in 1X PBS. Then, on ice, 100 µl lysis buffer (1% Triton X-100, 25 mM glycylglycine, pH 7.8, 15 mM MgSO<sub>4</sub>, 4 mM EGTA, 1mM DTT (added fresh just before use)) were added, and the  
20 cells were scraped off the bottom of the dish and transferred to a 1.5 ml microcentrifuge tube, spun for 5 minutes at 12000 x g, and the supernatant recovered.

To assay for luciferase activity, 20 µl supernatant from lysed, pelleted cells were transferred to a fresh tube. 80 µl luciferase assay buffer were added and a reading in millivolts in a luminometer was taken immediately. The luciferase assay buffer was 20 mM  
25 Tricine, 1.07 mM (MgCO<sub>3</sub>)<sub>4</sub>Mg(OH)<sub>2</sub>\*5H<sub>2</sub>O, 2.67 mM MgSO<sub>4</sub>, 0.1 mM EDTA, 33.3 mM DTT, 0.27 mM Coenzyme A, 0.47 mM Luciferin, 0.53 mM ATP. The results are shown in Figure 16, in which it can be seen that enhanced luciferase activity was observed in the presence of RA, RXRα and RARγ, for both orientations of the promoter sequence, although enhancement appears to be greater for the sense construct.

## 30 **EXAMPLE 12**

### **Inhibition of P450RAI Induction by 4-Hydroxyphenylretinamide**

It has recently been reported that 4-hydroxyphenylretinamide (4-HPR) inhibits RA-induced RA catabolism by NB4 cells [Taimi, 1997]. It was suggested that 4-HPR may inhibit the cytochrome P450 enzyme(s) responsible for RA oxidation to competitively inhibit RA  
35 catabolism. However, any such enzymes were not identified, no explanation of the nature of 4-HPR inhibition of RA catabolism was provided, and no evidence of 4-HPR metabolism was observed.

Experiments to determine the effect of 4-HPR on the induction of P450RAI were thus carried out. Figure 17 illustrates the ability of the synthetic retinoid, 4-HPR to inhibit

the induction of P450RAI by RA. MCF7 cells were grown in culture in minimal essential medium (MEM) (Gibco) supplemented with 10 % fetal calf serum, insulin (.01 mg/mL), MEM non-essential amino acids (as directed by the manufacturer - Gibco), sodium pyruvate (500 nM), L-glutamine (2 mM) gentamycin (10 µg/mL), penicillin (5 µg/mL), streptomycin (5 µg/mL), and fungizone (200 ng/mL). MCF7 cells cultured in parallel were treated for 12 hours with: 10 µM 4HPR; 1 µM 4HPR; 1 µM RA; 1 µM RA and 10 µM 4-HPR; or 1 µM RA and 1 µM 4-HPR. At the end of the 12 hour treatment, total RNA was extracted from the cells using TRIzol reagent (as outlined by the manufacturer - Gibco). P450RAI message in the total RNA preparations was analyzed by northern blot hybridization. The blot was reprobbed with a probe corresponding to the GAPDH cDNA to control for equivalent loading of RNA in each lane of the blot. The results indicate that 4-HPR treatment alone does not induce the P450RAI message. As expected, RA treatment of MCF7 cells results in a marked induction of P450RAI message following 12 hours of incubation. However, when cells are treated with RA in the presence of 10 µM 4-HPR, there is a noticeable suppression of P450RAI induction.

#### 15 **EXAMPLE 13**

##### **P450RAI expression in the presence of retinoic acid**

Figure 18 shows a time course of expression of cytochrome p450 RAI following treatment of MCF7 breast epithelium adenocarcinoma cells with 1 µM all-*trans* retinoic acid. In this northern blot analysis, total RNA was extracted at the indicated time points and transferred to nitrocellulose following electrophoresis in a 1% agarose, 0.66 M formaldehyde gel. The nitrocellulose was then probed with radioactively labelled human cytochrome p450RAI cDNA or GAPDH cDNA to control for equivalence of mRNA loading. This shows that after 3 hours of incubation with retinoic acid, the MCF7 cells express high levels of P450RAI message and following 12 hours of exposure, the message declines sharply, possibly indicating that the metabolic activity of induced P450RAI protein is reducing the concentration of active retinoic acid in the surrounding medium. This strongly suggests that the induction of P450RAI in MCF7 cells forms an autoregulatory negative feedback loop.

#### **EXAMPLE 14**

##### **P450RAI expression in the presence of retinoic acid and ketoconazole**

Figure 19 shows a time course of P450RAI mRNA expression in MCF7 cells similar to that described in Figure 18, except that the effect of the broad spectrum cytochrome p450 inhibitor ketoconazole on P450RAI expression was examined. In the absence of ketoconazole, lanes 1 to 5 in Figure 19 a time course similar to that shown in Figure 18 is shown. In cells which were exposed to 1 µM ketoconazole (replenished every 12 hours following initial treatment), cytochrome P450RAI message was detectable at high levels at 24 and 48 hour time points indicating that the breakdown of retinoic acid can be inhibited by a cytochrome P450 inhibitor and that P450RAI metabolism may be responsible for the sharp

drop in P450RAI message in the absence of ketoconazole. The example illustrates a method of identifying P450RAI inhibitors.

#### EXAMPLE 15

##### P450RAI expression in the presence of Am580

5                   Figure 20 shows a time course of P450RAI mRNA expression in MCF7 cells similar to that described in Figure 18, except that a comparison was made between the retinobenzoic acid derivative Am580 and all-*trans* retinoic acid. Retinoic acid shows a typical time course of induction of P450RAI message, lanes 1 to 4, Figure 20. Am580 induces P450RAI message to levels comparable to those observed following treatment with retinoic acid. Notably, whereas P450RAI message has declined sharply between 24 and 48 hours, for retinoic acid treated cells, the levels of P450RAI in Am580 treated cells remains high at this time point. This indicates that the synthetic retinoid Am580 is resilient to metabolism in these cells and illustrates the utility of identifying such compounds for therapeutic use. For example, the resistance to retinoic acid treatment observed in acute promyelocytic leukemia due to increased retinoic acid metabolism [Warrell, 1994] might be circumvented by treatment with a metabolism-resistant retinoid. P450RAI protein may be a useful agent for screening for these types of compounds.

#### EXAMPLE 16

##### Monoclonal antibodies to P450RAI

20                   Monoclonal antibodies (Mab's) specific for P450RAI are useful, for example, for diagnostic purposes such as for determining P450RAI protein levels in the identification of normal and tumor tissues which metabolize RA. To produce these antibodies, purified P450RAI protein is prepared. The human P450RAI protein is produced in bacterial cells as a fusion protein with glutathione-S-transferase using the vector pGEX2 (Pharmacia). This permits purification of the fusion protein by GSH affinity chromatography. In another approach, P450RAI is expressed as a fusion protein with the bacterial maltose binding domain. The fusion protein is thus recovered from bacterial extracts by passing the extract over an amylose resin column followed by elution of the fusion protein with maltose. For this fusion construct, the vector pMalC2, commercially available from New England Biolabs, is used. This vector has been used in the past, for example, to overexpress nuclear receptor proteins which were recovered in high yields for functional studies and the production of receptor specific antisera [Ohno, 1993]. The preparation of a second fusion protein is also useful in the preliminary screening of MAB's.

35                   The generation of hybridomas expressing monoclonal antibodies recognizing P450RAI protein is carried out as follows: BALB/c mice are injected intraperitoneally with protein/adjuvant three times at one-month intervals, followed by a final injection into the tail vein shortly prior to cell fusion. Spleen cells are harvested and fused with NS-1 myeloma cells (American Type Culture Collection, Rockville, MD) using polyethylene glycol 4000 according to

standard protocols [Kennett, 1979; Mirski, 1989]. The cell fusion process is carried out as described in more detail below.

The fused cells are plated into 96-well plates with peritoneal exudate cells and irradiated spleen cells from BALB/C mice as feeder layers and selection with hypoxanthine, aminopterin, and thymidine (HAT medium) is performed.

An ELISA assay is used as an initial screening procedure. 1-10 µg of purified P450RAI (cleaved from the fusion protein) in PBS is used to coat individual wells, and 50-100 µl per well of hybridoma supernatants is incubated. Horseradish peroxidase-conjugated anti-mouse antibodies are used for the colorimetric assay.

As a secondary screening, breast epithelial carcinoma MCF-7 cells are used as a source of P450RAI determinant. As indicated above, untreated MCF-7 cells exhibit no detectable expression of P450RAI message and no detectable RA metabolizing activity, whereas there is a rapid and striking induction of both in cells treated with RA. Untreated MCF-7 cells thus serve as a negative control for background. MCF-7 cells are aliquotted into 96 well plates one day prior to assay. Cells are fixed and permeabilized with methanol. Hybridoma supernatants are added and fluorescein isothiocyanate (FITC) -conjugated anti-mouse antibodies are used for screening using a standard fluorescence microscope.

Positive hybridomas are cloned by limiting-dilution and grown to large-scale for freezing and antibody production. Various positive hybridomas are selected for usefulness in western blotting and immunohistochemistry, as well as for cross reactivity with P450RAI proteins from different species.

The selected MAb's are useful for monitoring the levels of expression of P450RAI protein following RA treatment in cell culture and in tissues. P450RAI protein expression may be a prognostic indicator for determining whether a particular tumor will respond to RA treatment. There is also a wide intersubject variability in baseline RA metabolism and there is evidence suggesting that subjects with a high rate of RA metabolism have a higher incidence of squamous or large cell cancers of the lung [Rigas, 1996]. Once useful antibodies are characterized, these antibodies are used to survey tumor tissue samples for P450RAI expression.

### 30 Protocol For Production of Mouse Hybridomas

**Fusion.** Feeder cells (spleen and peritoneal exudate cells) are plated. 24 to 48 hours before fusion, mouse myeloma cells are taken off drug (8-azaguanine 20 µg/ml) and counted to ensure that there are at least  $50 \times 10^6$  cells. 2 g of PEG 4000 are autoclaved in a glass tube for 15 minutes and maintained at 60°C for use or alternatively stored at room temperature and remelted in a 60°C water bath when needed.

BALB/c mice are immunized as per desired schedule. The final injection is given intravenously in the tail vein. Fusion of immunized spleen cells is carried out 3 or 4 days after the intravenous injection. Spleen from each animal is collected separately; eye sera for ELISA if desired.



A single cell suspension is prepared using a Teflon pestle and decanting connective tissue. The suspension is washed 1x in serum-free medium. Each spleen has about  $10 \times 10^6$  cells. The myeloma cells are collected, counted and washed in serum-free medium.

- 5                   The cells are then fused. A small beaker of water, and serum-free medium ( $37^\circ\text{C}$ ) are prepared and the PEG melted at  $50-60^\circ\text{C}$ . The immunized spleen cells and myeloma cells are mixed in a 50 ml TC tube (recommended ratios vary from 1:1 to 2:1) and the cells are washed once with serum-free medium. The supernatant is carefully discarded. 2.4 ml pre-warmed serum-free medium is added immediately with pipette to the melted PEG and mixed, maintaining the temperature at  $37^\circ\text{C}$  in beaker of warm water. The PEG should be light pink. If it is yellow, another aliquot should be used. 0.5-1.0 ml of PEG is added dropwise to the cell pellet over 1 minute with gentle rotation of the tube or gentle stirring to ensure mixing. The tip of the addition pipette is placed directly over the cell pellet. The tube is swirled gently in  $37^\circ\text{C}$  water bath for 90 seconds with the blunt end of a 3 ml pipette tip and 10 ml warm serum-free medium added slowly over 6-10 minutes while rotating tube gently to bring the volume up to 20-50 ml. The tube is maintained at  $37^\circ\text{C}$  for at least 20 minutes to obtain cell fusion and then the cells are washed 2x with serum-free medium. The cells are centrifuged and gently resuspended in 100 ml of pre-warmed medium + 10-20% FBS. 100  $\mu\text{l}$ /well aliquotted in 96-well plates. Assuming one spleen fused with  $100 \times 10^6$  cells myeloma fusion partner, about 10 plates are needed. On the following day, 100  $\mu\text{l}$  medium is removed and 100  $\mu\text{l}$  2 x HAT added. Feed with 1 x HAT medium for 1 to 3 weeks, then feed with HT medium (i.e., remove 1/2 HAT medium and replace with equal volume HT medium).

- Preparation of Peritoneal Exudates and Spleen Feeder Cells.** A sacrificed mouse is sprayed with 70% alcohol, skin is nicked and torn apart, with care being taken not to cut the peritoneum. The peritoneum is lifted with forceps and a needle is introduced; 5 ml of serum-free medium is slowly injected. The abdomen is massaged and the fluid is slowly sucked up, collected in a sterile tube and kept on ice. The volume is brought up to 5 ml. The spleen is obtained and placed in a sterile tube containing serum-free medium. The spleen is gently mashed with a sterile Teflon pestle. Clumps are allowed to settle and the cells are decanted into a clean tube, care being taken to avoid including connective tissue, in order to minimize fibroblast growth. The sample is irradiated at 4500 R. Cells are washed once with serum-free medium, placed into 96-well plates [one spleen/10 plates (approximately  $2-5 \times 10^5$  cells/well) and peritoneal exudate cell suspension (PECS) ( $< 3 \times 10^3$  cells/well) in a total volume of 100  $\mu\text{l}$ /well] and incubated at  $37^\circ\text{C}$  until ready to be used. They can also be stored in sterile tube overnight at  $4^\circ\text{C}$ .

## Discussion

It is possible to compare the zP450RAI, mP450RAI and hP450RAI sequences described above. Of the 492 amino acids of zP450RAI (SEQ ID NO:2), it is possible to align 334 amino acids with the 497 amino acids of hP450RAI (SEQ ID NO:4). See Figure 9. On this

basis, there is about 68% homology between the human and fish proteins. The degree of homology between the two amino acid sequences is slightly greater towards the C-terminus than in the N-terminal region. It also appears as though nucleic acid sequences encoding the conserved sequence Met-Lys-Arg-Gln-Lys (amino acid numbers 70 to 74 of zP450RAI) can be used as a probe to obtain corresponding proteins from cDNA libraries of other species. The amino acid identity between the human and mouse P450RAI sequences is greater than 93%.

As shown above, RA-induced expression of a protein by the cells described herein involves a regulatory sequence which is located upstream of the coding sequence of DNA that it controls. As it occurs in nature, the protein is P450RAI, whether in cells of the zebrafish, human, mouse or other organism. Such a cell can be modified by incorporating DNA encoding a different protein into the region of the gene which encodes P450RAI, as exemplified above with DNA encoding luciferase. An aspect of the present invention is thus a regulatory DNA sequence, responsive to the presence of RA, operably linked to a protein-encoding sequence and incorporated into a conventional genetically engineered protein-producing cell. Production of the desired protein can be induced by exposure of the cell to RA.

Antisense nucleic acids or oligonucleotides (RNA or preferably DNA) that inhibit cellular RA-induced P450RAI production can be used to inhibit metabolism of RA by P450RAI [Monia, 1996]. Antisense oligonucleotides, typically 15 to 20 bases long, bind to the sense mRNA or pre mRNA region coding for the protein of interest, which can inhibit translation of the bound mRNA to protein. The cDNA sequence encoding hP450RAI can thus be used to design a series of oligonucleotides which together span a large portion, or even the entire cDNA sequence. These oligonucleotides can be tested to determine which provides the greatest inhibitory effect on the expression of the protein [Stewart, 1996]. This can be done by exposing cells to the various oligonucleotides and measuring subsequent changes in hP450RAI activity or by using antibodies to screen for inhibition of P450RAI synthesis. The most suitable mRNA target sites include 5'- and 3'-untranslated regions as well as the initiation codon. Other regions might be found to be more or less effective. Alternatively, an antisense nucleic acid or oligonucleotide may bind to P450RAI DNA coding or regulatory sequences.

Rather than reducing RA metabolism by inhibiting P450RAI gene expression at the nucleic acid level, activity of the P450RAI protein may be directly inhibited by binding to an agent, such as, for example, a suitable small molecule or a monoclonal antibody.

The present invention thus includes a method of screening drugs for their effect on activity of a retinoic acid inducible protein. The method includes exposing the protein to a prospective inhibitor drug and determining the effect on protein activity. The measured activity might be hydroxylation of a retinoid, particularly all-*trans* retinoic acid, or hydroxylation of a retinoic acid, particularly all-*trans* retinoic acid, at the 4 position of the  $\beta$ -ionone ring thereof. For screening drugs for use in humans, hP450RAI itself is particularly useful for testing the effectiveness of such drugs. Prospective drugs could also be tested for inhibition of the activity of other P450 cytochromes, which are desired not to be inhibited. In this way, drugs which selectively inhibit hP450RAI over other P450s could be identified.

Another system for screening for potential inhibitors of a P450RAI protein includes a stably transfected cell line having incorporated therein DNA of a reporter gene (e.g.,  $\beta$ -galactosidase, firefly luciferase, or the like) and of the P450RAI, in which expression of both genes is inducible by exposure of the cells to RA. Expression of the reporter gene provides a measure of the induction of the expression system and therefore provides an indication of the amount of RA present. Exposure of the cells to RA leads to RA metabolism and, with time, such metabolism leads to a decrease in the degree of induction which is indicated by the reporter protein. Exposure of the cells to RA in the presence of an agent that inhibits P450RAI metabolism of RA results in decreased RA metabolism, whereas exposure of the cells to RA in the presence of an agent that does not inhibit P450RAI metabolism of RA has no effect on RA metabolism. A comparison of expression of the reporter gene in the presence of RA alone and in the presence of both RA and a potential inhibitory drug thus gives a measure of the effectiveness of the drug in inhibiting metabolism of RA by the P450RAI protein.

One system for screening for potential inhibitors of a P450RAI protein includes a cell line in which the endogenous P450RAI gene is not present or not functional or not expressed. In this cell line, a cytochrome P450RAI expression vector and an RA-inducible reporter gene are incorporated such that exposure of the cell line to RA results in metabolism of RA by the expressed P450RAI protein and a degree of induction of the reporter gene based on remaining active RA. The addition of an inhibitor of P450RAI will decrease the rate of metabolism/degradation of RA and therefore increase the activation/induction of the RA sensitive reporter gene.

Given the high degree of conservation of the promoter regions of the mouse, human and zebrafish P450RAI promoter regions, it is likely that RA regulates P450RAI expression through a transcriptional mechanism involving the RARE conserved in the promoters of all three species. This is supported by studies which show the rapid and RA-dependent expression of P450RAI in a number of cell lines. Since P450RAI message is induced so strongly a reporter gene may be a useful indicator of RA activity *in vivo* as well as *in vitro*. Thus, the P450RAI promoter linked to a reporter gene provides a tool for screening retinoids or other compounds which have the ability to block or inhibit P450RAI induction. For example, the P450RAI reporter gene could be stably or transiently introduced into a cell line such that when the cells are exposed to a certain level of retinoid or other agent, the concentration will be reflected in reporter gene activity. Such transfection assays can be carried out in a manner similar to those described by Petkovich *et al.*, for example [Petkovich, 1987; Ohno, 1994].

The invention thus provides a system for screening potential inhibitors of RA catabolism by a P450RAI protein. The system includes a transfected cell line having incorporated therein DNA of a reporter gene, for example the luciferase gene exemplified above, in which expression of the reporter gene is inducible by exposure of the cells to RA. In this system, the P450RAI gene is omitted, that is the reporter gene is under the control of the native promoter for the P450RAI gene. Expression of the reporter gene provides a measure of

the induction of the expression system and therefore provides an indication of the amount of mRNA produced in response to exposure of the cells to RA. Exposure of the cells to RA in the presence of an agent that inhibits induction of the expression system indicates that the agent is a potential inhibitor of RA catabolism, i.e., provides a measure of the effectiveness of the agent as a drug in inhibiting the expression of P450RAI gene and thus metabolism of RA.

There is the possibility that cellular retinoic acid-binding protein (CRABP) [Adamson, 1993] is involved in binding of a retinoid substrate to a P450RAI protein of the present invention. The effect of the presence of CRABP, derivatives, synthetic fragments or analogs thereof could thus be determined according to screening methods of the present invention; effectiveness of such agents in enhancing RA metabolism can also be determined.

It will of course be understood, without the intention of being limited thereby, that a variety of substitutions of amino acids is possible while preserving the structure responsible for retinoid metabolizing activity of the proteins disclosed herein. Conservative substitutions are described in the patent literature, as for example, in United States Patent No. 5,264,558. It is thus expected, for example, that interchange among non-polar aliphatic neutral amino acids, glycine, alanine, proline, valine and isoleucine, would be possible. Likewise, substitutions among the polar aliphatic neutral amino acids, serine, threonine, methionine, asparagine and glutamine could possibly be made. Substitutions among the charged acidic amino acids, aspartic acid and glutamic acid, could probably be made, as could substitutions among the charged basic amino acids, lysine and arginine. Substitutions among the aromatic amino acids, including phenylalanine, histidine, tryptophan and tyrosine would also likely be possible. These sorts of substitutions and interchanges are well known to those skilled in the art. Other substitutions might well be possible. Of course, it would also be expected that the greater the percentage of homology, i.e., sequence similarity, of a variant protein with a naturally occurring protein, the greater the retention of metabolic activity. Of course, as protein variants having the activity of P450RAI as described herein are intended to be within the scope of this invention, so are nucleic acids encoding such variants.

A further advantage may be obtained through chimeric forms of the protein, as known in the art. A DNA sequence encoding the entire protein, or a portion of the protein, could thus be linked, for example, with a sequence coding for the C-terminal portion of *E. coli*  $\beta$ -galactosidase to produce a fusion protein. GST-CYPRAI fusion proteins are described in the above examples. An expression system for human respiratory syncytial virus glycoproteins F and G is described in United States Patent No. 5,288,630 issued February 22, 1994 and references cited therein, for example.

A recombinant expression vector of the invention can be a plasmid, as described above. The recombinant expression vector of the invention further can be a virus, or portion thereof, which allows for expression of a nucleic acid introduced into the viral nucleic acid. For example, replication defective retroviruses, adenoviruses and adeno-associated viruses can be used.

The invention provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression, by transcription of the DNA molecule, of an RNA molecule which is antisense to the nucleotide sequence of SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:31. Regulatory sequences operatively linked to the antisense nucleic acid can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance a viral promoter and/or enhancer, or other regulatory sequences can be chosen which direct tissue or cell type specific expression of antisense RNA.

The recombinant expression vectors of the invention can be used to make a transformant host cell including the recombinant expression vector. The term "transformant host cell" is intended to include prokaryotic and eukaryotic cells which have been transformed or transfected with a recombinant expression vector of the invention. The terms "transformed with", "transfected with", "transformation" and "transfection" are intended to encompass introduction of nucleic acid (e.g. a vector) into a cell by one of many possible techniques known in the art. Prokaryotic cells can be transformed with nucleic acid by, for example, electroporation or calcium-chloride mediated transformation. Nucleic acid can be introduced into mammalian cells via conventional techniques such as calcium phosphate or calcium chloride coprecipitation, DEAE-dextran-mediated transfection, lipofection, electroporation or microinjection. Suitable methods for transforming and transfecting host cells are known [Sambrook, 1989].

The number of host cells transformed with a recombinant expression vector of the invention by techniques such as those described above will depend upon the type of recombinant expression vector used and the type of transformation technique used. Plasmid vectors introduced into mammalian cells are integrated into host cell DNA at only a low frequency. In order to identify these integrants, a gene that contains a selectable marker (e.g. resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those which confer resistance to certain drugs, such as G418 and hygromycin. Selectable markers can be introduced on a separate plasmid from the nucleic acid of interest or, preferably, are introduced on the same plasmid. Host cells transformed with one or more recombinant expression vectors containing a nucleic acid of the invention and a gene for a selectable marker can be identified by selecting for cells using the selectable marker. For example, if the selectable marker encodes a gene conferring neomycin resistance (such as pRc/CMV), transformant cells can be selected with G418. Cells that have incorporated the selectable marker gene will survive, while the other cells die.

Certain nucleic acids of the invention encode proteins which "have biological activity of a cytochrome P450RAI". The biological activity of a cytochrome P450RAI is the ability to oxidize a retinoid. Such activity can be tested for as described above.

The invention provides purified proteins having biological activity of P450RAI. The terms "isolated" and "purified" each refer to a protein substantially free of cellular material

or culture medium when produced by recombinant DNA techniques, or chemical precursors or other chemicals when chemically synthesized. In certain preferred embodiments, the protein having biological activity of P450RAI comprises an amino acid sequence identified as SEQ ID NO:2 or SEQ ID NO:4 or SEQ ID NO:32. Alternatively, preferred proteins encoded by a nucleic acid comprising the nucleotide sequence identified as SEQ ID NO:3 or SEQ ID NO:5 or SEQ ID NO:31, as defined above, are encompassed by the invention. Furthermore, proteins having biological activity of P450RAI that are encoded by nucleic acids which hybridize under stringent conditions, as discussed above, to a nucleic acid comprising a nucleotide sequence identified as SEQ ID NO:3 or SEQ ID NO:5 or SEQ ID NO:31 are encompassed by the invention.

10 P450RAIs of the invention can be obtained by expression in a suitable host cell using techniques known in the art. Suitable host cells include prokaryotic or eukaryotic organisms or cell lines, for example, yeast, *E. coli*, insect cells and COS 1 cells. The recombinant expression vectors of the invention, described above, can be used to express a protein having P450RAI activity in a host cell in order to isolate the protein. The invention provides a method of

15 preparing an purified protein of the invention comprising introducing into a host cell a recombinant nucleic acid encoding the protein, allowing the protein to be expressed in the host cell and isolating and purifying the protein. Preferably, the recombinant nucleic acid is a recombinant expression vector. Proteins can be isolated from a host cell expressing the protein and purified according to standard procedures of the art, including ammonium sulfate

20 precipitation, column chromatography (e.g. ion exchange, gel filtration, affinity chromatography, etc.), electrophoresis, and ultimately, crystallization [see generally, "Enzyme Purification and Related Techniques", *Methods in Enzymology*, 22, 233-577 (1971)].

Alternatively, the protein or parts thereof can be prepared by chemical synthesis using techniques well known in the chemistry of proteins such as solid phase

25 synthesis [Merrifield, 1964], or synthesis in homogeneous solution [Houbenwyl, 1987].

The protein of the invention, or portions thereof, can be used to prepare antibodies specific for the proteins. Antibodies can be prepared which bind to a distinct epitope in an unconserved region of a particular protein. An unconserved region of the protein is one which does not have substantial sequence homology to other proteins, for example other

30 members of the P450 family of cytochromes. Conventional methods can be used to prepare the antibodies. For example, by using a peptide of a P450RAI protein, polyclonal antisera or monoclonal antibodies can be made using standard methods. As demonstrated in Example 16, a mammal, (e.g. a mouse, hamster, or rabbit) can be immunized with an immunogenic form of the peptide which elicits an antibody response in the mammal. Techniques for

35 conferring immunogenicity on a peptide include conjugation to carriers or other techniques well known in the art. For example, the peptide can be administered in the presence of adjuvant. The progress of immunization can be monitored by detection of antibody titers in plasma or serum. Standard ELISA or other immunoassay can be used to assess the levels of antibodies. Following immunization, antisera can be obtained and, if desired, polyclonal antibodies isolated

40 from the sera.

To produce monoclonal antibodies, antibody producing cells (lymphocytes) can be harvested from an immunized animal and fused with myeloma cells by standard somatic cell fusion procedures, thus immortalizing these cells and yielding hybridoma cells. Such techniques are well known in the art. For example, the hybridoma technique originally developed by Kohler and Milstein [Kohler, 1975] as well as other techniques such as the human B-cell hybridoma technique [Kozbor, 1983], the EBV-hybridoma technique to produce human monoclonal antibodies [Cole, 1985], and screening of combinatorial antibody libraries [Huse, 1989]. Hybridoma cells can be screened immunochemically for production of antibodies specifically reactive with the peptide, and monoclonal antibodies isolated.

The term antibody as used herein is intended to include fragments thereof which are also specifically reactive with a protein having the biological activity of P450RAI, or a peptide fragment thereof. Antibodies can be fragmented using conventional techniques and the fragments screened for utility in the same manner as described above for whole antibodies. For example, F(ab')<sub>2</sub> fragments can be generated by treating antibody with pepsin. The resulting F(ab')<sub>2</sub> fragment can be treated to reduce disulfide bridges to produce Fab' fragments.

It is also known in the art to make chimeric antibody molecules with human constant regions. See, for example, Morrison *et al.*, Takeda *et al.*, Cabilly *et al.*, Boss *et al.*, Tanaguchi *et al.*, Teng *et al.* [Morrison, 1985; Takeda, 1985; Cabilly; Boss; Tanaguchi; Teng, 1982], European Patent Publication 0173494, United Kingdom Patent GB 2177096B, PCT Publication WO92/06193 and EP 0239400. It is expected that such chimeric antibodies would be less immunogenic in a human subject than the corresponding non-chimeric antibody.

Another method of generating specific antibodies, or antibody fragments, reactive against protein having the biological activity of a P450RAI, or a peptide fragment thereof, is to screen expression libraries encoding immunoglobulin genes, or portions thereof, expressed in bacteria, with peptides produced from the nucleic acid molecules of the present invention. For example, complete Fab fragments, VH regions and FV regions can be expressed in bacteria using phage expression libraries. See for example Ward *et al.*, Huse *et al.*, and McCafferty *et al.* [Ward, 1989; Huse, 1989; McCafferty, 1990]. Screening such libraries with, for example, a P450RAI peptide can identify immunoglobulin fragments reactive with P450RAI. Alternatively, the SCID-hu mouse developed by Genpharm can be used to produce antibodies, or fragments thereof.

The polyclonal, monoclonal or chimeric monoclonal antibodies can be used to detect the proteins of the invention, portions thereof or closely related isoforms in various biological materials, for example they can be used in an ELISA, radioimmunoassay or histochemical tests. Thus, the antibodies can be used to quantify the amount of a P450RAI protein of the invention, portions thereof or closely related isoforms in a sample in order to determine the role of P450RAI proteins in particular cellular events or pathological states. Using methods described hereinbefore, polyclonal, monoclonal antibodies, or chimeric monoclonal antibodies can be raised to nonconserved regions of P450RAI and used to distinguish a particular P450RAI from other proteins.

The polyclonal or monoclonal antibodies can be coupled to a detectable substance or reporter system. The term "coupled" is used to mean that the detectable substance is physically linked to the antibody. Suitable detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase,  $\beta$ -galactosidase, and acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride and phycoerythrin; an example of a luminescent material includes luminol; and examples of suitable radioactive material include  $^{125}\text{I}$ ,  $^{131}\text{I}$ ,  $^{35}\text{S}$  and  $^3\text{H}$ . In a preferred embodiment, the reporter system allows quantitation of the amount of protein (antigen) present.

Such an antibody-linked reporter system could be used in a method for determining whether a fluid or tissue sample of a subject contains a deficient amount or an excessive amount of the protein. Given a normal threshold concentration of such a protein for a given type of subject, test kits could thus be developed.

The present invention allows the skilled artisan to prepare bispecific antibodies and tetrameric antibody complexes. Bispecific antibodies can be prepared by forming hybrid hybridomas [Staerz, 1986a &b].

The present invention includes three types of compounds related to retinoids: those that inhibit enzymatic activity of P450RAI, thereby inhibiting metabolism of RA; those retinoids that evade metabolism by P450RAI, for example Am580, shown above; and those compounds that repress induction of P450RAI gene expression, for example 4-HPR, as shown above.

Compositions of the invention are administered to subjects in a biologically compatible form suitable for pharmaceutical administration *in vivo*. By "biologically compatible from suitable for administration *in vivo*" is meant a form of the composition to be administered in which any toxic effects are outweighed by the therapeutic effects of the composition. The term "subject" is intended to include living organisms in which a desired therapeutic response can be elicited, e.g. mammals. Examples of subjects include human, dogs, cats, mice, rats and transgenic species thereof. Administration of a therapeutically active amount of the therapeutic compositions of the present invention is defined as an amount effective, at dosages and for periods of time necessary to achieve the desired result. For example, a therapeutically active amount of a compound that inhibits catabolism of RA by a P450RAI protein may vary according to factors such as the disease state, age, sex, and weight of the individual, as well as target tissue and mode of delivery. Dosage regimes may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation.

Compounds of the present invention, such as those that are found to inhibit metabolism of RA by P450RAI enzymes and that are useful as anticancer agents and in the



treatment, amelioration, or prevention of skin disorders for which retinoic acid is useful, for example, may be used topically. In this regard they may be included in compositions for therapy in animals, including humans, for premalignant epithelial cell lesions, as a prophylaxis against tumor promotion in epithelial cells and treatment for dermatoses such as ichthyoses, follicular disorders, benign epithelial disorders, and other proliferative skin diseases, such as acne, psoriasis, eczema, atopic dermatitis, nonspecific dermatitis and the like.

Topical compositions are usually formulated with a pharmaceutically acceptable carrier in liquid, semi-solid or solid form. A pharmaceutically acceptable carrier is a material that is nontoxic and generally inert and does not affect the functionality of the active ingredients adversely. Such materials are well known and include those materials sometimes referred to as diluents or vehicles (excipients) in the pharmaceutical formulation art. The carrier may be organic or inorganic in nature. Examples of pharmaceutically acceptable carriers are water, gelatin, lactose, starch, mineral oil, cocoa butter, dextrose, sucrose, sorbitol, mannitol, gum, acacia, alginates, cellulose, talc, magnesium stearate, polyoxyethylene sorbitan monolaurate, and other commonly used pharmaceutical carriers. In addition to an active ingredient and carrier, the formulation may contain minor amounts of additives such as flavoring agents, coloring agents, thickening or gelling agents, emulsifiers, wetting agents, buffers, stabilizers, and preservatives such as antioxidants.

Certain compositions may be administered enterally. For oral administration, suitable forms are, for example, tablets, pills, syrups, suspensions, emulsions, solutions, powders and granules.

As anti-tumor agents or as part of an anti-tumor formulation, for example, compounds of the present invention can be used in a similar manner to retinoids used for treating various tumours, such as all-*trans* retinoic acid. The dose to be administered, whether a single dose, multiple does or daily dose, will of course vary with the particular compound employed because of the varying potency of the active ingredient, the chosen route of administration, the size of the recipient, the type of tumor, and the nature of the patient's condition. The dosage to be administered is not subject to definite bounds, but it will usually be an effective amount, or the equivalent on a molar basis of the pharmacologically active free form produced from a dosage formulation upon the metabolic release of the active drug to achieve its desired pharmacological and physiological effects. An oncologist skilled in the art of cancer treatment will be able to ascertain without undue experimentation, appropriate protocols for the effective administration of the compounds of this present invention.

Nucleic acids which encode proteins having biological activity of a P450RA1 protein can be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. A transgenic animal (e.g., a mouse) is an animal having cells that contain a transgene, which transgene was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A transgene is a DNA which is integrated into the genome of a cell from which a transgenic animal develops. In one embodiment, a human P450RA1 cDNA, comprising the nucleotide

sequence shown in SEQ ID NO:5, or an appropriate variant or subsequence thereof, can be used to generate transgenic animals that contain cells which express human P450RAI protein. Methods for generating transgenic animals, particularly animals such as mice, have become conventional in the art are described, for example, in U.S. Patent Nos. 4,736,866 and

5 4,870,009. In a preferred embodiment, plasmids containing recombinant molecules of the invention are microinjected into mouse embryos. In particular, the plasmids are microinjected into the male pronuclei of fertilized one-cell mouse eggs; the injected eggs are transferred to pseudo-pregnant foster females; and, the eggs in the foster females are allowed to develop to term. [Hogan, 1986]. Alternatively, an embryonal stem cell line can be transfected with an  
10 expression vector comprising nucleic acid encoding a protein having P450RAI activity, and cells containing the nucleic acid can be used to form aggregation chimeras with embryos from a suitable recipient mouse strain. The chimeric embryos can then be implanted into a suitable pseudopregnant female mouse of the appropriate strain and the embryo brought to term. Progeny harboring the transfected DNA in their germ cells can be used to breed uniformly  
15 transgenic mice.

Typically, particular cells would be targeted for P450RAI transgene incorporation by use of tissue specific enhancers operatively linked to the P450RAI encoding gene. For example, promoters and/or enhancers which direct expression of a gene to which they are operatively linked preferentially in cardiac muscle cells can be used to create a  
20 transgenic animal which expresses a P450RAI protein preferentially in cardiac muscle tissue. Examples of suitable promoters and enhancers include those which regulate the expression of the genes for cardiac myosin and cardiac actin. Transgenic animals that include a copy of an P450RAI transgene introduced into the germ line of the animal at an embryonic stage can also be used to examine the effect of increased P450RAI expression in various tissues.

25 The pattern and extent of expression of a recombinant molecule of the invention in a transgenic mouse is facilitated by fusing a reporter gene to the recombinant molecule such that both genes are co-transcribed to form a polycistronic mRNA. The reporter gene can be introduced into the recombinant molecule using conventional methods such as those described in Sambrook *et al.*, [Sambrook, 1989]. Efficient expression of both cistrons of  
30 the polycistronic mRNA encoding the protein of the invention and the reporter protein can be achieved by inclusion of a known internal translational initiation sequence such as that present in poliovirus mRNA. The reporter gene should be under the control of the regulatory sequence of the recombinant molecule of the invention and the pattern and extent of expression of the gene encoding a protein of the invention can accordingly be determined by assaying for the  
35 phenotype of the reporter gene. Preferably the reporter gene codes for a phenotype not displayed by the host cell and the phenotype can be assayed quantitatively. Examples of suitable reporter genes include lacZ ( $\beta$ -galactosidase), neo (neomycin phosphotransferase), CAT (chloramphenicol acetyltransferase) dhfr (dihydrofolate reductase), aphIV (hygromycin phosphotransferase), lux (luciferase), uidA ( $\beta$ -glucuronidase). Preferably, the reporter gene is  
40 lacZ which codes for  $\beta$ -galactosidase.  $\beta$ -galactosidase can be assayed using the lactose

analogue X-gal (5-bromo-4-chloro-3-indolyl-b-D-galactopyranoside) which is broken down by  $\beta$ -galactosidase to a product that is blue in color [Old].

Although experimental animals used in the preferred embodiment disclosed are mice, the invention should not be limited thereto. It can be desirable to use other species  
5 such as, for example, rats, hamsters, rabbits and sheep.

The transgenic animals of the invention can be used to investigate the molecular basis of RA metabolism. The transgenic animals of the invention can also be used to test substances for the ability to prevent, slow or enhance RA metabolism. A transgenic animal can be treated with the substance in parallel with an untreated control transgenic  
10 animal.

Cells from the transgenic animals of the invention can be cultured using standard tissue culture techniques. In particular, cells carrying the recombinant molecule of the invention can be cultured and used to test substances for the ability to prevent, slow or enhance RA metabolism.

15 Additionally, the non-human homologues of genes encoding proteins having P450RAI activity can be used to construct a "knock out" animal which has a defective or altered P450RAI gene. For example, with established techniques, a portion of murine genomic P450RAI DNA (e.g., an exon), can be deleted or replaced with another gene, such as a gene encoding a selectable marker which can be used to monitor integration. The altered P450RAI  
20 DNA can then be transfected into an embryonal stem cell line. The altered P450RAI DNA will homologously recombine with the endogenous P450RAI gene in certain cells and clones containing the altered gene can be selected. Cells containing the altered gene are injected into a blastocyst of an animal, such as a mouse, to form aggregation chimeras as described for transgenic animals. Chimeric embryos are implanted as described above. Transmission of the  
25 altered gene into the germline of a resultant animal can be confirmed using standard techniques and the animal can be used to breed animals having an altered P450RAI gene in every cell [Lemoine, 1996]. Accordingly, a knockout animal can be made which cannot express a functional P450RAI protein. Such a knockout animal can be used, for example, to test the effectiveness of an agent in the absence of a P450RAI protein.

30 The antisense nucleic acids and oligonucleotides of the invention are useful for inhibiting expression of nucleic acids (e.g. mRNAs) encoding proteins having P450RAI activity. Since proteins having P450RAI activity are associated with metabolism of agents which can act on the cell, e.g., RA, decreasing expression of such proteins can increase sensitivity of the cell to such agents. Antisense nucleic acids can be introduced into a drug resistant cell in culture to  
35 inhibit P450RAI expression. One or more antisense nucleic acids, such as oligonucleotides, can be added to cells in culture media, typically, for example, at 200  $\mu$ g/ml.

The antisense nucleic acids of the invention, or oligonucleotides thereof, can thus be used in gene therapy to correct or prevent retinoic acid or other retinoid resistance in a subject. For example, antisense sequences can be used to render retinoic acid or other  
40 retinoid resistant malignant cells sensitive to chemotherapeutic agents. Administration of

antisense nucleic acids to a subject may be most effective when the antisense nucleic acid is contained in a recombinant expression vector which allows for continuous production of antisense RNA. Recombinant molecules comprising an antisense nucleic acid or oligonucleotide thereof, can be directly introduced into tissues, including lung tissue *in vivo*,  
5 using delivery vehicles such as liposomes, retroviral vectors, adenoviral vectors and DNA virus vectors. A delivery vehicle can be chosen which can be targeted to a cell of interest in the subject (e.g. a retinoid resistant tumor cell). Antisense nucleic acids can also be introduced into isolated cells, such as those of the haematopoietic system, *ex vivo* using viral vectors or physical techniques such as microinjection and electroporation or chemical methods such as  
10 coprecipitation and incorporation of DNA into liposomes, and such cells can be returned to the donor. Recombinant molecules can be delivered in the form of an aerosol or by lavage.

Accordingly, the invention provides a method for inhibiting retinoic acid or other retinoid resistance of a resistant cell by introducing into the resistant cell a nucleic acid which is antisense to a nucleic acid which encodes the protein identified as SEQ ID NO:2, SEQ ID  
15 NO:32, or particularly, the in case of human cells SEQ ID NO:4.

The nucleic acids of the invention can further be used to design ribozymes which are capable of cleaving a single-stranded nucleic acid encoding a protein having P450RAI activity, such as an mRNA. A catalytic RNA (ribozyme) having ribonuclease activity can be designed which has specificity for a P450RAI-encoding mRNA based upon the  
20 sequence of a nucleic acid of the invention. For example, a derivative of a *Tetrahymena* L-19IVS RNA can be constructed in which the base sequence of the active site is complementary to the base sequence to be cleaved in a P450RAI-encoding mRNA. [Cech a and b]. Alternatively, a nucleic acid of the invention could be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules [Bartel, 1993].

25 The isolated nucleic acids and antisense nucleic acids of the invention can be used to construct recombinant expression vectors as described previously. These recombinant expression vectors are then useful for making transformant host cells containing the recombinant expression vectors, for expressing protein encoded by the nucleic acids of the invention, and for isolating proteins of the invention as described previously. The isolated  
30 nucleic acids and antisense nucleic acids of the invention can also be used to construct transgenic and knockout animals as described previously.

The isolated proteins of the invention are useful for making antibodies reactive against proteins having P450RAI activity, as described previously. Alternatively, the antibodies of the invention can be used to isolate a protein of the invention by standard immunoaffinity  
35 techniques. Furthermore, the antibodies of the invention, including bispecific antibodies are useful for diagnostic purposes.

Molecules which bind to a protein comprising an amino acid sequence shown in SEQ ID NO:4 can also be used in a method for killing a cell which expresses the protein, wherein the cell takes up the molecule. Preferably, the cell is a tumor cell. Destruction of such  
40 cells can be accomplished by labeling the molecule with a substance having toxic or

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Those skilled in the art will know, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

## References

Particulars of references cited above are given below. All of the listed references are incorporated herein by reference.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Petkovich, P. Martin, White, Jay A.,  
Beckett, Barbara R., Jones, Glenville
- (ii) TITLE OF INVENTION: Retinoid Metabolizing Protein
- (iii) NUMBER OF SEQUENCES: 35
- (iv) CORRESPONDENCE ADDRESS:  
(A) ADDRESSEE: Blake, Cassels & Graydon  
(B) STREET: Box 25, Commerce Court West  
(C) CITY: Toronto  
(D) PROVINCE: Ontario  
(E) COUNTRY: Canada  
(F) ZIP: M5L 1A9
- (v) COMPUTER READABLE FORM:  
(A) MEDIUM TYPE: Diskette, 3 1/2 inch, 1.4 Mb storage  
(B) COMPUTER: COMPAQ, IBM PC compatible  
(C) OPERATING SYSTEM: MS-DOS 5.1  
(D) SOFTWARE: WORD PERFECT
- (vi) CURRENT APPLICATION DATA:  
(A) APPLICATION NUMBER:  
(B) FILING DATE:
- (vii) PRIOR APPLICATION DATA:  
(A) APPLICATION NUMBERS: 08/667,546; 08/724,466  
(B) FILING DATE: June 21, 1996; October 1, 1996
- (viii) ATTORNEY/AGENT INFORMATION:  
(A) NAME: Hunt, John C.  
(B) REGISTRATION NUMBER: 36,424  
(C) REFERENCE/DOCKET NUMBER: 50767/00010
- (ix) TELECOMMUNICATION INFORMATION:  
(A) TELEPHONE: (416) 863-4344  
(B) TELEFAX: (416) 863-2653

(2) INFORMATION FOR SEQ ID NO:1

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 337 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1

TGCCAGTGGA CAATCTCCCT ACCAAATTCA CTAGTTATGT CCAGAAATTA GCCTAAACCG	60
GAGCCTTTGT ACATATGTTT TTATTTTAGA TGAAGTGTGA TGTATTGGAT ATTTTCTAAT	120
TTGTTTATAT AAAGCAGATG TGTATATAAG TCTATGCGAA GAAGCGAAAA CGAGGGCACT	180
ACTTTCTCAT GGATCACTGT AATGCTACAG AGTGTCTGTG ATGTATATTT ATAATGTAGT	240
TGTGTCATAT AGCTTTTGTA CTGTATGCAA CTTATTTAAC TCGCTCTTTA TCTCATGGGT	300
TTTATTTAAT AAAACATGTT CTTACAAAAA AAAAAAA	337

(2) INFORMATION FOR SEQ ID NO:2

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 492 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2

Met	Gly	Leu	Tyr	Thr	Leu	Met	Val	Thr	Phe	Leu	Cys	Thr	Ile	Val	Leu	1	5	10	15
Pro	Val	Leu	Leu	Phe	Leu	Ala	Ala	Val	Lys	Leu	Trp	Glu	Met	Leu	Met	20	25	30	
Ile	Arg	Arg	Val	Asp	Pro	Asn	Cys	Arg	Ser	Pro	Leu	Pro	Pro	Gly	Thr	35	40	45	
Met	Gly	Leu	Pro	Phe	Ile	Gly	Glu	Thr	Leu	Gln	Leu	Ile	Leu	Gln	Arg	50	55	60	
Arg	Lys	Phe	Leu	Arg	Met	Lys	Arg	Gln	Lys	Tyr	Gly	Cys	Ile	Tyr	Lys	65	70	75	80
Thr	His	Leu	Phe	Gly	Asn	Pro	Thr	Val	Arg	Val	Met	Gly	Ala	Asp	Asn	85	90	95	
Val	Arg	Gln	Ile	Leu	Leu	Gly	Glu	His	Lys	Leu	Val	Ser	Val	Gln	Trp	100	105	110	
Pro	Ala	Ser	Val	Arg	Thr	Ile	Leu	Gly	Ser	Asp	Thr	Leu	Ser	Asn	Val	115	120	125	
His	Gly	Val	Gln	His	Lys	Asn	Lys	Lys	Lys	Ala	Ile	Met	Arg	Ala	Phe	130	135	140	
Ser	Arg	Asp	Ala	Leu	Glu	His	Tyr	Ile	Pro	Val	Ile	Gln	Gln	Glu	Val	145	150	155	160
Lys	Ser	Ala	Ile	Gln	Glu	Trp	Leu	Gln	Lys	Asp	Ser	Cys	Val	Leu	Val	165	170	175	
Tyr	Pro	Glu	Met	Lys	Lys	Leu	Met	Phe	Arg	Ile	Ala	Met	Arg	Ile	Leu	180	185	190	
Leu	Gly	Phe	Glu	Pro	Glu	Gln	Ile	Lys	Thr	Asp	Glu	Gln	Glu	Leu	Val	195	200	205	
Glu	Ala	Phe	Glu	Glu	Met	Ile	Lys	Asn	Leu	Phe	Ser	Leu	Pro	Ile	Asp	210	215	220	
Val	Pro	Phe	Ser	Gly	Leu	Tyr	Arg	Gly	Leu	Arg	Ala	Arg	Asn	Phe	Ile	225	230	235	240
His	Ser	Lys	Ile	Glu	Glu	Asn	Ile	Arg	Lys	Lys	Ile	Gln	Asp	Asp	Asp	245	250	255	
Asn	Glu	Asn	Glu	Gln	Lys	Tyr	Lys	Asp	Ala	Leu	Gln	Leu	Leu	Ile	Glu	260	265	270	
Asn	Ser	Arg	Arg	Ser	Asp	Glu	Pro	Phe	Ser	Leu	Gln	Ala	Met	Lys	Glu	275	280	285	
Ala	Ala	Thr	Glu	Leu	Leu	Phe	Gly	Gly	His	Glu	Thr	Thr	Ala	Ser	Thr	290	295	300	
Ala	Thr	Ser	Leu	Val	Met	Phe	Leu	Gly	Leu	Asn	Thr	Glu	Val	Val	Gln				

305		310		315		320
Lys Val Arg Glu	Glu Val Gln Glu Lys Val Glu Met Gly Met Tyr Thr					
	325			330		335
Pro Gly Lys Gly Leu Ser Met Glu Leu Leu Asp Gln Leu Lys Tyr Thr						
	340			345		350
Gly Cys Val Ile Lys Glu Thr Leu Arg Ile Asn Pro Pro Val Pro Gly						
	355			360		365
Gly Phe Arg Val Ala Leu Lys Thr Phe Glu Leu Asn Gly Tyr Gln Ile						
	370			375		380
Pro Lys Gly Trp Asn Val Ile Tyr Ser Ile Cys Asp Thr His Asp Val						
	385			390		395
Ala Asp Val Phe Pro Asn Lys Glu Glu Phe Gln Pro Glu Arg Phe Met						
	405			410		415
Ser Lys Gly Leu Glu Asp Gly Ser Arg Phe Asn Tyr Ile Pro Phe Gly						
	420			425		430
Gly Gly Ser Arg Met Cys Val Gly Lys Glu Phe Ala Lys Val Leu Leu						
	435			440		445
Lys Ile Phe Leu Val Glu Leu Thr Gln His Cys Asn Trp Ile Leu Ser						
	450			455		460
Asn Gly Pro Pro Thr Met Lys Thr Gly Pro Thr Ile Tyr Pro Val Asp						
	465			470		475
Asn Leu Pro Thr Lys Phe Thr Ser Tyr Val Arg Asn						
	485			490		

(2) INFORMATION FOR SEQ ID NO:3

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1850 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3

TGTCGCCGTT GCTGTCGGTT GCTGTCGGAC GCTGTCTCCT CTCCAGAAGC TTGTTTTTCG	60
TTTTGGCGAT CAGTTGCGCG CTTCAAC ATG GGG CTG TAC ACC CTT ATG GTC ACC	114
Met Gly Leu Tyr Thr Leu Met Val Thr	
1 5	
TTT CTC TGC ACC ATC GTG CTA CCC GTT TTA CTC TTT CTC GCC GCG GTG	162
Phe Leu Cys Thr Ile Val Leu Pro Val Leu Leu Phe Leu Ala Ala Val	
10 15 20 25	
AAG TTG TGG GAG ATG TTA ATG ATC CGA CGA GTC GAT CCG AAC TGC AGA	210
Lys Leu Trp Glu Met Leu Met Ile Arg Arg Val Asp Pro Asn Cys Arg	
30 35 40	
AGT CCT CTA CCG CCA GGT ACC ATG GGC TTG CCG TTC ATT GGA GAA ACG	258
Ser Pro Leu Pro Pro Gly Thr Met Gly Leu Pro Phe Ile Gly Glu Thr	
45 50 55	
CTC CAG CTG ATC CTC CAG AGA AGG AAG TTT CTG CGC ATG AAA CGG CAG	306
Leu Gln Leu Ile Leu Gln Arg Arg Lys Phe Leu Arg Met Lys Arg Gln	
60 65 70	

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AAA	TAC	GGG	TGC	ATC	TAC	AAG	ACG	CAC	CTC	TTC	GGG	AAC	CCG	ACT	GTC	354
Lys	Tyr	Gly	Cys	Ile	Tyr	Lys	Thr	His	Leu	Phe	Gly	Asn	Pro	Thr	Val	
	75					80					85					
AGG	GTG	ATG	GGA	GCT	GAT	AAT	GTG	AGG	CAG	ATT	CTG	CTG	GGC	GAA	CAC	402
Arg	Val	Met	Gly	Ala	Asp	Asn	Val	Arg	Gln	Ile	Leu	Leu	Gly	Glu	His	
	90				95					100					105	
AAG	CTG	GTG	TCT	GTT	CAG	TGG	CCA	GCA	TCA	GTG	AGA	ACC	ATC	CTG	GGC	450
Lys	Leu	Val	Ser	Val	Gln	Trp	Pro	Ala	Ser	Val	Arg	Thr	Ile	Leu	Gly	
				110					115					120		
TCT	GAC	ACC	CTC	TCC	AAT	GTC	CAT	GGA	GTT	CAA	CAC	AAA	AAC	AAG	AAA	498
Ser	Asp	Thr	Leu	Ser	Asn	Val	His	Gly	Val	Gln	His	Lys	Asn	Lys	Lys	
			125					130					135			
AAG	GCC	ATT	ATG	AGG	GCG	TTC	TCT	CGA	GAT	GCT	CTG	GAG	CAC	TAC	ATT	546
Lys	Ala	Ile	Met	Arg	Ala	Phe	Ser	Arg	Asp	Ala	Leu	Glu	His	Tyr	Ile	
	140						145					150				
CCC	GTG	ATC	CAG	CAG	GAG	GTG	AAG	AGC	GCC	ATA	CAG	GAA	TGG	CTG	CAA	594
Pro	Val	Ile	Gln	Gln	Glu	Val	Lys	Ser	Ala	Ile	Gln	Glu	Trp	Leu	Gln	
	155					160						165				
AAA	GAC	TCC	TGC	GTG	CTG	GTT	TAT	CCA	GAA	ATG	AAG	AAA	CTC	ATG	TTT	642
Lys	Asp	Ser	Cys	Val	Leu	Val	Tyr	Pro	Glu	Met	Lys	Lys	Leu	Met	Phe	
	170				175					180					185	
CGG	ATA	GCT	ATG	AGA	ATC	CTG	CTT	GGT	TTT	GAA	CCA	GAG	CAA	ATA	AAG	690
Arg	Ile	Ala	Met	Arg	Ile	Leu	Leu	Gly	Phe	Glu	Pro	Glu	Gln	Ile	Lys	
			190						195					200		
ACG	GAC	GAG	CAA	GAA	CTG	GTG	GAA	GCT	TTT	GAG	GAA	ATG	ATC	AAA	AAC	738
Thr	Asp	Glu	Gln	Glu	Leu	Val	Glu	Ala	Phe	Glu	Glu	Met	Ile	Lys	Asn	
			205					210					215			
TTG	TTC	TCC	TTG	CCA	ATC	GAC	GTT	CCT	TTC	AGT	GGT	CTG	TAC	AGG	GGT	786
Leu	Phe	Ser	Leu	Pro	Ile	Asp	Val	Pro	Phe	Ser	Gly	Leu	Tyr	Arg	Gly	
		220					225					230				
TTG	AGG	GCA	CGC	AAT	TTC	ATT	CAC	TCC	AAA	ATT	GAG	GAA	AAC	ATC	AGG	834
Leu	Arg	Ala	Arg	Asn	Phe	Ile	His	Ser	Lys	Ile	Glu	Glu	Asn	Ile	Arg	
	235				240						245					
AAG	AAA	ATT	CAA	GAT	GAC	GAC	AAT	GAA	AAC	GAA	CAG	AAA	TAC	AAA	GAC	882
Lys	Lys	Ile	Gln	Asp	Asp	Asp	Asn	Glu	Asn	Glu	gln	Lys	Tyr	Lys	Asp	
	250				255					260					265	
GCC	CTT	CAG	CTG	TTG	ATC	GAG	AAC	AGC	AGA	AGA	AGT	GAC	GAA	CCT	TTT	930
Ala	Leu	Gln	Leu	Leu	Ile	Glu	Asn	Ser	Arg	Arg	Ser	Asp	Glu	Pro	Phe	
			270						275					280		
AGT	TTG	CAG	GCG	ATG	AAA	GAA	GCA	GCT	ACA	GAG	CTT	CTA	TTT	GGA	GGT	978
Ser	Leu	Gln	Ala	Met	Lys	Glu	Ala	Ala	Thr	Glu	Leu	Leu	Phe	Gly	Gly	
			285					290					295			
CAT	GAA	ACC	ACC	GCC	AGC	ACT	GCA	ACC	TCA	CTT	GTC	ATG	TTT	CTG	GGT	1026
His	Glu	Thr	Thr	Ala	Ser	Thr	Ala	Thr	Ser	Leu	Val	Met	Phe	Leu	Gly	
		300					305					310				
CTG	AAC	ACA	GAA	GTG	GTG	CAG	AAG	GTC	AGA	GAG	GAG	GTT	CAG	GAG	AAG	1074
Leu	Asn	Thr	Glu	Val	Val	Gln	Lys	Val	Arg	Glu	Glu	Val	Gln	Glu	Lys	
	315					320					325					

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GTT GAA ATG GGC ATG TAT ACA CCT GGA AAG GGC TTG AGT ATG GAG CTG	1122
Val Glu Met Gly Met Tyr Thr Pro Gly Lys Gly Leu Ser Met Glu Leu	
330 335 340 345	
TTG GAC CAG CTG AAG TAC ACT GGA TGT GTG ATT AAA GAG ACT CTT AGA	1170
Leu Asp Gln Leu Lys Tyr Thr Gly Cys Val Ile Lys Glu Thr Leu Arg	
350 355 360	
ATC AAC CCT CCT GTT CCC GGA GGA TTC AGA GTC GCA CTC AAA ACC TTT	1218
Ile Asn Pro Pro Val Pro Gly Gly Phe Arg Val Ala Leu Lys Thr Phe	
365 370 375	
GAA TTG AAT GGT TAC CAA ATT CCT AAA GGA TGG AAC GTC ATT TAC AGC	1266
Glu Leu Asn Gly Tyr Gln Ile Pro Lys Gly Trp Asn Val Ile Tyr Ser	
380 385 390	
ATC TGT GAC ACG CAC GAT GTG GCC GAC GTC TTT CCA AAC AAA GAG GAG	1314
Ile Cys Asp Thr His Asp Val Ala Asp Val Phe Pro Asn Lys Glu Glu	
395 400 405	
TTC CAG CCG GAG AGA TTC ATG AGC AAA GGT CTG GAG GAC GGG TCC AGG	1362
Phe Gln Pro Glu Arg Phe Met Ser Lys Gly Leu Glu Asp Gly Ser Arg	
410 415 420 425	
TTT AAC TAC ATC CCC TTC GGA GGA GGA TCC AGG ATG TGT GTG GGC AAA	1410
Phe Asn Tyr Ile Pro Phe Gly Gly Gly Ser Arg Met Cys Val Gly Lys	
430 435 440	
GAG TTC GCC AAA GTG TTA CTC AAG ATC TTT TTA GTT GAG TTA ACG CAG	1458
Glu Phe Ala Lys Val Leu Leu Lys Ile Phe Leu Val Glu Leu Thr Gln	
445 450 455	
CAT TGC AAT TGG ATT CTC TCA AAC GGA CCC CCG ACA ATG AAA ACA GGC	1506
His Cys Asn Trp Ile Leu Ser Asn Gly Pro Pro Thr Met Lys Thr Gly	
460 465 470	
CCG ACT ATT TAC CCA GTG GAC AAT CTC CCT ACC AAA TTC ACT AGT TAT	1554
Pro Thr Ile Tyr Pro Val Asp Asn Leu Pro Thr Lys Phe Thr Ser Tyr	
475 480 485	
GTC AGA AAT TAGCCTAACC GGAGCTTTGT ACATATGTTT TTATTTTAGA	1603
Val Arg Asn	
490	
TGAACTGTGA TGTATTGGAT ATTTTCTATT TTGTTTATAT AAAGCAGATG TGTATATAAG	1663
TCTATGCGAG GAAGCGAAAA CGAGGGCACT ACTTTCTCAT GGATCACTGT AATGCTACAG	1723
AGTGTCTGTG ATGTATATTT ATAATGTAGT TGTGTTATAT AGCTTTTGTA CTGTATGCAA	1783
CTTATTTAAC TCGCTCTTTA TCTCATGGGT TTTATTTAAT AAAACATGTT CTTACAAAAA	1843
AAAAAAA	1850

(2) INFORMATION FOR SEQ ID NO:4

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 497 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4

Met Gly Leu Pro Ala Leu Leu Ala Ser Ala Leu Cys Thr Phe Val Leu
1 5 10 15



Pro Leu Leu Leu Phe Leu Ala Ala Ile Lys Leu Trp Asp Leu Tyr Cys  
20 25 30

Val Ser Gly Arg Asp Arg Ser Cys Ala Leu Pro Leu Pro Pro Gly Thr  
35 40 45

Met Gly Phe Pro Phe Phe Gly Glu Thr Leu Gln Met Val Leu Gln Arg  
50 55 60

Arg Lys Phe Leu Gln Met Lys Arg Arg Lys Tyr Gly Phe Ile Tyr Lys  
65 70 75 80

Thr His Leu Phe Gly Arg Pro Thr Val Arg Val Met Gly Ala Asp Asn  
85 90 95

Val Arg Arg Ile Leu Leu Gly Asp Asp Arg Leu Val Ser Val His Trp  
100 105 110

Pro Ala Ser Val Arg Thr Ile Leu Gly Ser Gly Cys Leu Ser Asn Leu  
115 120 125

His Asp Ser Ser His Lys Gln Arg Lys Lys Val Ile Met Arg Ala Phe  
130 135 140

Ser Arg Glu Ala Leu Glu Cys Tyr Val Pro Val Ile Thr Glu Glu Val  
145 150 155 160

Gly Ser Ser Leu Glu Gln Trp Leu Ser Cys Gly Glu Arg Gly Leu Leu  
165 170 175

Val Tyr Pro Glu Val Lys Arg Leu Met Phe Arg Ile Ala Met Arg Ile  
180 185 190

Leu Leu Gly Cys Glu Pro Gln Leu Ala Gly Asp Gly Asp Ser Glu Gln  
195 200 205

Gln Leu Val Glu Ala Phe Glu Glu Met Thr Arg Asn Leu Phe Ser Leu  
210 215 220

Pro Ile Asp Val Pro Phe Ser Gly Leu Tyr Arg Gly Met Lys Ala Arg  
225 230 235 240

Asn Leu Ile His Ala Arg Ile Glu Gln Asn Ile Arg Ala Lys Ile Cys  
245 250 255

Gly Leu Arg Ala Ser Glu Ala Gly Gln Gly Cys Lys Asp Ala Leu Gln  
260 265 270

Leu Leu Ile Glu His Ser Trp Glu Arg Gly Glu Arg Leu Asp Met Gln  
275 280 285

Ala Leu Lys Gln Ser Ser Thr Glu Leu Leu Phe Gly Gly His Glu Thr  
290 295 300

Thr Ala Ser Ala Ala Thr Ser Leu Ile Thr Tyr Leu Gly Leu Tyr Pro  
305 310 315 320

His Val Leu Gln Lys Val Arg Glu Glu Leu Lys Ser Lys Gly Leu Leu  
325 330 335

Cys Lys Ser Asn Gln Asp Asn Lys Leu Asp Met Glu Ile Leu Glu Gln  
340 345 350

Leu Lys Tyr Ile Gly Cys Val Ile Lys Glu Thr Leu Arg Leu Asn Pro  
355 360 365

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Pro Val Pro Gly Gly Phe Arg Val Ala Leu Lys Thr Phe Glu Leu Asn  
370 375 380

Gly Tyr Gln Ile Pro Lys Gly Trp Asn Val Ile Tyr Ser Ile Cys Asp  
385 390 395 400

Thr His Asp Val Ala Glu Ile Phe Thr Asn Lys Glu Glu Phe Asn Pro  
405 410 415

Asp Arg Phe Ser Ala Pro His Pro Glu Asp Ala Ser Arg Phe Ser Phe  
420 425 430

Ile Pro Phe Gly Gly Gly Leu Arg Ser Cys Val Gly Lys Glu Phe Ala  
435 440 445

Lys Ile Leu Leu Lys Ile Phe Thr Val Glu Leu Ala Arg His Cys Asp  
450 455 460

Trp Gln Leu Leu Asn Gly Pro Pro Thr Met Lys Thr Ser Pro Thr Val  
465 470 475 480

Tyr Pro Val Asp Asn Leu Pro Ala Arg Phe Thr His Phe His Gly Glu  
485 490 495

Ile

(2) INFORMATION FOR SEQ ID NO:5

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1494 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5

ATG GGG CTC CCG GCG CTG CTG GCC AGT GCG CTC TGC ACC TTC GTG CTG	48
Met Gly Leu Pro Ala Leu Leu Ala Ser Ala Leu Cys Thr Phe Val Leu	
1 5 10 15	
CCG CTG CTG CTC TTC CTG GCT GCG ATC AAG CTC TGG GAC CTG TAC TGC	96
Pro Leu Leu Leu Phe Leu Ala Ala Ile Lys Leu Trp Asp Leu Tyr Cys	
20 25 30	
GTG AGC GGC CGC GAC CGC AGT TGT GCC CTC CCA TTG CCC CCC GGG ACT	144
Val Ser Gly Arg Asp Arg Ser Cys Ala Leu Pro Leu Pro Pro Gly Thr	
35 40 45	
ATG GGC TTC CCC TTC TTT GGG GAA ACC TTG CAG ATG GTA CTG CAG CGG	192
Met Gly Phe Pro Phe Phe Gly Glu Thr Leu Gln Met Val Leu Gln Arg	
50 55 60	
AGG AAG TTC CTG CAG ATG AAG CGC AGG AAA TAC GGC TTC ATC TAC AAG	240
Arg Lys Phe Leu Gln Met Lys Arg Arg Lys Tyr Gly Phe Ile Tyr Lys	
65 70 75 80	
ACG CAT CTG TTC GGG CGG CCC ACC GTA CGG GTG ATG GGC GCG GAC AAT	288
Thr His Leu Phe Gly Arg Pro Thr Val Arg Val Met Gly Ala Asp Asn	
85 90 95	
GTG CGG CGC ATC TTG CTC GGA GAC GAC CGG CTG GTG TCG GTC CAC TGG	336
Val Arg Arg Ile Leu Leu Gly Asp Asp Arg Leu Val Ser Val His Trp	
100 105 110	

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CCA	GCG	TCG	GTG	CGC	ACC	ATT	CTG	GGA	TCT	GGC	TGC	CTC	TCT	AAC	CTG	384
Pro	Ala	Ser	Val	Arg	Thr	Ile	Leu	Gly	Ser	Gly	Cys	Leu	Ser	Asn	Leu	
		115					120					125				
CAC	GAC	TCC	TCG	CAC	AAG	CAG	CGC	AAG	AAG	GTG	ATT	ATG	CGG	GCC	TTC	432
His	Asp	Ser	Ser	His	Lys	Gln	Arg	Lys	Lys	Val	Ile	Met	Arg	Ala	Phe	
		130				135					140					
AGC	CGC	GAG	GCA	CTC	GAA	TGC	TAC	GTG	CCG	GTG	ATC	ACC	GAG	GAA	GTG	480
Ser	Arg	Glu	Ala	Leu	Glu	Cys	Tyr	Val	Pro	Val	Ile	Thr	Glu	Glu	Val	
		145			150					155					160	
GGC	AGC	AGC	CTG	GAG	CAG	TGG	CTG	AGC	TGC	GGC	GAG	CGC	GGC	CTC	CTG	528
Gly	Ser	Ser	Leu	Glu	Gln	Trp	Leu	Ser	Cys	Gly	Glu	Arg	Gly	Leu	Leu	
				165					170					175		
GTC	TAC	CCC	GAG	GTG	AAG	CGC	CTC	ATG	TTC	CGA	ATC	GCC	ATG	CGC	ATC	576
Val	Tyr	Pro	Glu	Val	Lys	Arg	Leu	Met	Phe	Arg	Ile	Ala	Met	Arg	Ile	
			180					185					190			
CTA	CTG	GGC	TGC	GAA	CCC	CAA	CTG	GCG	GGC	GAC	GGG	GAC	TCC	GAG	CAG	624
Leu	Leu	Gly	Cys	Glu	Pro	Gln	Leu	Ala	Gly	Asp	Gly	Asp	Ser	Glu	Gln	
		195					200					205				
CAG	CTT	GTG	GAG	GCC	TTC	GAG	GAA	ATG	ACC	CGC	AAT	CTC	TTC	TCG	CTG	672
Gln	Leu	Val	Glu	Ala	Phe	Glu	Glu	Met	Thr	Arg	Asn	Leu	Phe	Ser	Leu	
		210				215					220					
CCC	ATC	GAC	GTG	CCC	TTC	AGC	GGG	CTG	TAC	CGG	GGC	ATG	AAG	GCG	CGG	720
Pro	Ile	Asp	Val	Pro	Phe	Ser	Gly	Leu	Tyr	Arg	Gly	Met	Lys	Ala	Arg	
		225			230					235					240	
AAC	CTC	ATT	CAC	GCG	CGC	ATC	GAG	CAG	AAC	ATT	CGC	GCC	AAG	ATC	TGC	768
Asn	Leu	Ile	His	Ala	Arg	Ile	Glu	Gln	Asn	Ile	Arg	Ala	Lys	Ile	Cys	
				245					250					255		
GGG	CTG	CGG	GCA	TCC	GAG	GCG	GGC	CAG	GGC	TGC	AAA	GAC	GCG	CTG	CAG	816
Gly	Leu	Arg	Ala	Ser	Glu	Ala	Gly	Gln	Gly	Cys	Lys	Asp	Ala	Leu	Gln	
			260				265						270			
CTG	TTG	ATC	GAG	CAC	TCG	TGG	GAG	AGG	GGA	GAG	CGG	CTG	GAC	ATG	CAG	864
Leu	Leu	Ile	Glu	His	Ser	Trp	Glu	Arg	Gly	Glu	Arg	Leu	Asp	Met	Gln	
		275					280					285				
GCA	CTA	AAG	CAA	TCT	TCA	ACC	GAA	CTC	CTC	TTT	GGA	GGA	CAC	GAA	ACC	912
Ala	Leu	Lys	Gln	Ser	Ser	Thr	Glu	Leu	Leu	Phe	Gly	Gly	His	Glu	Thr	
		290				295					300					
ACG	GCC	AGT	GCA	GCC	ACA	TCT	CTG	ATC	ACT	TAC	CTG	GGG	CTC	TAC	CCA	960
Thr	Ala	Ser	Ala	Ala	Thr	Ser	Leu	Ile	Thr	Tyr	Leu	Gly	Leu	Tyr	Pro	
		305			310					315					320	
CAT	GTT	CTC	CAG	AAA	GTG	CGA	GAA	GAG	CTG	AAG	AGT	AAG	GGT	TTA	CTT	1008
His	Val	Leu	Gln	Lys	Val	Arg	Glu	Glu	Leu	Lys	Ser	Lys	Gly	Leu	Leu	
				325					330					335		
TGC	AAG	AGC	AAT	CAA	GAC	AAC	AAG	TTG	GAC	ATG	GAA	ATT	TTG	GAA	CAA	1056
Cys	Lys	Ser	Asn	Gln	Asp	Asn	Lys	Leu	Asp	Met	Glu	Ile	Leu	Glu	Gln	
			340					345					350			
CTT	AAA	TAC	ATC	GGG	TGT	GTT	ATT	AAG	GAG	ACC	CTT	CGA	CTG	AAT	CCC	1104
Leu	Lys	Tyr	Ile	Gly	Cys	Val	Ile	Lys	Glu	Thr	Leu	Arg	Leu	Asn	Pro	
		355					360					365				

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CCA GTT CCA GGA GGG TTT CGG GTT GCT CTG AAG ACT TTT GAA TTA AAT	1152
Pro Val Pro Gly Gly Phe Arg Val Ala Leu Lys Thr Phe Glu Leu Asn	
370 375 380	
GGA TAC CAG ATT CCC AAG GGC TGG AAT GTT ATC TAC AGT ATC TGT GAT	1200
Gly Tyr Gln Ile Pro Lys Gly Trp Asn Val Ile Tyr Ser Ile Cys Asp	
385 390 395 400	
ACT CAT GAT GTG GCA GAG ATC TTC ACC AAC AAG GAA GAA TTT AAT CCT	1248
Thr His Asp Val Ala Glu Ile Phe Thr Asn Lys Glu Glu Phe Asn Pro	
405 410 415	
GAC CGA TTC AGT GCT CCT CAC CCA GAG GAT GCA TCC AGG TTC AGC TTC	1296
Asp Arg Phe Ser Ala Pro His Pro Glu Asp Ala Ser Arg Phe Ser Phe	
420 425 430	
ATT CCA TTT GGA GGA GGC CTT AGG AGC TGT GTA GGC AAA GAA TTT GCA	1344
Ile Pro Phe Gly Gly Gly Leu Arg Ser Cys Val Gly Lys Glu Phe Ala	
435 440 445	
AAA ATT CTT CTC AAA ATA TTT ACA GTG GAG CTG GCC AGG CAT TGT GAC	1392
Lys Ile Leu Leu Lys Ile Phe Thr Val Glu Leu Ala Arg His Cys Asp	
450 455 460	
TGG CAG CTT CTA AAT GGA CCT CCT ACA ATG AAA ACC AGT CCC ACC GTG	1440
Trp Gln Leu Leu Asn Gly Pro Pro Thr Met Lys Thr Ser Pro Thr Val	
465 470 475 480	
TAT CCT GTG GAC AAT CTC CCT GCA AGA TTC ACC CAT TTC CAT GGG GAA	1488
Tyr Pro Val Asp Asn Leu Pro Ala Arg Phe Thr His Phe His Gly Glu	
485 490 495	
ATC TGA	1494
Ile	

- (2) INFORMATION FOR SEQ ID NO:6  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 20 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6

Pro Phe Gly Gly Gly Pro Arg Leu Cys Pro Gly Tyr Glu Leu Ala Arg
1 5 10 15
Val Ala Leu Ser
20

- (2) INFORMATION FOR SEQ ID NO:7  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 20 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7

Pro Phe Ser Gly Gly Ala Arg Asn Cys Ile Gly Lys Gln Phe Ala Met
1 5 10 15
Ser Glu Met Lys
20

(2) INFORMATION FOR SEQ ID NO:8

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 20 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8

Pro Phe Ser Gly Gly Ala Arg Asn Cys Ile Gly Lys Gln Phe Ala Met  
 1 5 10 15  
 Asn Glu Leu Lys  
 20

(2) INFORMATION FOR SEQ ID NO:9

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 20 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9

Pro Phe Gly Thr Gly Pro Arg Asn Cys Ile Gly Met Arg Phe Ala Ile  
 1 5 10 15  
 Met Asn Met Lys  
 20

(2) INFORMATION FOR SEQ ID NO:10

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 20 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10

Pro Phe Ser Gly Gly Ser Arg Asn Cys Ile Gly Lys Gln Phe Ala Met  
 1 5 10 15  
 Asn Glu Leu Lys  
 20

(2) INFORMATION FOR SEQ ID NO:11

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 351 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11

GAACCTCTCT TTGGAGGACA CGAAACCACG GCCAGTGCAG CCACATCTCT GATCACTTAC	60
CTGGGGCTCT ACCCACATGT TCTCCAGAAA GTGCGAGAAG AGCTGAAGAG TAAGGGTTTA	120
CTTTGCAAGA GCAATCAAGA CAACAAGTTG GACATGGAAA TTTTGGAACA ACTTAAATAC	180
ATCGGGTGTG TTATTAAGGA GACCCTTCGA CTGAATCCCC CAGTTCCAGG AGGGTTTCGG	240
GTTGCTCTGA AGACTTTTGA ATTAAATGGA TACCAGATTC CCAAGGGCTG GAATGTTATC	300
TACAGTATCT GTGATACTCA TGATGTGGCA GAGATCTTCA CCAACAAGGA A	351

- (2) INFORMATION FOR SEQ ID NO:12  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12

TTTTTTTTTT TTGG

14

- (2) INFORMATION FOR SEQ ID NO:13  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13

TTTTTTTTTT TTGA

14

- (2) INFORMATION FOR SEQ ID NO:14  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14

TTTTTTTTTT TTGT

14

- (2) INFORMATION FOR SEQ ID NO:15  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15

TTTTTTTTTT TTGC

14

- (2) INFORMATION FOR SEQ ID NO:16  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16

TTTTTTTTTT TTAG

14

- (2) INFORMATION FOR SEQ ID NO:17  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 14 base pairs  
(B) TYPE: nucleic acid  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17

TTTTTTTTTT TTAA

14

- (2) INFORMATION FOR SEQ ID NO:18  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18

TTTTTTTTTTT TTAT 14

- (2) INFORMATION FOR SEQ ID NO:19  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19

TTTTTTTTTTT TTAC 14

- (2) INFORMATION FOR SEQ ID NO:20  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20

TTTTTTTTTTT TTCG 14

- (2) INFORMATION FOR SEQ ID NO:21  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21

TTTTTTTTTTT TTCA 14

- (2) INFORMATION FOR SEQ ID NO:22  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22

TTTTTTTTTTT TTCT 14

- (2) INFORMATION FOR SEQ ID NO:23  
  (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 14 base pairs  
    (B) TYPE: nucleic acid  
    (D) TOPOLOGY: linear  
  
  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23

TTTTTTTTTTT TTCC 14

- (2) INFORMATION FOR SEQ ID NO:24  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 10 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24

AAGCGACCGA

10

- (2) INFORMATION FOR SEQ ID NO:25  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 10 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25

TGTTCGCCAG

10

- (2) INFORMATION FOR SEQ ID NO:26  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 10 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26

TGCCAGTGGA

10

- (2) INFORMATION FOR SEQ ID NO:27  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 10 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27

GGCTGCAAAC

10

- (2) INFORMATION FOR SEQ ID NO:28  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 10 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28

CCTAGCGTTG

10

- (2) INFORMATION FOR SEQ ID NO:29  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 21 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear  
  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29

GTCGCGCCG CTGCCAGTGG A

21



- (2) INFORMATION FOR SEQ ID NO:30  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 12 base pairs  
 (B) TYPE: nucleic acid  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30

GTAGCGGCCG CT

12

- (2) INFORMATION FOR SEQ ID NO:31  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 1725 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31 .

GCACGAGGGA GGCTGAAGCG TGCC ATG GGG CTC CCG GCG CTG CTG GCC AGT	51
Met Gly Leu Pro Ala Leu Leu Ala Ser	
1 5	
GCG CTC TGC ACC TTC GTG CTG CCG CTG CTG CTC TTC CTG GCG GCG CTC	99
Ala Leu Cys Thr Phe Val Leu Pro Leu Leu Leu Phe Leu Ala Ala Leu	
10 15 20 25	
AAG CTC TGG GAC CTG TAC TGT GTG AGC AGC CGC GAT CGC AGC TGC GCC	147
Lys Leu Trp Asp Leu Tyr Cys Val Ser Ser Arg Asp Arg Ser Cys Ala	
30 35 40	
CTC CCC TTG CCC CCC GGT ACC ATG GGC TTC CCA TTC TTT GGG GAA ACA	195
Leu Pro Leu Pro Pro Gly Thr Met Gly Phe Pro Phe Phe Gly Glu Thr	
45 50 55	
TTG CAG ATG GTG CTT CAG CGG AGG AAG TTT CTG CAG ATG AAG CGC AGG	243
Leu Gln Met Val Leu Gln Arg Arg Lys Phe Leu Gln Met Lys Arg Arg	
60 65 70	
AAA TAC GGC TTC ATC TAC AAG ACG CAT CTG TTT GGG CGG CCC ACG GTG	291
Lys Tyr Gly Phe Ile Tyr Lys Thr His Leu Phe Gly Arg Pro Thr Val	
75 80 85	
CGG GTG ATG GGC GCG GAT AAT GTG CGG CGC ATC TTG CTG GGA GAG CAC	339
Arg Val Met Gly Ala Asp Asn Val Arg Arg Ile Leu Leu Gly Glu His	
90 95 100 105	
CGG TTG GTG TCG GTG CAC TGG CCC GCG TCG GTG CGC ACC ATC CTG GGC	387
Arg Leu Val Ser Val His Trp Pro Ala Ser Val Arg Thr Ile Leu Gly	
110 115 120	
GCT GGC TGC CTC TCC AAC CTG CAC GAT TCC TCG CAC AAG CAG CGA AAG	435
Ala Gly Cys Leu Ser Asn Leu His Asp Ser Ser His Lys Gln Arg Lys	
125 130 135	
AAG GTG ATT ATG CAG GCC TTC AGC CGC GAG GCA CTC CAG TGC TAC GTG	483
Lys Val Ile Met Gln Ala Phe Ser Arg Glu Ala Leu Gln Cys Tyr Val	
140 145 150	
CTC GTG ATC GCT GAG GAA GTC AGC AGT TGT CTG GAG CAG TGG CTA AGC	531
Leu Val Ile Ala Glu Glu Val Ser Ser Cys Leu Glu Gln Trp Leu Ser	
155 160 165	

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TGC	GGC	GAG	CGC	GGC	CTC	CTG	GTC	TAC	CCC	GAG	GTG	AAG	CGC	CTC	ATG	579
Cys	Gly	Glu	Arg	Gly	Leu	Leu	Val	Tyr	Pro	Glu	Val	Lys	Arg	Leu	Met	
170					175					180					185	
TTC	CGC	ATC	GCC	ATG	CGC	ATC	CTG	CTG	GGC	TGC	GAG	CCG	GGT	CCA	GCG	627
Phe	Arg	Ile	Ala	Met	Arg	Ile	Leu	Leu	Gly	Cys	Glu	Pro	Gly	Pro	Ala	
				190					195					200		
GGC	GGC	GGG	GAG	GAC	GAG	CAA	CAG	CTC	GTG	GAG	GCT	TTC	GAG	GAG	ATG	675
Gly	Gly	Gly	Glu	Asp	Glu	Gln	Gln	Leu	Val	Glu	Ala	Phe	Glu	Glu	Met	
			205					210					215			
ACC	CGC	AAT	CTC	TTC	TCT	CTT	CCC	ATT	GAC	GTG	CCC	TTT	AGC	GGC	CTG	723
Thr	Arg	Asn	Leu	Phe	Ser	Leu	Pro	Ile	Asp	Val	Pro	Phe	Ser	Gly	Leu	
		220					225					230				
TAC	CGG	GGC	GTG	AAG	GCG	CGG	AAC	CTT	ATA	CAC	GCG	CGC	ATC	GAG	GAG	771
Tyr	Arg	Gly	Val	Lys	Ala	Arg	Asn	Leu	Ile	His	Ala	Arg	Ile	Glu	Glu	
	235				240						245					
AAC	ATT	CGC	GCC	AAG	ATC	CGC	CGG	CTT	CAG	GCT	ACA	GAG	CCG	GAT	GGG	819
Asn	Ile	Arg	Ala	Lys	Ile	Arg	Arg	Leu	Gln	Ala	Thr	Glu	Pro	Asp	Gly	
250					255				260						265	
GGT	TGC	AAG	GAC	GCG	CTG	CAG	CTC	CTG	ATT	GAG	CAC	TCG	TGG	GAG	AGG	867
Gly	Cys	Lys	Asp	Ala	Leu	Gln	Leu	Leu	Ile	Glu	His	Ser	Trp	Glu	Arg	
			270					275						280		
GGA	GAG	AGG	CTG	GAT	ATG	CAG	GCA	CTA	AAA	CAA	TCG	TCA	ACA	GAG	CTC	915
Gly	Glu	Arg	Leu	Asp	Met	Gln	Ala	Leu	Lys	Gln	Ser	Ser	Thr	Glu	Leu	
			285					290					295			
CTC	TTT	GGT	GGT	CAT	GAA	ACT	ACA	GCC	AGT	GCT	GCG	ACA	TCA	CTG	ATC	963
Leu	Phe	Gly	Gly	His	Glu	Thr	Thr	Ala	Ser	Ala	Ala	Thr	Ser	Leu	Ile	
		300					305					310				
ACT	TAC	CTA	GGA	CTC	TAC	CCA	CAT	GTC	CTC	CAG	AAA	GTT	CGA	GAA	GAG	1011
Thr	Tyr	Leu	Gly	Leu	Tyr	Pro	His	Val	Leu	Gln	Lys	Val	Arg	Glu	Glu	
	315					320					325					
ATA	AAG	AGC	AAG	GGC	TTA	CTT	TGC	AAG	AGC	AAT	CAA	GAC	AAC	AAG	TTA	1059
Ile	Lys	Ser	Lys	Gly	Leu	Leu	Cys	Lys	Ser	Asn	Gln	Asp	Asn	Lys	Leu	
330					335					340					345	
GAC	ATG	GAA	ACT	TTG	GAA	CAG	CTT	AAA	TAC	ATT	GGG	TGT	GTC	ATT	AAG	1107
Asp	Met	Glu	Thr	Leu	Glu	Gln	Leu	Lys	Tyr	Ile	Gly	Cys	Val	Ile	Lys	
				350				355						360		
GAG	ACC	CTG	CGA	TTG	AAT	CCT	CCG	GTT	CCA	GGA	GGG	TTT	CGG	GTT	GCT	1155
Glu	Thr	Leu	Arg	Leu	Asn	Pro	Pro	Val	Pro	Gly	Gly	Phe	Arg	Val	Ala	
			365					370					375			
CTG	AAG	ACT	TTT	GAG	CTG	AAT	GGA	TAC	CAG	ATC	CCC	AAG	GGC	TGG	AAT	1203
Leu	Lys	Thr	Phe	Glu	Leu	Asn	Gly	Tyr	Gln	Ile	Pro	Lys	Gly	Trp	Asn	
		380					385					390				
GTT	ATT	TAC	AGT	ATC	TGT	GAC	ACC	CAC	GAT	GTG	GCA	GAT	ATC	TTC	ACT	1251
Val	Ile	Tyr	Ser	Ile	Cys	Asp	Thr	His	Asp	Val	Ala	Asp	Ile	Phe	Thr	
	395					400					405					
AAC	AAG	GAG	GAA	TTT	AAT	CCC	GAC	CGC	TTT	ATA	GTG	CCT	CAT	CCA	GAG	1299
Asn	Lys	Glu	Glu	Phe	Asn	Pro	Asp	Arg	Phe	Ile	Val	Pro	His	Pro	Glu	
410					415					420					425	

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GAT GCT TCC CGG TTC AGC TTC ATT CCA TTT GGA GGA GGC CTT CGG AGC	1347
Asp Ala Ser Arg Phe Ser Phe Ile Pro Phe Gly Gly Gly Leu Arg Ser	
430 435 440	
TGT GTA GGC AAA GAG TTT GCA AAA ATT CTT CTT AAG ATA TTT ACA GTG	1395
Cys Val Gly Lys Glu Phe Ala Lys Ile Leu Leu Lys Ile Phe Thr Val	
445 450 455	
GAG CTG GCT AGG CAC TGT GAT TGG CAG CTT CTA AAT GGA CCT CCT ACA	1443
Glu Leu Ala Arg His Cys Asp Trp Gln Leu Leu Asn Gly Pro Pro Thr	
460 465 470	
ATG AAG ACA AGC CCC ACT GTG TAC CCT GTG GAC AAT CTC CCT GCA AGA	1491
Met Lys Thr Ser Pro Thr Val Tyr Pro Val Asp Asn Leu Pro Ala Arg	
475 480 485	
TTC ACC TAC TTC CAG GGA GAT ATC TGATAGCTAT TTCAATTCTT	1535
Phe Thr Tyr Phe Gln Gly Asp Ile	
490 495	
GGACTTATTT GAAGTGATA TTGGTTTTTTT TAAAAAATAG TGTCATGTTG ACTTTATTTA	1595
ATTTCTAAAT GTATAGTATG ATATTTATGT GTCTCTACTA CAGTCCCGTG GTCTTTAAAT	1655
ATTAAAAATAA TGAATTTGTA TGATTTCCCA ATAAAGTAAA ATTAAAAAGT GAAAAAAAAA	1715
AAAAAAAAAA	1725

(2) INFORMATION FOR SEQ ID NO:32

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 497 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32

Met Gly Leu Pro Ala Leu Leu Ala Ser Ala Leu Cys Thr Phe Val Leu	
1 5 10 15	
Pro Leu Leu Leu Phe Leu Ala Ala Leu Lys Leu Trp Asp Leu Tyr Cys	
20 25 30	
Val Ser Ser Arg Asp Arg Ser Cys Ala Leu Pro Leu Pro Pro Gly Thr	
35 40 45	
Met Gly Phe Pro Phe Phe Gly Glu Thr Leu Gln Met Val Leu Gln Arg	
50 55 60	
Arg Lys Phe Leu Gln Met Lys Arg Arg Lys Tyr Gly Phe Ile Tyr Lys	
65 70 75 80	
Thr His Leu Phe Gly Arg Pro Thr Val Arg Val Met Gly Ala Asp Asn	
85 90 95	
Val Arg Arg Ile Leu Leu Gly Glu His Arg Leu Val Ser Val His Trp	
100 105 110	
Pro Ala Ser Val Arg Thr Ile Leu Gly Ala Gly Cys Leu Ser Asn Leu	
115 120 125	
His Asp Ser Ser His Lys Gln Arg Lys Lys Val Ile Met Gln Ala Phe	
130 135 140	

Ser	Arg	Glu	Ala	Leu	Gln	Cys	Tyr	Val	Leu	Val	Ile	Ala	Glu	Glu	Val
145					150					155					160
Ser	Ser	Cys	Leu	Glu	Gln	Trp	Leu	Ser	Cys	Gly	Glu	Arg	Gly	Leu	Leu
			165						170					175	
Val	Tyr	Pro	Glu	Val	Lys	Arg	Leu	Met	Phe	Arg	Ile	Ala	Met	Arg	Ile
			180					185					190		
Leu	Leu	Gly	Cys	Glu	Pro	Gly	Pro	Ala	Gly	Gly	Gly	Glu	Asp	Glu	Gln
		195					200					205			
Gln	Leu	Val	Glu	Ala	Phe	Glu	Met	Thr	Arg	Asn	Leu	Phe	Ser	Leu	
	210					215				220					
Pro	Ile	Asp	Val	Pro	Phe	Ser	Gly	Leu	Tyr	Arg	Gly	Val	Lys	Ala	Arg
225					230					235					240
Asn	Leu	Ile	His	Ala	Arg	Ile	Glu	Glu	Asn	Ile	Arg	Ala	Lys	Ile	Arg
				245					250					255	
Arg	Leu	Gln	Ala	Thr	Glu	Pro	Asp	Gly	Gly	Cys	Lys	Asp	Ala	Leu	Gln
			260					265					270		
Leu	Leu	Ile	Glu	His	Ser	Trp	Glu	Arg	Gly	Glu	Arg	Leu	Asp	Met	Gln
		275					280					285			
Ala	Leu	Lys	Gln	Ser	Ser	Thr	Glu	Leu	Leu	Phe	Gly	Gly	His	Glu	Thr
	290					295					300				
Thr	Ala	Ser	Ala	Ala	Thr	Ser	Leu	Ile	Thr	Tyr	Leu	Gly	Leu	Tyr	Pro
305					310					315					320
His	Val	Leu	Gln	Lys	Val	Arg	Glu	Glu	Ile	Lys	Ser	Lys	Gly	Leu	Leu
				325					330					335	
Cys	Lys	Ser	Asn	Gln	Asp	Asn	Lys	Leu	Asp	Met	Glu	Thr	Leu	Glu	Gln
			340					345					350		
Leu	Lys	Tyr	Ile	Gly	Cys	Val	Ile	Lys	Glu	Thr	Leu	Arg	Leu	Asn	Pro
		355					360					365			
Pro	Val	Pro	Gly	Gly	Phe	Arg	Val	Ala	Leu	Lys	Thr	Phe	Glu	Leu	Asn
	370					375					380				
Gly	Tyr	Gln	Ile	Pro	Lys	Gly	Trp	Asn	Val	Ile	Tyr	Ser	Ile	Cys	Asp
385					390					395					400
Thr	His	Asp	Val	Ala	Asp	Ile	Phe	Thr	Asn	Lys	Glu	Glu	Phe	Asn	Pro
				405					410					415	
Asp	Arg	Phe	Ile	Val	Pro	His	Pro	Glu	Asp	Ala	Ser	Arg	Phe	Ser	Phe
			420					425					430		
Ile	Pro	Phe	Gly	Gly	Gly	Leu	Arg	Ser	Cys	Val	Gly	Lys	Glu	Phe	Ala
		435					440					445			
Lys	Ile	Leu	Leu	Lys	Ile	Phe	Thr	Val	Glu	Leu	Ala	Arg	His	Cys	Asp
	450					455					460				
Trp	Gln	Leu	Leu	Asn	Gly	Pro	Pro	Thr	Met	Lys	Thr	Ser	Pro	Thr	Val
465					470					475					480

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Tyr Pro Val Asp Asn Leu Pro Ala Arg Phe Thr Tyr Phe Gln Gly Asp  
485 490 495

Ile

(2) INFORMATION FOR SEQ ID NO:33

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33

CGCACCCCAG GAGGCGCGCT CGGAGGGAAG CCGCCACCGC CGCCGCCTCT GCCTCGGCGC 60  
GGAACAAACG GTTAAAGATT TTGGGCCASC GCCTCCGCGG GGGGAGGAGC CAGGGGCCCC 120  
AATCCCGCAA TTAAAGATGA ACTTTGGGTG AACTAATTGT CTGACCAAGG TAACGTGGGC 180  
AGCAACCTGG GCCGCCTATA AAGCGGCAGC GCCGTGGGGT TTGAAGCGCT GCGGCGGCGC 240  
GCAGGTGGCG CGGGAGGTCG CGGCGCGCCA TGG 273

(2) INFORMATION FOR SEQ ID NO:34

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34

CGCACCCCCA GGAGGCGCGC TCAGAGGGAA GCCGCCAGTG CGCCGCCTCT GCCTCGGCGC 60  
GGAACAAACG GTTAAAGATT TTTTGGGCA GCGCCTCGAG GGGGAGGAG CCAGGGGCCC 120  
GATCCGCAAT TAAAGATGAA CTTTGGGTGA ACTAATTTGT CTGACCAAGG TAACGTGGGC 180  
AGTAACCTGG GCGGCCTTAT AAAGAGGGCG CGCGGCGGGG TTCGAGCTA GGGAGGCGGC 240  
GGCAGGTGGC GCGGGAGGCT GAAGCGTGCC ATGG 274

(2) INFORMATION FOR SEQ ID NO:35

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 319 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35

TCGGGGGAAT TAACACCTTT TCAAAGTGAA ATCTCAGGAT TGTCTGCCTT CTACAGGAGG 60  
TGGTATTAAA ATGCGCCTAT AACAAATGGT TGAGAGTTTG GAGCCGCTTC TGCCCTGTGG 120  
GCGGGGCGAG ATGACACCAC AATTAAAGAT GAACTTTGGG TGAATAATT TATCTGAGGA 180  
AGTTAACAGG AGGAGACCTG CGCGCAATGG ATATATAAGG GCGCGCAGGC GAGGACGCCC 240  
TCAGTTTGTG CGTAAAGACG CGTCTCCTCT CCAGAAGCTT GTTTTTCGTT TTGGCGATCA 300  
GTTGCGCGCT TCAACATGG 319

(2) INFORMATION FOR SEQ ID NO:36

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36

GATCCCAGATCTGCCTATTGCGCCCGATGCCCCGAGGCTCTCTCTTGGAC  
TCTGGCCCTGAGTTCTTCTGCGCGATCCTTCGGAGACGTCTGGAGGCCTG  
CTTTATGCATCTCTCTTGGACCTCAGTTTCCCCACACGTGGGAGGAGGCA  
GCTGGACGATTCTGAAAGGACTTTCCTTGCTTCCTCATCACGTGGAAG  
AGAGCCCAACCGGCACCTGGAATGGAAAGCCAGTGAAGGCTGCTTGGG  
CCGGGGCAKCGGTGGGACCGGGCGGGAGGGATTCCAAGAGACCGCCGG  
GAAGGCTAGAGCTTGAATTCGGCTCCTCGGAGTCTGGCCCTCCCCCA  
CCGCCGCTCGGAGCTCAGCACACCTTGGATGGGGGAGGCGGGCAGCTCC  
TAGCCCCGACCCAGGAGGCGCTCGGAGGGAAGCCGCCACCGCCGCC  
GCCTCTGCCTCGGCGCGGAACAAACGGTTAAAGATTTTGGGCCASCCT  
CCGCGGGGGAGGAGCCAGGGGCCCAATCCCGCAATTAAAGATGAACTT  
TGGGTGAATAATTGTCTGACCAAGGTAACGTGGGACGCAACCTGGGCG  
CCTATAAAGCGGCAGCGCCCTGGGGTTTGAAGCGCTGGCGCGGCGCAG  
GTGGCGCGGGAGGTGCGGCGCGCCATGGGGCTCCGCGCGCTGCTGGCCA  
GTGCGCTCTGCACCTTCGTGCTGCCGCTGCTGCTCTTCTGGCTGCGATC  
AAGCTCTGGGACCTGTACTGCGTGAGCGGCCGCGACCGCAGTTGTGCCCT  
CCCATTGCCCCCGGACTATSGGSTTCCCCCTTCTTGGGGAACCTTGC  
AGATNTACTNCAGGTAAGGGAGGGTGGGGCGGACAGGCTGCTTCCCCG  
GAGCCCGCGCGGCTCTGGGCTTCTGCTGAAGTCGGGGTAGGCGCCCCG  
GGAGGCATGCTATTGCGGCTAGGAGCAGGGCTGGCGGGAGCGCGGCGCTC  
CCCGGMKYMCSCTCAWGCSCRCWWKTMWCCTCCGCCTYMCTCCAMAGCG  
GARSARWKYKGMRGATGAAGCGCAGGAAATACGGCTTCATCTACAAGA  
CGCATCTGTTGGGGCGGCCACCGTACGGGTGATGGGCGCGGACAATGTG  
CGCGCATCTTGCTCGGAGAGCACCGGCTGGTGTGCGTCCACTGGCCAGC  
GTGCGTGCGACCATTTCTGGGATCTGGCTGCCTCTCTAACCTGCACGACT  
CCTCGCACAGCAGCGCAAGAGGTGGGGGAGGAGGCGACGGCTGGACA  
GGGAGGGGGACCCATTTATGAGCGGAATTCGGCTGATGGATGCTAGGC  
GCGGGCTAGCAGCTTGGGTGGGCTAGGACCTCTGCCAGCTCCAGGTTA  
GCTTTCCAGCTCGGAGAGTGCCATGTGTCTGGCAGGACTGGGGGTGTCT  
GGAAGGGGACGGCGGTAGACGAGAGGGGCGGATGGAGGCTTTTAACGCTG  
TCCCCCTCCTCGGACTCAGGTGATTATGCGGGCCTTCAGCCGCGAGGCAC  
TCGAATGCTACGTGCCGGTGATCACCAGGAAGTGGGCAGCAGCCTGGAG  
CAGTGGCTGAGCTGCGGCGAGCGCGGCTCCTGGTCTACCCCGAGGTGAA  
GCGCCTCATGTTCCGAATCGCCATGCGCATCCTACTGGGCTGCGAACCC  
AAGTGGCGGGCGACGGGACTCCGAGCAGCAGCTTGTGGAGGCCTTCGAG  
GAAATGACCCGCAATCTCTCTCGCTGCCCATCGACGTGCCCTTCAGCGG  
GCTGTACCGGTAAGGGCGGCAACGGGCTGCGGACTAGGGGCGCGGGAC  
CTGGGCGTCTGCTACCGCCGCGCGCTCTCTGCGCTCAGGGCATGAAGGC  
GCGGAACCTCATTCACGCGCGCATCGAGCAGAACATTGCGGCCAAGATCT  
GCGGGCTGCGGGCATCCGAGCGGGCCAGGGCTGCAAGACGCGCTGCAG  
CTGTTGATCGAGCACTCGTGGGAGAGGGGAGAGCGGCTGGACATGCAGGT  
GAGTAGCAGCTTCAGACCAGGCACTGCGGAGTTTGGTCCCCCTGGCTTTCC  
AAGGCGCTGTTCTGGGGCCCCAAAGCGCGCGCTGGGGCCAGCTTTC  
TGGAGTGGGCGGCCGGCTCAGACTACAGCTATGGAATCCCGAAGGAAGGC  
TGAGACACCCGTCAGGAGAGCTGCGGAAGGGGCTGCGMGGAACCTGGG  
AGACTCCCTAGCCTTTAMCAGGTTTCAAAGGGAAAGTTGGAATTTGCAA  
AAATGTTAATAAAGAACCTTGCATTTTAATAAACTAAGACTTTAACTC  
AGGAGTTTCCGGTAGRCGGGGTCTGACTCGCCTTACTGCTCCAGCTGAA  
CTAAAGGGACGTTGCATTTTGTAAAGATATTGCTTTCCTTGACTTTCT  
GTCAGCAAAACATTTAGCCCTTCTAGTCTTCCCTCCAGAACTCTCAGTTC  
GATTCTGAGTAATCCTTCTGTCAAACCGCAGGCAGACTTGTGAGAATGTG  
GGTCTCACTTATTTAGGCACTAAAGCAATCTTCAACCGAACTCCTCT  
TTGGAGGACACGAACACGCGCAGTGCAGCCACATCTCTGATCACTTAC  
CTGGGGCTCTACCCACATGTTCTCCAG

(2) INFORMATION FOR SEQ ID NO:37

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37

GATCCAGGTTGCTGAAACATATCTCCATATAGGGCAGAACAATTATCAAA  
AGCATAAGAATTGCAGCCACAGCATAGGGAAGAAAGAGGAGTTTTTAAAC  
CACAACAAAAGGGAGAAAGAAGAGAATTTTAACTTACATTTAATTCAAAA  
GTCTTCAGAGCAACCCGAAACCTCCTGGAAGTGGGGATTTCAGTCGAAG  
GGTCTCCTTAATAACACACCCGATGTATYTAAGTTGTTCCAAAATTTCCA  
TGTCCAAGTTGTTGCTTGTGATTGCTCTTGCAGTAACCCCTAYCAAAAY  
AGTCATACAGAGGTGAACAGTYATTTTGTGCTCCAATTAAATCAGCCCA  
GCAGACGTAAACAGGGCTTAAGTGGAGACTAAACCCAAAGGGCCCCATGA  
TGGGAGAGACTGGGAGGGGGAAACAGCAGCTAATGGCCATTTGCCTGCCC  
AAATCCACTATCTATTTACAATCCCAGGAGAATGCTGCTCACCAGTTAGA  
AGGACCAAGTTTCTCCCCACGCCCCCCCCACCCACACTCACCACCACCAC  
CCACACTAATCAGCTATTACACTATGTATGCCCTTGGACACACCAATTC  
AAGAAAAGTGAACCTATCTGAGAATCTCCACGGTTCACAAAAGGTGGA  
GGAGGGGTAGGAATACAAGGTCAAACCCCTGCCC

(2) INFORMATION FOR SEQ ID NO:38

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38

TCGCGAGGAGCGACCACGGCTTGAAGAGGGGTAGACGAGACCAGATGCTC  
CCCGGCGCCCCCTCATGCGGGTTGCGGTCTCTCTCCTCCACCTCCCTCTC  
AGCGGAGGAAGTTTCTGCAGATGAAGCGCAGGAAATACGGCTTCATCTAC  
AAGACGCATCTGTTTGGGCGGGCCACGGTGCGGGTGATGGGCGCGGATAA  
TGTGCGGCGCATCTTGCTGGGAGAGCACCAGTTGGTGTCTGGTGCCTGGC  
CCGCTCGGTGCGCACCATCCTGGGCGCTGGCTGCCTCTCCAACCTGCAC  
GATTCCTCGCACAGCAGCGAAAGAAGGTGAGGGTGAGCTGGCAACTCCT  
TGGCTGGCAGGGAGACCTCATCCTATGGCTTGGTTCAGGCAAAATAGAAT  
GCGGGGCGAGGGCTAGTCTATGTGGTGGGGACCAGGACCCTCTCTATCT  
GAGATCCACTTTAGCTTTTCTGCTAGCACGTGGGTAGTCTGGGGGGGA  
CTGAAATCTTTGAAGGGTACTCGGAAAGGCGAAGGGGGGGGGCTGAGG  
GAAAGTAGAGGATTGTAACACTCTCTGCTCCTGGGGGGTGCTCAGGTGAT  
TATGCAGGCCTTCAGCCGCGAGGCACTCCAGTGCTACGTGCCCCTGATCG  
CTGAGGAAGTCAGCAGTTGTCTGGAGCAGTGGCTAAGCTGCGGCGAGCGC  
GGCCTCCTGGTCTACCCGAGGTGAAGCGCCTCATGTTCCGCATCGCCAT  
GCGCATCCTGCTGGGCTGCGAGCCGGGTCCAGCGGGCGGGGAGGACG  
AGCAGCAGCTCGTGGAGGCTTTTCGAGGAGATGACCGCAATCTCTCTCT  
CTTCCCATTTACGTGCCCTTTAGCGGCTGTACCGGGTAAGGGCGGTTTG  
CGGAGTCGGAGTAGGGGAACGCAAGCTCGGGCATCCGCTCACCGCCACGC  
TCTCTCCGCGCTCAGGGCGTGAAGGCGCGGAACCTTATACACGCGCGCAT  
CGAGGAGAACATTCGCGCCAAGATCCGCCGGCTTCAGGCTACAGAGCCGG  
ATGGGGGTTGCAAGGACGCGCTGCAGCTCCTGATTGAGCACTCGTGGGAG  
AGGGGAGAGAGGCTGGATATGCAGGTGAGAAGCAATTTCAAAGGTGCCA  
AGGGCCGGGGAGTGCCCTCTGACTTTCAGACACACTTTCTGGGGTCTCCA  
AAGCCCTGTCAAGGCCCCAGCTACTTCCAAGTGGGCGGCGATGCTAGGTC  
TAGAGCTTTTCAACCTGTGGGTCTGACCCCTTCACGGAGCCAAACAACC  
CTTTCAGAGGGTTCGCTAAGAGCATCTGCATATCCGATATTTACATCAA  
GAAACATAACAGTACGATAAATTACCGTTATGAAGTAGCAACAAAGATAAT  
TTTATCGTTGGGGGTACCACACACGAGGAACCGTATTAAGGGTGCGCA  
TTGGTCTAGAGAGCTGTGGAAGGGGGTGGCTGAGCAATGGGGAAGATCCC  
AAAGTTCAAAGGGCAAGGCTCATCTACAAAGGTTAAAGCGGAAGAGCAGG  
ATTAAGGGAGTTTTGCGTTTTTGTGTTGTGGTCTTTGACTTCTATGAACA  
AAACGGATTTTACCCTTGAAGTCTTCCGTGCAATATTCTCAGGTCAGGTC  
TTTGTAACAGTGCTATAAACTGCACTCAGATCTGTATAAACTTCCGTTTT

GAAGTGCTTCAGGA



**WHAT IS CLAIMED IS:**

1. A purified protein having the ability to oxidize a retinoid, and having an amino acid sequence which is at least about 30% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or identified as SEQ ID NO:32, or a functionally equivalent homolog thereof.
2. A said protein of claim 1, in which the protein is at least about 35% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or identified as SEQ ID NO:32, or a functionally equivalent homolog thereof.
3. A said protein of claim 1, in which the protein is at least about 50% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or identified as SEQ ID NO:32, or a functionally equivalent homolog thereof.
4. A protein according to claim 1, or a conservatively substituted variant thereof.
5. The protein of claim 1, wherein the retinoid is a retinol and/or a retinoic acid.
6. The protein of claim 1, wherein the protein has the ability to oxidize the carbon at the 4-position of the  $\beta$ -ionone ring of the retinoid.
7. The protein of claim 6, wherein the retinoid is an all-*trans* retinoid.
8. A purified protein having the ability to hydroxylate retinoic acid at the 4 position of the  $\beta$ -ionone ring of retinoic acid, and having an amino acid sequence which is at least about 30% conserved in relation to the amino acid sequence identified as SEQ ID NO:2 or identified as SEQ ID NO:4 or identified as SEQ ID NO:32, or a functionally equivalent homolog thereof.
9. A purified protein having the ability to oxidize a retinoid at the carbon in the 4-position of the  $\beta$ -ionone ring of the retinoid.
10. The protein of claim 9 wherein the retinoid is an all-*trans* retinoid.
11. The protein of claim 10 wherein the retinoid is a retinoic acid.
12. The protein of claim 1, or a conservatively substituted variant thereof.
13. An isolated nucleic acid molecule encoding a protein of claim 1, or a nucleic acid strand capable of hybridizing with the nucleic acid molecule under stringent hybridization conditions.
14. An isolated nucleic acid molecule encoding a protein of claim 8.
15. An isolated nucleic acid molecule comprising a DNA molecule having the sequence identified as SEQ ID NO:3 or SEQ ID NO:5 or SEQ ID NO:31, or which varies from the sequence due to the degeneracy of the genetic code, or a nucleic acid strand capable of hybridizing with at least one said nucleic acid molecule under stringent hybridization conditions.
16. A said nucleic acid molecule of claim 15 wherein the sequence of the nucleic acid molecule corresponds to a part of a human genome or of a fish genome or of a mouse genome, or varies therefrom due to the degeneracy of the genetic code.
17. Isolated mRNA transcribed from DNA having a sequence which corresponds to a nucleic acid molecule according to claim 15.
18. A microbial cell containing and expressing heterologous DNA encoding a retinoid inducible protein.
19. A microbial cell according to claim 18 in which the protein is capable of oxidizing a retinoid.

20. A microbial cell containing and expressing heterologous DNA encoding a retinoid inducible protein having all-*trans* retinoic acid 4-hydroxylase activity.
21. The cell of claim 18 wherein the retinoid is retinoic acid.
22. A microbial cell containing and expressing heterologous DNA which is complementary a  
5 nucleic acid molecule of claim 13.
23. Isolated DNA having a sequence according to claim 15 in a recombinant cloning vector.
24. A stably transfected cell line which expresses a protein according to claim 1.
25. A culture of cells transformed with a recombinant DNA molecule having a nucleic acid sequence according to claim 15.
- 10 26. A host cell that has been engineered genetically to produce a protein according to claim 1, the cell having incorporated expressibly therein heterologous DNA encoding said protein.
27. The cell of claim 26 wherein production of the protein is inducible by exposing the cell to retinoic acid.
28. The cell of claim 26 wherein the cell is eukaryotic.
- 15 29. A process for producing the protein of claim 1 comprising:  
preparing a DNA fragment including a nucleotide sequence which encodes said protein;  
incorporating the DNA fragment into an expression vector to obtain a recombinant DNA  
molecule which includes the DNA fragment and is capable of undergoing replication;  
transforming a host cell with said recombinant DNA molecule to produce a transformant  
20 which can express said protein;  
culturing the transformant to produce said protein; and  
recovering said protein from resulting cultured mixture.
30. An antibody to a protein of claim 1.
31. An antibody of claim 30 wherein the antibody is a monoclonal antibody.
- 25 32. An antibody to a protein having a sequence included in the sequence identified as SEQ ID NO:4.
33. A protein of claim 1 for use in metabolizing retinoic acid in an organism or cell in need of such metabolizing.
34. A method for metabolizing retinoic acid in an organism or cell in need of retinoic acid  
30 metabolizing comprising administering a protein of claim 1.
35. A method for inhibiting retinoic acid hydroxylation in an organism in need of such inhibition, comprising administering to the organism an effective amount of an antisense nucleic acid or oligonucleotide substantially complementary to at least a portion of the sequence identified as SEQ ID NO:3 or SEQ ID NO:5 or SEQ ID NO:31.
- 35 36. The method of claim 35 wherein the portion is at least 5 bases in length.
37. The method of claim 35 wherein the organism is human.
38. The method of claim 35 wherein the organism is being treated for a disease selected from the group consisting of cancer, actinic keratosis, oral leukoplakia, a secondary tumor of the head and/or neck, a non-small cell lung carcinoma, a basal cell carcinoma, acute

promyelocytic leukemia, skin cancer, and a premalignancy associated actinic keratosis, acne, psoriasis and/or ichthyosis.

39. The method of claim 38 wherein the disease is acute promyelocytic leukemia.

40. A method for producing a desired protein, comprising:

- 5 providing a cell which can produce an endogenous protein in response to exposure to a retinoid;  
incorporating into DNA of the cell a DNA sequence encoding the desired protein at or near a site which is normally occupied by a DNA sequence encoding the endogenous protein; and
- 10 exposing the cell to the retinoid so as to induce production of the desired protein.

41. A kit for determining the presence of a protein according to claim 1 or having an amino acid sequence identified as SEQ ID NO:4, comprising an antibody to said protein linked to a reporter system, wherein the reporter system produces a detectable response when a predetermined amount of the protein and the antibody are bound together.

- 15 42. A kit for determining the presence of a first said nucleic acid molecule of claim 13, the kit comprising a second nucleic acid molecule capable of hybridizing with at least a portion of a the first nucleic acid molecule under stringent conditions, in which the second nucleic acid molecule is linked to a reporter system wherein the reporter system produces a detectable response when a predetermined amount of the first and second molecules are hybridized
- 20 together.

43. The kit of claim 42, wherein the nucleic acid molecule is at least about 5 bases in length.

44. An isolated DNA molecule having a nucleotide sequence selected from the group consisting of:

- (a) SEQ ID NO:33;
- 25 (b) SEQ ID NO:34;
- (c) SEQ ID NO:35; and
- (d) a fragment of (a), (b) or (c),

wherein said DNA molecule possesses promoter activity.

45. A DNA molecule of claim 44 wherein said DNA molecule includes the sequence

- 30 TGAAC(T)<sub>x</sub>TGAAC, wherein x has a value of up to 5.

46. A DNA molecule of claim 45 wherein x is 5.

47. A DNA molecule of claim 45 wherein said DNA molecule further includes the sequence TCTGASSAAGKTAAC downstream from the sequence TGAAC(T)<sub>x</sub>TGAAC.

48. A DNA molecule of claim 47 further including the sequence AATT between the sequence

- 35 TGAAC(T)<sub>x</sub>TGAAC and the sequence TCTGASSAAGKTAAC.

49. A DNA molecule of claim 48 wherein there are up to six nucleotides between the sequence TGAAC(T)<sub>x</sub>TGAAC and the sequence TCTGASSAAGKTAAC.

50. A DNA molecule of claim 45 further comprising the sequence CAATTAAAGA upstream of the sequence TGAAC(T)<sub>x</sub>TGAAC.

51. A DNA molecule of claim 44 wherein said DNA molecule includes the sequence CAATTAAAGATGAACTTTGGGTGAACTAATT.
52. A DNA molecule of claim 44, 45, 46, 47, 48, 49, 50 or 51 wherein the said DNA molecule includes the sequence TATAA.
- 53 A DNA molecule of claim 45, 46 or 50, wherein the said DNA molecule includes the sequence TATAA downstream of the sequence TGAAC(N)<sub>x</sub>TGAAC.
54. A DNA molecule of claim 47 or 48, wherein the said DNA molecule includes the sequence TATAA downstream of the sequence TCTGASSAAGKTAAC.
55. A DNA molecule of claim 45 wherein said DNA molecule includes the sequence TATAA located downstream of the sequence TGAAC(N)<sub>x</sub>TGAAC and spaced up to about 55 nucleotides therefrom.
56. A recombinant DNA comprising the isolated DNA molecule according to claim 44.
57. A recombinant DNA according to claim 51, further comprising one or more structural genes.
58. A recombinant DNA according to claim 56, wherein the one or more structural genes encodes cytochrome P450 protein(s).
59. An expression plasmid comprising the recombinant DNA according to claim 56.
60. An isolated cell containing the recombinant DNA according to claim 57.
61. An isolated cell according to claim 56 wherein said cell is eukaryotic.
62. A process for the production of recombinant protein(s), comprising culturing a cell according to claim 60 and recovering the protein(s) produced.
63. A process according to claim 62 wherein the protein(s) comprise a cytochrome P450.
64. A process according to claim 62 wherein the protein(s) comprise a fusion protein.
65. A method of screening drugs for their effect on activity of a retinoic acid inducible protein comprising exposing a purified said protein to a said drug and determining the effect on the activity.
66. The method of claim 65 wherein the activity is hydroxylation of a retinoic acid, particularly all-*trans* retinoic acid.
67. A method of screening drugs for their effect on activity of a protein of claim 1 comprising exposing a purified said protein to a said drug and determining the effect on the activity.
68. The method of claim 67 wherein said method includes ascertaining the effect a said drug has on the oxidation of retinoic acid.
69. The method of claim 68 wherein said retinoid is all-*trans* retinoic acid.
70. A method of screening drugs for their effect on expression of a gene wherein the gene is inducible by a retinoid, comprising exposing a recombinant DNA of claim 60 to a said drug and determining the effect on gene expression.
71. The method of claim 70 including exposing the recombinant DNA in the presence of a said retinoid.
72. The method of claim 70 wherein said structural gene is a reporter gene which does not metabolize retinoic acid.

73. The method of claim 70 wherein determining the effect on gene expression includes ascertaining the effect on transcription of the gene.

74. A method of screening a drug for its effect on the metabolism of a retinoid by a cytochrome P450 encoded by a nucleotide sequence identified according to claim 1 and incorporated into  
5 an expression system so as to be under control of a nucleotide sequence having native promoter activity for a said cytochrome P450, comprising:

exposing the system to the drug in the presence of the retinoid so as to determine the effect of the drug on metabolism of the retinoid.

75. The method of claim 74 wherein the retinoid is retinoic acid.

- 10 76. The method of claim 74 or claim 75 wherein the retinoic acid is all *trans*-retinoic acid.

77. A drug identified according to a method of claim 65, 66, 67, 68 or 69 as having the effect of modulating the activity of a said protein.

78. A drug identified according to a method of claim 70, 71, 72, 73, 74, or 75 as having the effect of modulating gene expression.

- 15 79. A drug identified according to a method of claim 76 as having the effect of modulating gene expression.

80. A method for inhibiting retinoic acid metabolism in an organism in need of such inhibition, comprising administering to the organism an effective amount of a drug of claim 77.

- 20 81. A method for inhibiting retinoic acid metabolism in an organism in need of such inhibition, comprising administering to the organism an effective amount of a drug of claim 78.

82. A method for inhibiting retinoic acid metabolism in an organism in need of such inhibition, comprising administering to the organism an effective amount of a drug of claim 79.

**ABSTRACT OF THE DISCLOSURE**

Amino acid sequences and corresponding nucleic acid sequence of retinoid metabolizing protein found in human, mouse and zebrafish are described, as well as methods of using same.

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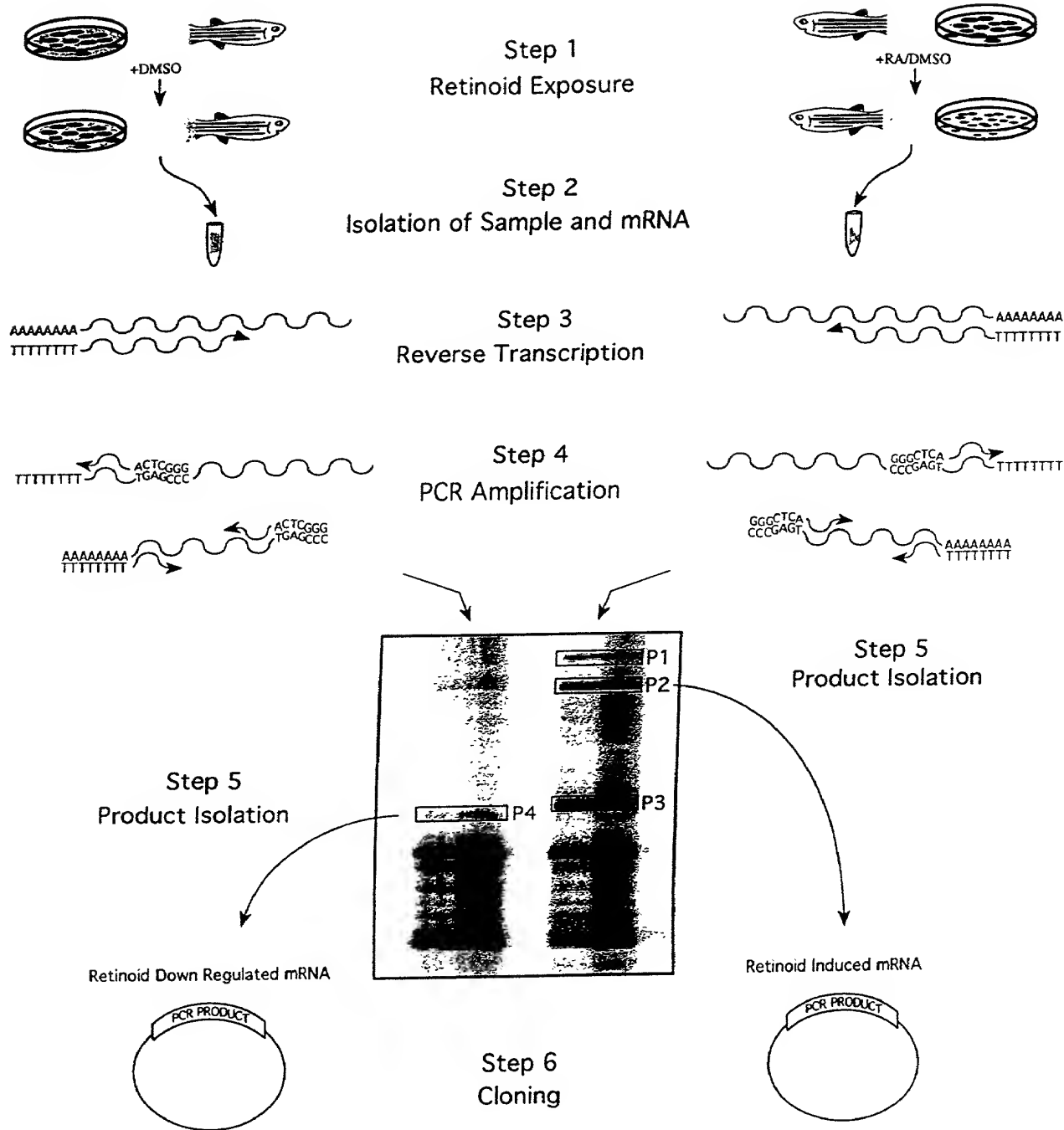
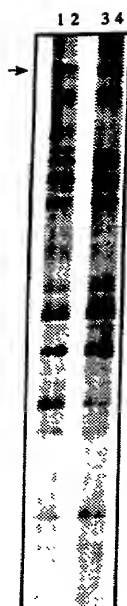


Fig. 1. Schematic representation of the steps involved in the isolation of retinoid-regulated genes using the differential display technique. The cloned products isolated in step 6 can then be used for sequencing, Northern blotting or screening of cDNA libraries. P1, P2 and P3 correspond to fragments from RA induced mRNAs. P4 represents a PCR product from an mRNA which is down-regulated.

FIG. 1

FIG. 2

A



C

MetGlyLeuTyrThrLeuMetValThrPheLeuCysThrIleValLeuProValLeuLeu 20  
PheLeuAlaAlaValLysLeuTrpGluMetLeuMetIleArgArgValAspProAsnCys 40  
ArgSerProLeuProProGlyThrMetGlyLeuProPheIleGlyGluThrLeuGlnLeu 60  
IleLeuGlnArgArgLysPheLeuArgMetLysArgGlnLysTyrGlyCysIleTyrLys 80  
ThrHisLeuPheGlyAsnProThrValArgValMetGlyAlaAspAsnValArgGlnIle 100  
LeuLeuGlyGluHisLysLeuValSerValGlnTrpProAlaSerValArgThrIleLeu 120  
GlySerAspThrLeuSerAsnValHisGlyValGlnHisLysAsnLysLysAlaIle 140  
MetArgAlaPheSerArgAspAlaLeuGluHisTyrIleProValIleGlnGlnGluVal 160  
LysSerAlaIleGlnGluTrpLeuGlnLysAspSerCysValLeuValTyrProGluMet 180  
LysLysLeuMetPheArgIleAlaMetArgIleLeuLeuGlyPheGluProGluGlnIle 200  
LysThrAspGluGlnGluLeuValGluAlaPheGluGluMetIleLysAsnLeuPheSer 220  
LeuProIleAspValProPheSerGlyLeuTyrArgGlyLeuArgAlaArgAsnPheIle 240  
HisSerLysIleGluGluAsnIleArgLysLysIleGlnAspAspAspAsnGluAsnGlu 260  
GlnLysTyrLysAspAlaLeuGlnLeuLeuIleGluAsnSerArgArgSerAspGluPro 280  
PheSerLeuGlnAlaMetLysGluAlaAlaThrGluLeuLeuPheGlyGlyHisGluThr 300  
ThrAlaSerThrAlaThrSerLeuValMetPheLeuGlyLeuAsnThrGluValValGln 320  
LysValArgGluGluValGlnGluLysValGluMetGlyMetTyrThrProGlyLysGly 340  
LeuSerMetGluLeuLeuAspGlnLeuLysTyrThrGlyCysValIleLysGluThrLeu 360  
ArgIleAsnProProValProGlyGlyPheArgValAlaLeuLysThrPheGluLeuAsn 380  
GlyTyrGlnIleProLysGlyTrpAsnValIleTyrSerIleCysAspThrHisAspVal 400  
AlaAspValPheProAsnLysGluGluPheGlnProGluArgPheMetSerLysGlyLeu 420  
GluAspGlySerArgPheAsnTyrIleProPheGlyGlyGlySerArgMetCysValGly 440  
LysGluPheAlaLysValLeuLeuLysIlePheLeuValGluLeuThrGlnHisCysAsn 460  
TrpIleLeuSerAsnGlyProProThrMetLysThrGlyProThrIleTyrProValAsp 480  
AsnLeuProThrLysPheThrSerTyrValArgAsn 492

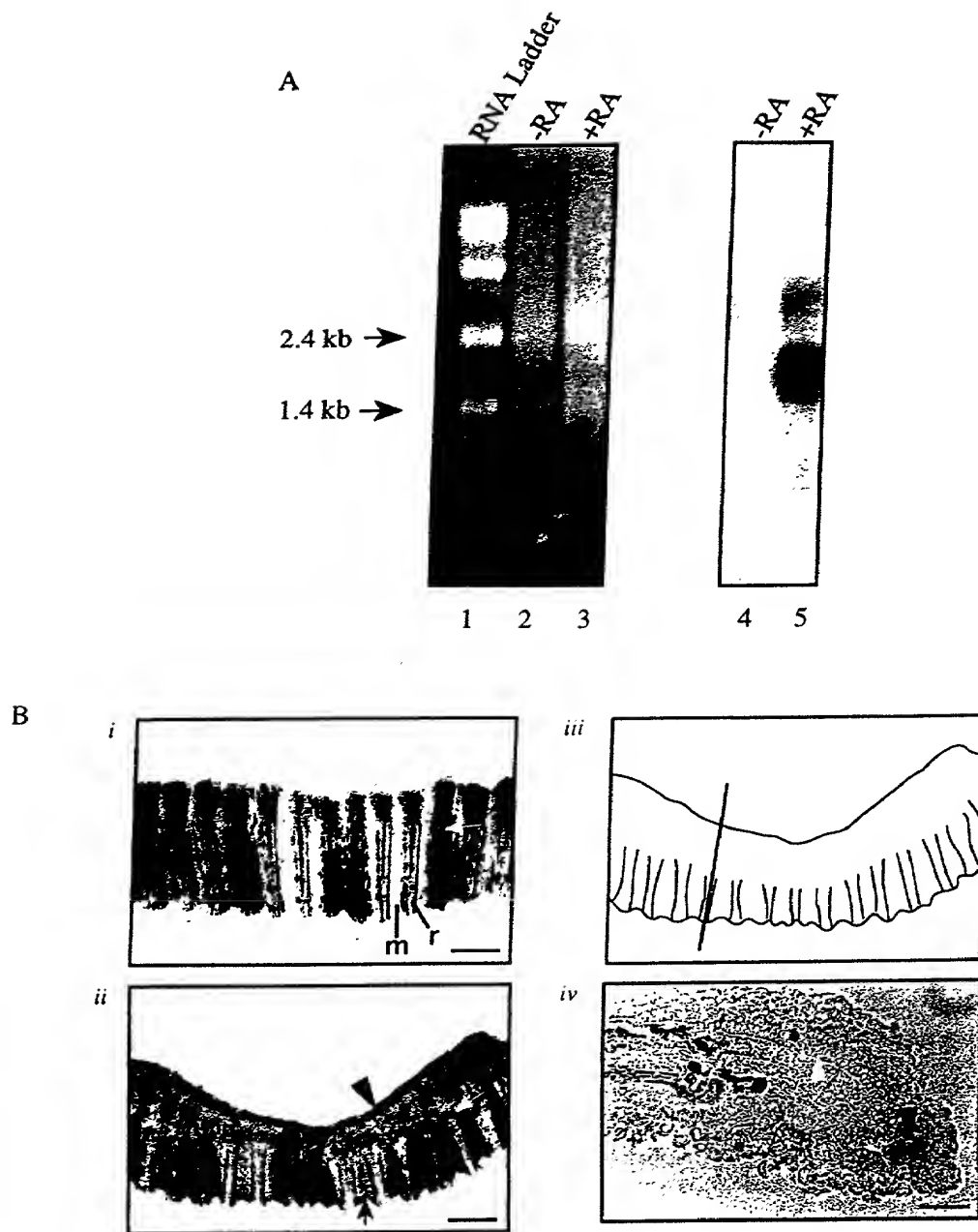
B

→  
TGCCAGTGGACAATCTCCCTACCAAATCACTAGTTATGTCAGAAATTA 50  
GCCTAAACCGAGCCCTTGTACATATGTTTTATTTTAGATGAAGTGA 100  
TOTATTGGATATTTCTAATTTGTTTATATAAAGCAGATGTGTATATAAG 150  
TCTATGCGAAGAAAGCGAAACGAGGCACTACTTTCTCATGGATCACTGT 200  
AATGCTACAGAGTGTCTGTGATGTATATTATAATGTAGTTGTGTATAT 250  
AGCTTTTGTACTGTATGCAACTTATTTAACTGCTCTTTATCTCATGGGT 300  
TTTATTATAAACAATGTTCTTACAAAAA 337  
←

D

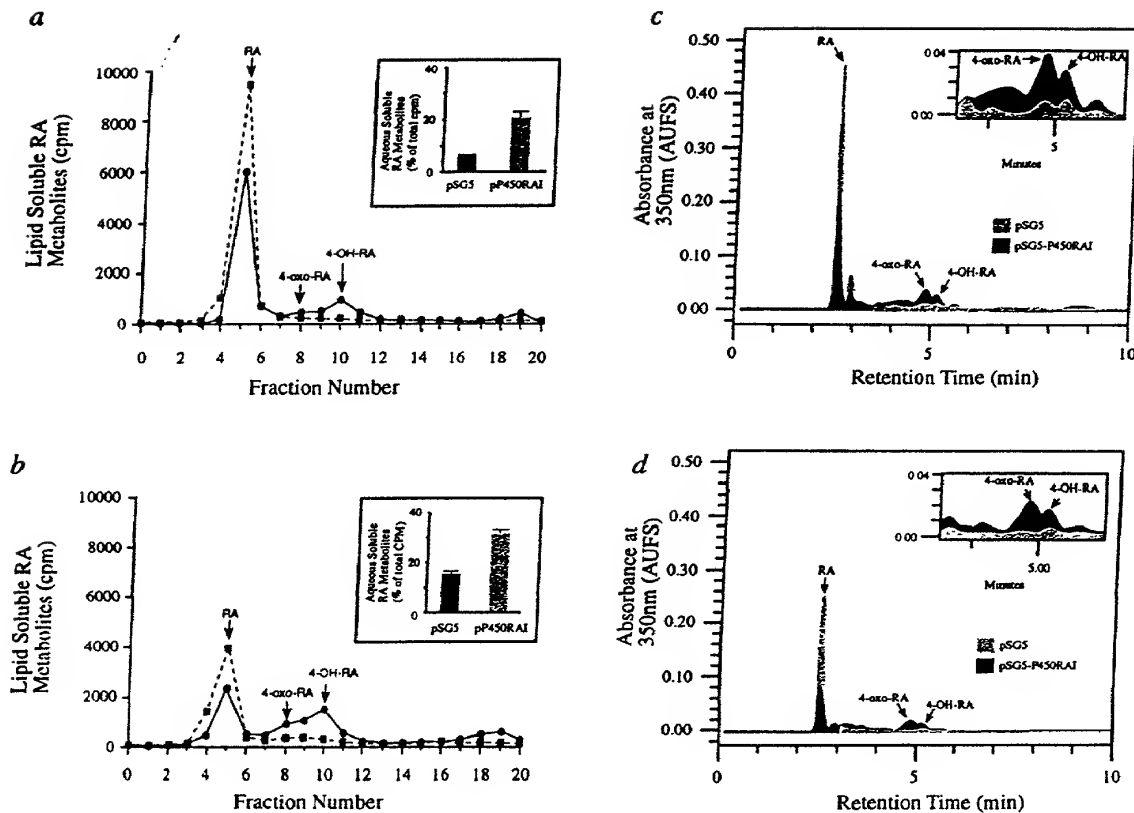
P450RAI	-8 -4 0 4 8	PFGGGSRMCVGKEFAKVLLK
ATCYP450		*****P*L*P*Y*L*R*A*S
RATCYP4A1		**S**A*N*I**Q**MSEM*
RABCYP4A5		**S**A*N*I**Q**MNE**
CYP4503A12		***T*P*N*I**MR**IMNM*
hCYP4A12		**S*****I**Q**MNE**





**FIG. 3**

FIG. 4



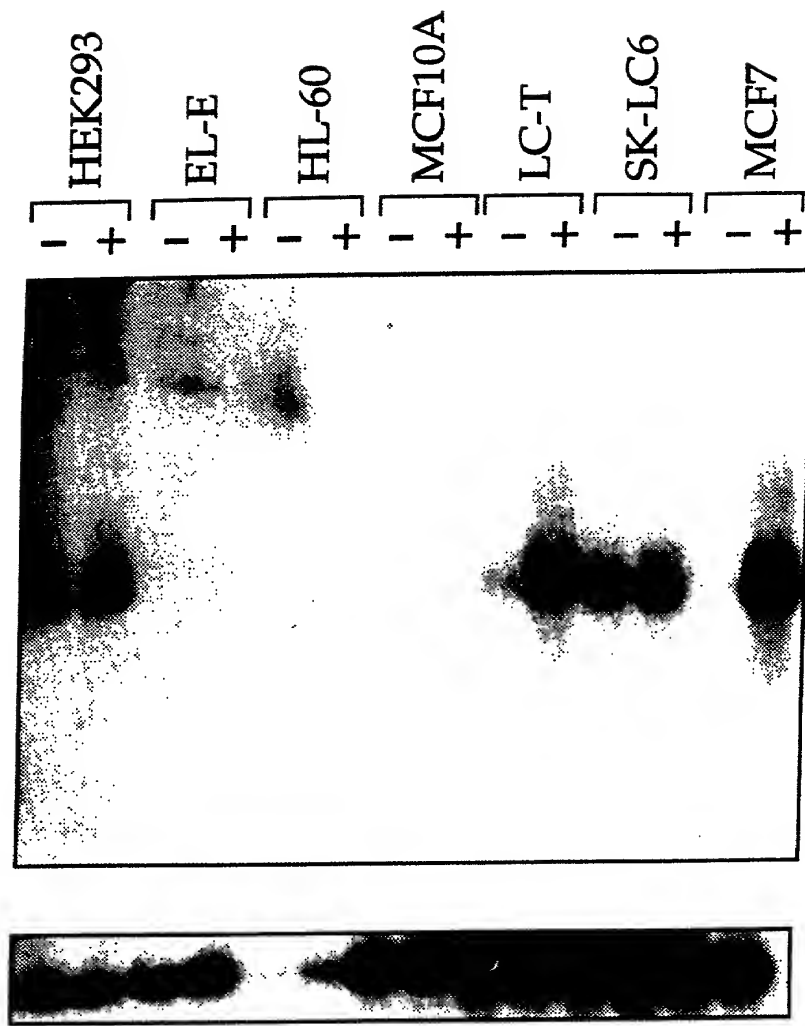


FIG. 5

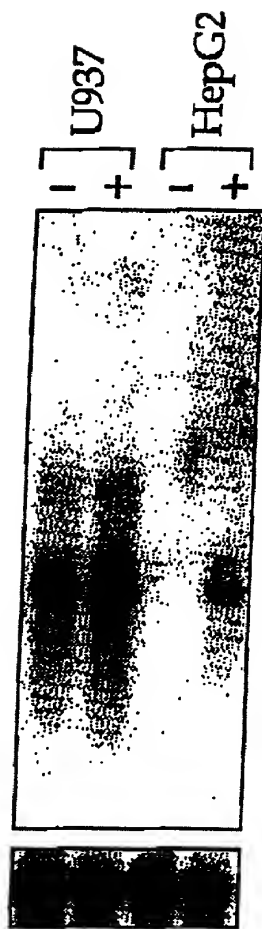


FIG. 6

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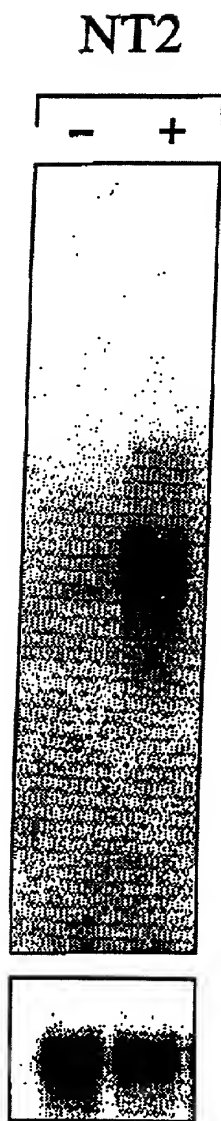


FIG. 7

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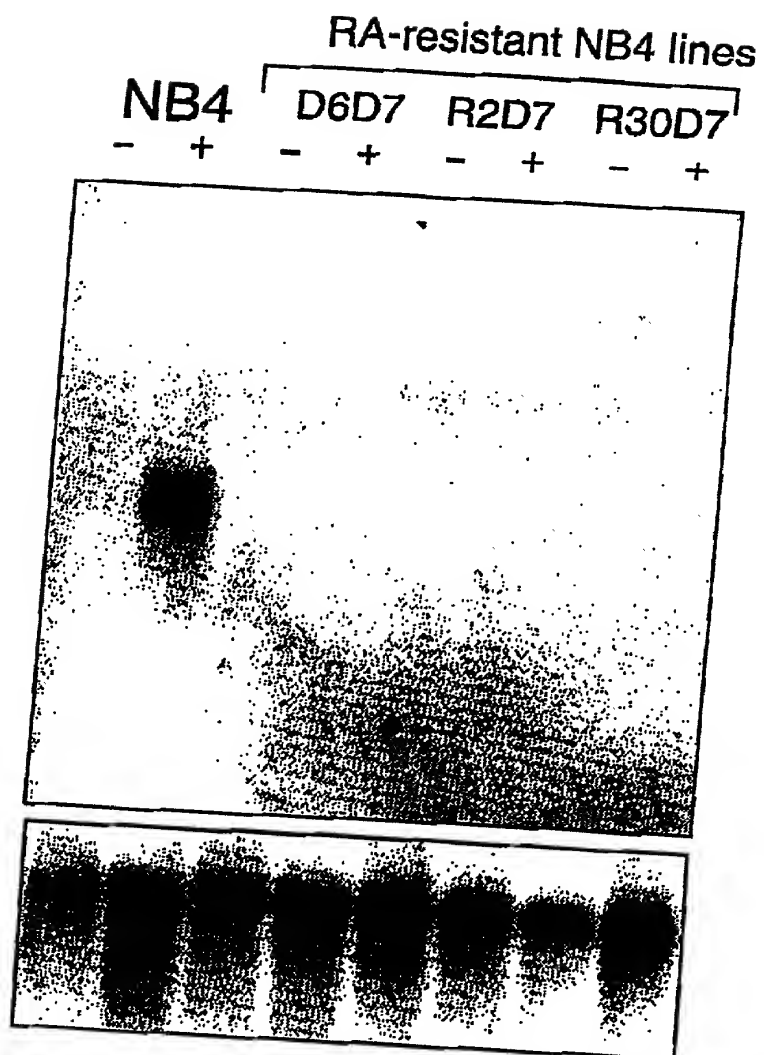


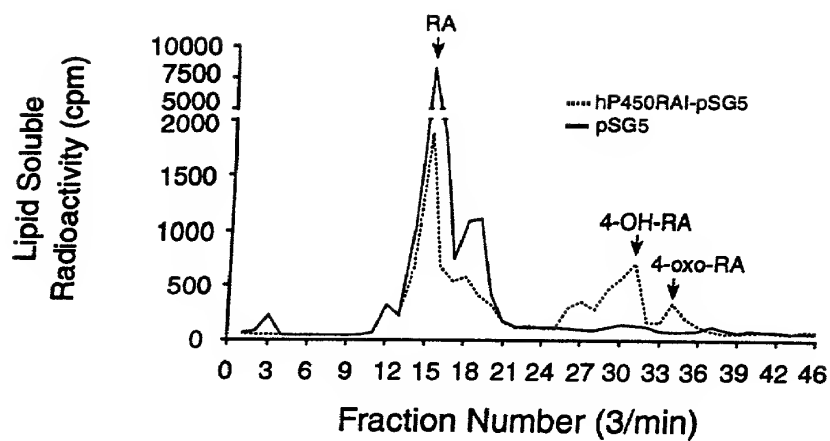
FIG. 8

FIG. 9

mP450RAI	MGLPALLASALCTFVLPLLLFLAALKLWDLYCVSSRDRSCALPLPPGTMGFPFFGETLQM	60
hP450RAI	.....I.....G.....	60
zP450RAI	...YT.MVTF...I...V.....V...EMLMIRRV.PN.RS.....L..I.....L	60
mP450RAI	VLQRRKFLQMKRRKYGFIYKTHLFGRTVVRVMGADNVRRILLGEHRLVSVHWPASVRTIL	120
hP450RAI	.....DD.....	120
zP450RAI	I.....R...Q...C.....N.....Q.....K...Q.....	120
mP450RAI	GAGCLSNLHDSSHKQRKKVIMQAFSREALQCYVLVIAEEVSSCLEQWLSCGERGLLVYPE	180
hP450RAI	.S.....R.....E...P..T...G.S.....	180
zP450RAI	.SDT...V.GVQ..NK..A..R....D..EH.IP..QQ..K.AIQE..Q-KDSCV....	179
mP450RAI	VKRLMFRIAMRILLGCEPGPAGGGEDEQQLVEAFEEMTRNLFSLPIDVPFSGLYRGVKAR	240
hP450RAI	.....QL..D.DS.....M...	240
zP450RAI	M.K.....F..EQI--KT...E.....IK.....LR...	237
mP450RAI	NLIHARIEENIRAKIRRLQATEPDGGCKDALQLLIEHSWERGERLDMQALKQSSTELLFG	300
hP450RAI	.....Q.....CG.R.S.AGQ.....	300
zP450RAI	.F..SK.....K..QDDDNENEQ-KY.....N.RRSD.PFSL..M.EAA.....	296
mP450RAI	GHETTASAATSLITYLGLYPHVLQKVREEIKSKGLLCKSNQDNKLDMETLEQLKYIGCVI	360
hP450RAI	.....L.....I.....	360
zP450RAI	.....T...VMP...NTE.V.....VQE.VEMGMYTPGKG.S..L.D...T...	356
mP450RAI	KETLRNLNPPVPGGFRVALKTFELNGYQIPKGWNVISICDTHDVADIFTNKEEFNPDRFI	420
hP450RAI	.....E.....S	420
zP450RAI	....I.....V.P....Q.E..M	416
mP450RAI	VPHPEDASRFSFIPFGGGLRSCVKGFEKILLKIFTVELARHCDWQLLNGPPTMKTSPTV	480
hP450RAI	A.....	480
zP450RAI	SKGL..G...NY.....S.M.....V....L...TQ..N.I.S.....G..I	476
mP450RAI	YPVDNLPARFTYFQGDI	497
hP450RAI	.....H.H.E.	497
zP450RAI	.....TK..SYVRN-	492

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*a*



*b*

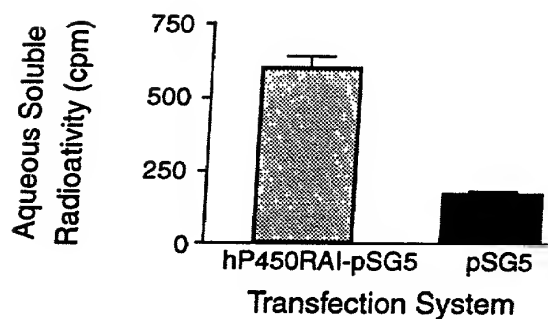
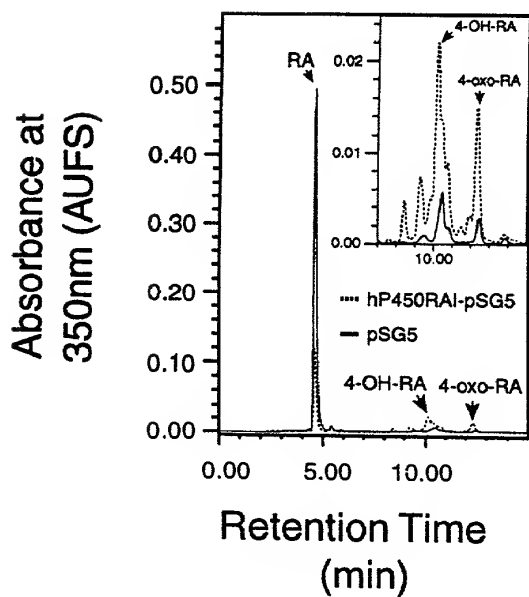


FIG. 10

*c*





4-oxo-RA Production in Transfected COS-1 Cells

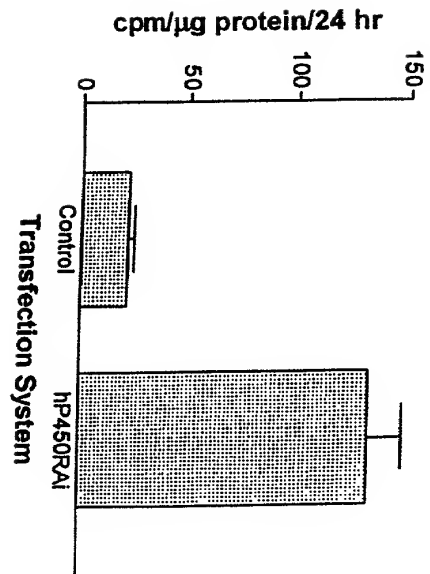


FIG. 11(a)

4-OH-RA Production in Transfected COS-1 cells

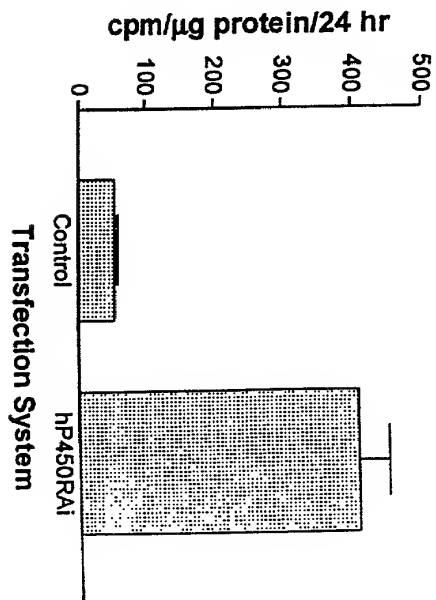


FIG. 11(b)

Formation of Aqueous Soluble Metabolites in Transfected COS-1 Cells

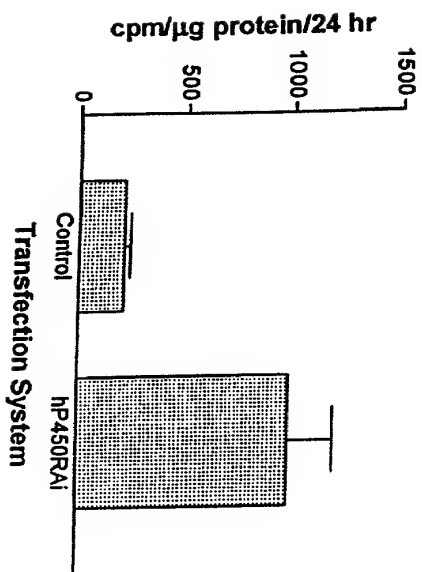


FIG. 11(c)

Unmetabolized RA in Transfected COS-1 Cells

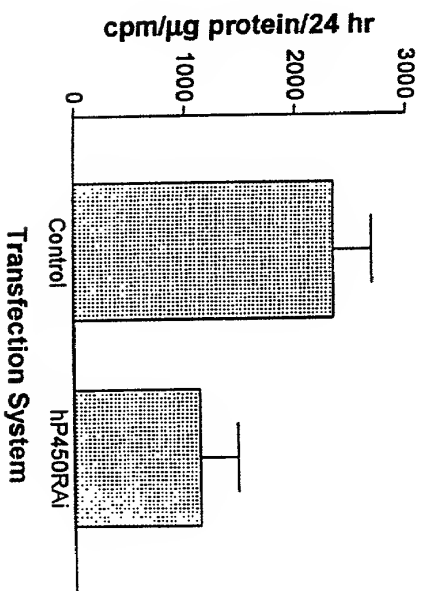
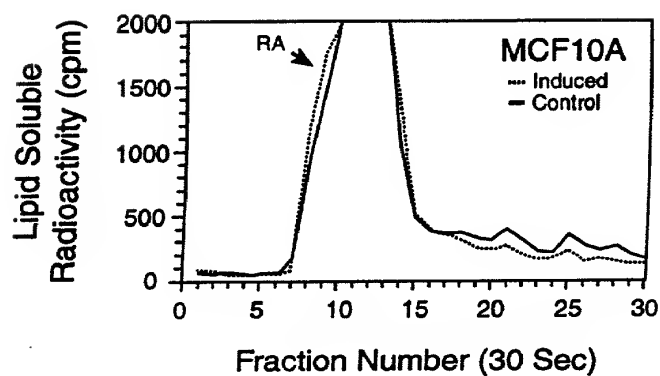


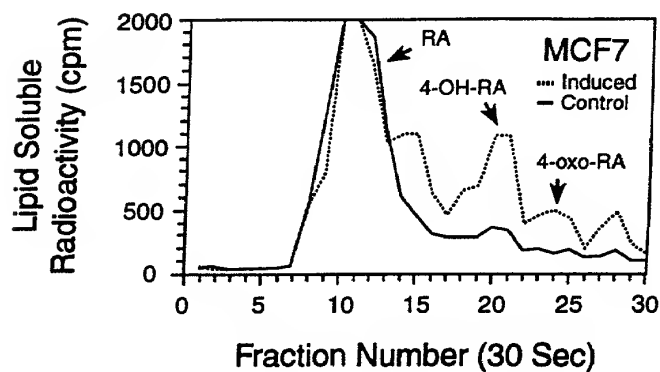
FIG. 11(d)

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*a*



*b*



*c*

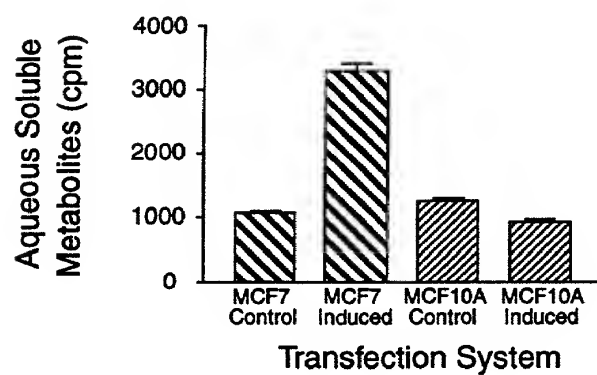
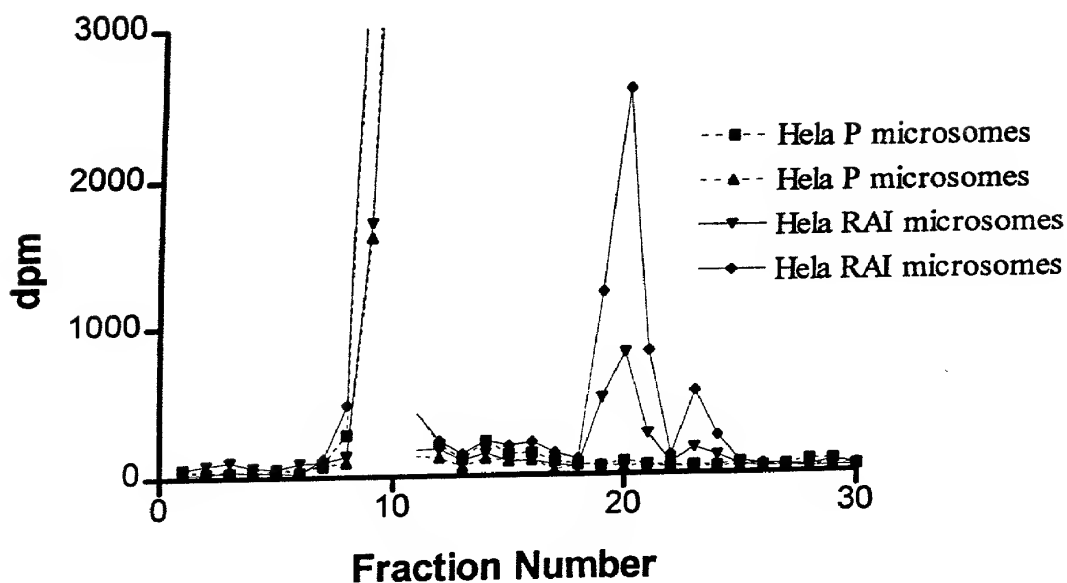


FIG. 12

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### Microsomal Preparations (90 minutes)



### Microsomal Preparations (90 minutes)

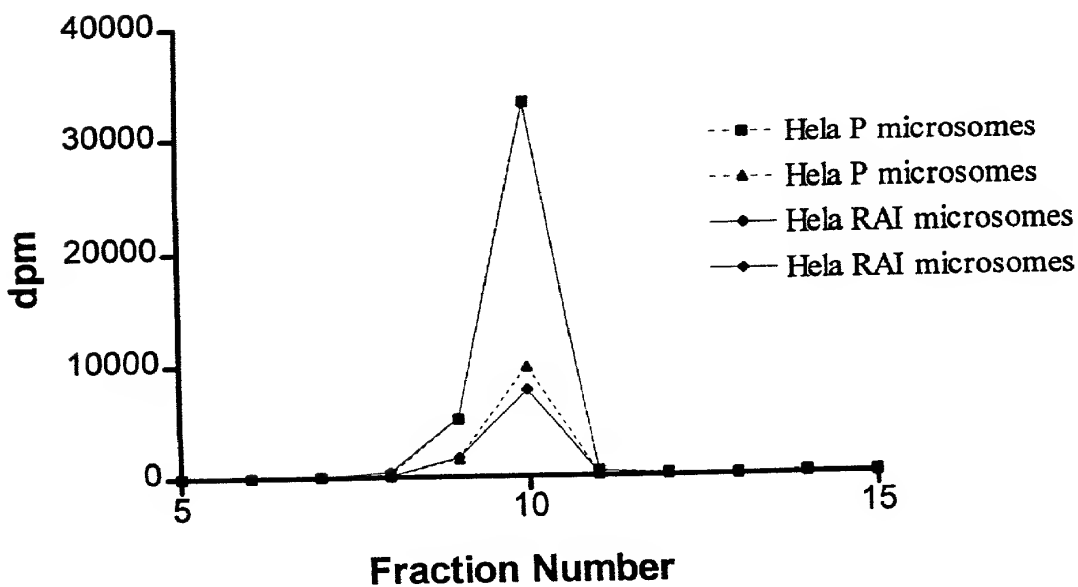
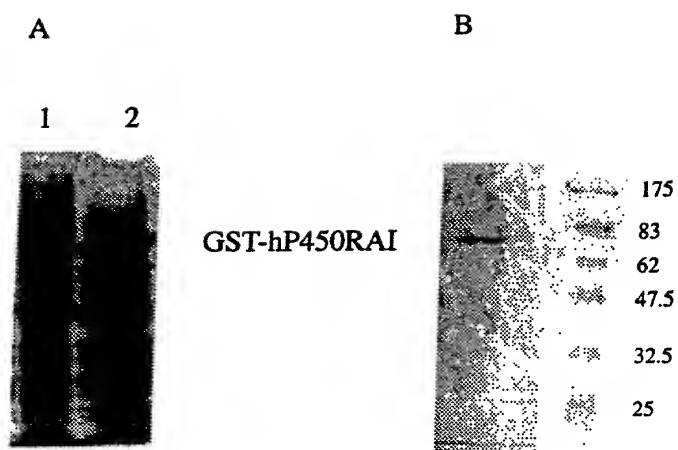


FIG. 13



**FIG. 14**

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hRAI promoter	-CG-----CA-C-----CC-----CAGGAGG
mRAI promoter	-CG-----CACC-----CC-----CAGGAGG
zRAI promoter	TCGGGGGAATTAACACCTTTTCAAAGTGAAATCTCAGGATTGTCTGCCTTCTACAGGAGG
hRAI promoter	-----CGCGCTCGGAGGGAA--GCCGCCAC--CGCCGCCCTCTGCCTCGGCG
mRAI promoter	-----CGCGCTCAGAGGGAA--GCCGCCAG--TG-CGCCGCTCTGCCTCGGCG
zRAI promoter	TGGTATTAAATGCGCCTATAACAAATGGTTGAGAGTTTGGAGCCGCTTCTGCCCTG---
hRAI promoter	CGGAACAAACGGTTAAAGATTTT---GGGCCASCCTCCGCGGGGGAGGAGCCAGGGG
mRAI promoter	CGGAACAAACGGTTAAAGATTTT---TGGGC-AGCGCCT-CGAGGGGGAGGAGCCAGGGG
zRAI promoter	-----TGGGC-----GGGGCGAG-----ATG---
hRAI promoter	CCCCAATCCCGCAATTAAAGATGAACCTTGGCTGAACCTAATT-GTCTGACCAAGGTAACG
mRAI promoter	CCCGA--TCCGCAATTAAAGATGAACCTTGGCTGAACCTAATTGTCTGACCAAGGTAACG
zRAI promoter	-----ACACCACAATTAAAGATGAACCTTGTGTGAACCTAATTATCTGAGGAAGTTAACA
hRAI promoter	TGGGCAGCAACCTGGGCCGCC---TATAAGCGGCAGCGCCGTGGGGTTGAAGCGCTG
mRAI promoter	TGGGCAGTAACCTGGGCCGCC---TATAAGAGGGCGCGCGGGGTTGCGAGCTAGG
zRAI promoter	GGAGGAGACCTGCGCGCAATGGATATATAAGGGCGCGCAGGCGAGGACGCCCTCAGTTTG
hRAI promoter	GCGGCGGCGGCAGGTGGCGCGGGAGGTCG-----CG
mRAI promoter	GCGGCGGCGGCAGGTGGCGCGGGAGGCTG-----AA
zRAI promoter	TGCGTAAAGACGCGTCTCCTCTCCAGAAGCTTGTTTTCGTTTGGCGATCAGTTGCGCG
hRAI promoter	GCGCGCCATGG
mRAI promoter	GCGTGCCATGG
zRAI promoter	CTTCAACATGG

FIG. 15

09653442, 092500

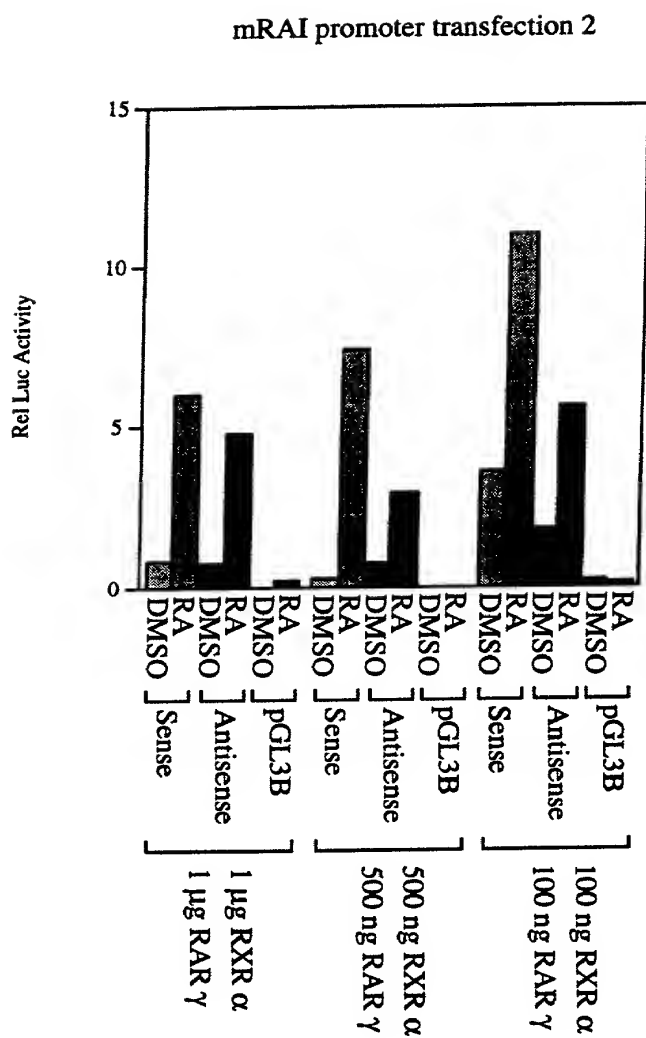
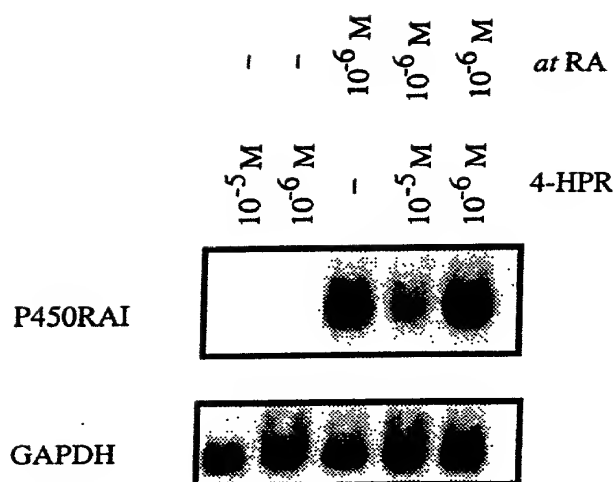


FIG. 16



Cells were treated for twelve hours with the indicated concentrations of *all-trans* retinoic acid (*at*RA) and 4-HPR. Total RNA was extracted using TRIzol, and, following electrophoresis, Northern blotting was performed as described. The nitrocellulose was probed with radiolabelled P450RAI or GAPDH.

FIG. 17

005250" 28489960

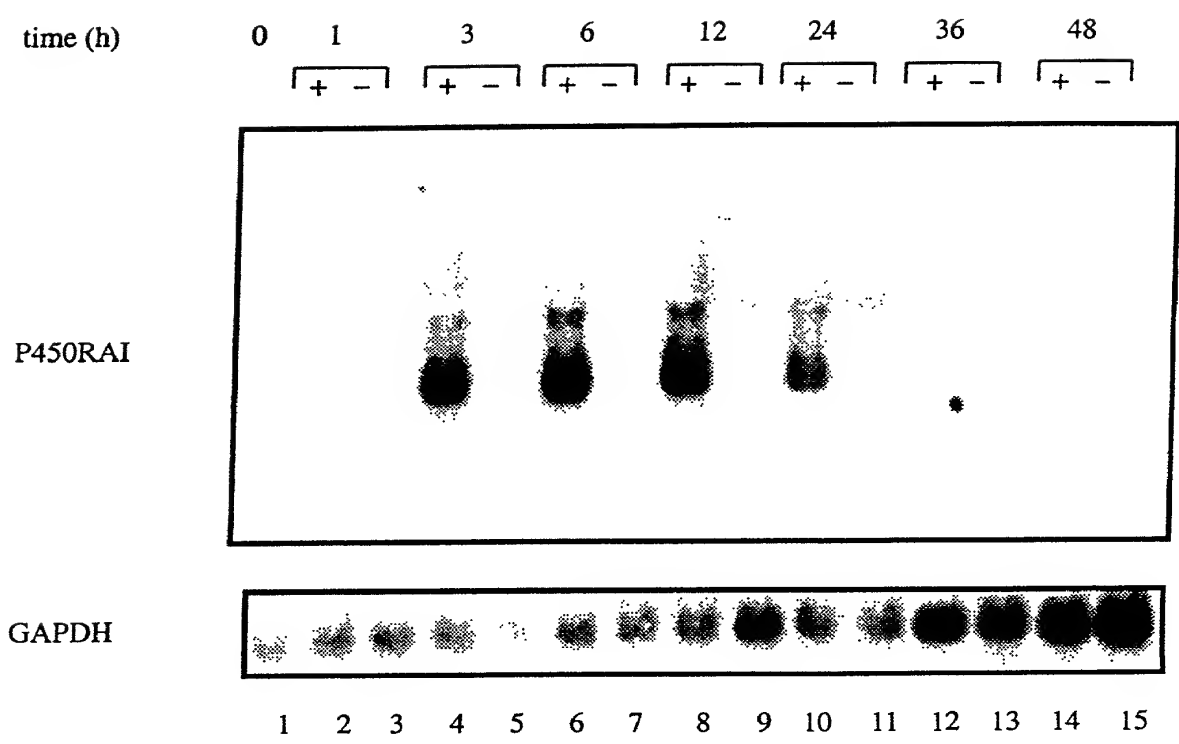


FIG. 18



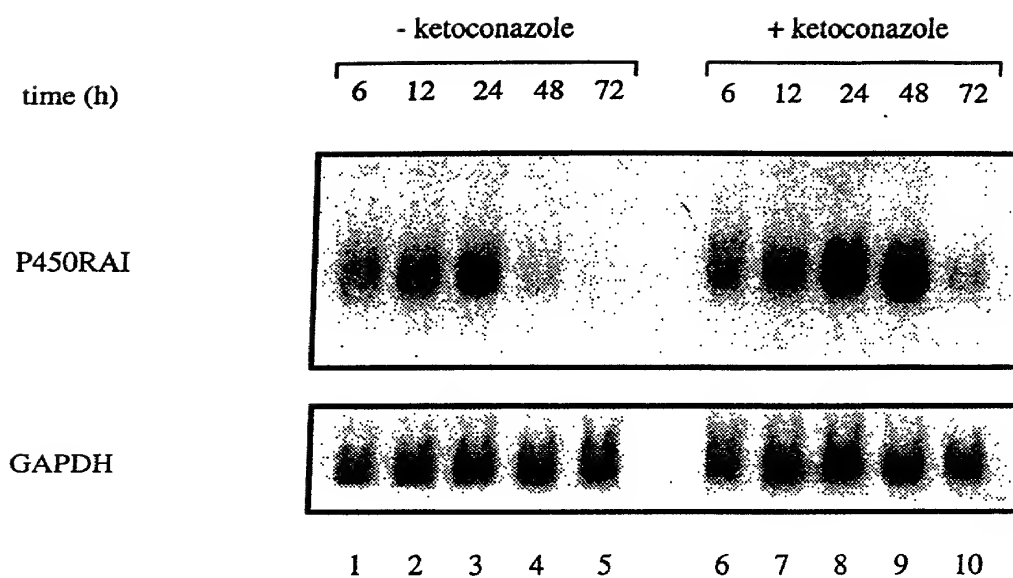


FIG. 19

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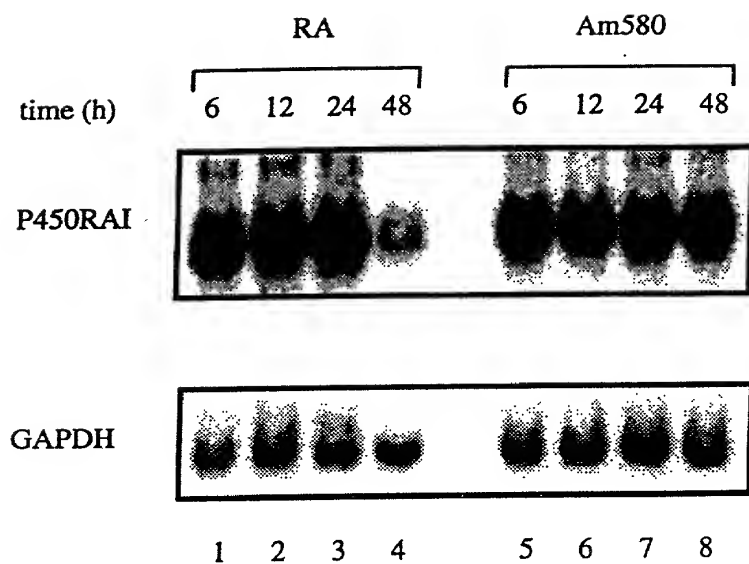


FIG. 20

## DECLARATION

This application is a Continuation-in-Part application of U.S. Serial No. 08/724,466, filed October 1, 1996, which is a Continuation-in-Part application of U.S. Serial No. 08/667,546, filed June 21, 1996.

As the below named inventors, we hereby declare that:

Our residences, post office addresses and citizenships are as stated below next to our names.

We believe that we are the original, first and joint inventors of the subject matter which is claimed and for which a patent is sought on the invention entitled:

### **Retinoid Metabolizing Protein**

as set forth in U.S. Patent Application Serial No. 08/882,164, filed June 25, 1997.

We hereby state that we have reviewed and understand the contents of the above-identified specification, including the claims.

We acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56, and in particular information which has occurred between the filing date of our earlier application(s) and the filing date of this application.

We hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

#### ***Prior Foreign Application(s)***

(Number)	(Country)	(Day/Month/Year Filed)	Priority Claimed [ ] [X] Yes No
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We hereby claim internal priority benefits under Title 35, United States Code, § 119(e) of any U.S. application(s) for patent listed below and have also identified below any U.S. application(s) for patent having a filing date before that of the application on which priority is claimed.

***Prior U.S. Application(s)***

08/667,546	United States	21 June 1996	Priority
(Number)	(Country)	(Day/Month/Year Filed)	Claimed
			[X] [ ]
			Yes No

08/724,466	United States	01 October 1996	Priority
(Number)	(Country)	(Day/Month/Year Filed)	Claimed
			[X] [ ]
			Yes No

**POWER OF ATTORNEY**

We hereby appoint Brian W. Gray (Reg. No. 30,017); Sheldon Burshtein (Reg. No. 32,851); John C. Hunt (Reg. No. 36,424); Alfred A. Macchione (Reg. No. 40,333); Kenneth Lloyd Bousfield (Reg. No. 40,460), care of Blake, Cassels & Graydon, Box 25, Commerce Court West, Toronto, Ontario, M5L 1A9, Canada, and Carol Miernicki Steeg (Reg. No. 39,539) care of PARTEQ Innovations, Queen's University, Kingston, Ontario, K7L 3N6, Canada, as our attorneys or agents to prosecute this application, to make alterations and amendments therein, to receive the patent and all correspondence relating to this application, and to transact all business in the United States Patent and Trademark Office connected therewith, and the said attorneys or agents are hereby given full power of substitution and revocation.

Please address all correspondence concerning this application to John C. Hunt, c/o Blake, Cassels & Graydon, Box 25, Commerce Court West, 28th Floor, Toronto, Ontario, M5L 1A9, Canada.

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful, false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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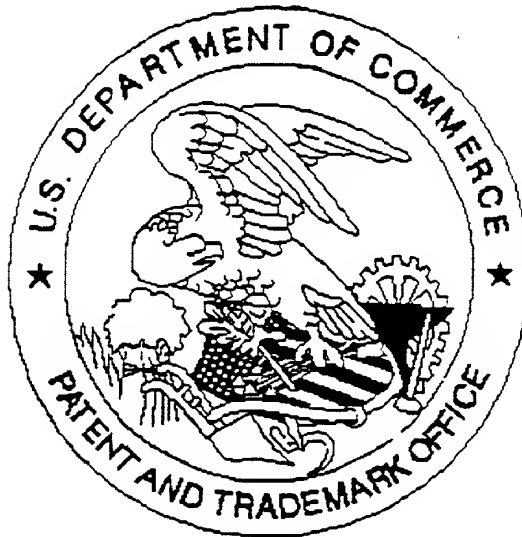
  
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